



Philippine COVID-19 Living Clinical Practice Guidelines

Institute of Clinical Epidemiology, National Institutes of Health, UP Manila

In cooperation with the Philippine Society for Microbiology and Infectious Diseases

Funded by the Department of Health

EVIDENCE SUMMARY

Among patients suspected to have COVID-19, how accurate are rapid antigen tests compared to RT-PCR for the diagnosis of COVID-19?

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RECOMMENDATION

We suggest the use of rapid antigen test for the diagnosis of symptomatic individuals suspected of COVID-19 as an alternative to RT-PCR if all the following conditions are met: *(Low certainty of evidence; Weak recommendation)*

- Individuals are in the early phase of illness (less than or equal to 7 days from onset of symptoms)
- Testing kits demonstrated sensitivity of more than or equal to 80% AND have very high specificity of more than or equal to 97%

We suggest against the use of rapid antigen test for screening purposes. *(Low certainty of evidence; Weak recommendation)*

We suggest against the use of saliva as specimen for rapid antigen test in patients suspected of COVID-19 infection. *(Low certainty of evidence; Weak recommendation)*

We suggest against the use of rapid antigen tests alone in asymptomatic patients suspected of COVID-19 infection. *(Low certainty of evidence; Weak recommendation)*

We suggest the use of rapid antigen tests for the diagnosis of individuals suspected of COVID-19 during the setting of an outbreak provided that all the following conditions are met: *(Very low certainty of evidence; Weak recommendation)*

- Individuals are in the early phase of illness (less than or equal to 7 days from onset of symptoms); AND
- Testing kits demonstrated sensitivity of more than or equal to 80% AND have very high specificity of more than or equal to 97%.

There is insufficient evidence to recommend for or against the use of repeat antigen testing for screening or diagnosis of COVID-19. *(Very low certainty of evidence)*

A negative rapid antigen test should be confirmed with an RT-PCR in settings or situations wherein COVID-19 is highly suspected (e.g., symptomatic or asymptomatic close contacts of probable or confirmed COVID-19 individuals).



Consensus Issues

The panel was unanimous against (1) the use of rapid antigen test for screening purposes, (2) the use of saliva as specimen for rapid antigen tests, and (3) the use of rapid antigen test alone in asymptomatic patients suspected of COVID-19 infection due to the observed lower sensitivity of these tests under such conditions. A unanimous decision on the insufficiency of evidence to recommend for or against the use of repeat antigen testing was also made.

Majority of the panelists agreed that the following conditions should be met when using rapid antigen tests:

- a. Individuals are in the early phase of illness, because antigen tests perform best during this period; and
- b. Testing kits have a sensitivity of more than or equal to 80% and specificity of more than or equal to 97%, because the quality of the test kit should be ensured.

One of eleven panelists raised a concern on the specified sensitivity and specificity of the testing kits, as these are based on the Health Technology Assessment Council (HTAC) of the local Department of Health (DOH).

A weak recommendation on the use of rapid antigen tests for diagnosing COVID-19 suspects during outbreaks was made based on nine observational studies with unclear patient selection, conduct of reference standard, and patient flow and timing. The risk of exposure was an important consideration for the panel, citing that it is not cost-effective to test everyone during an outbreak. However, the risk stratification of participants was not specified in any of the studies.

PREVIOUS RECOMMENDATION

We recommend the use of rapid antigen test in patients suspected of COVID-19 infection meeting all the following conditions: (*Moderate quality of evidence; Strong recommendation*)

Symptomatic AND

Early phase ≤ 7 days from onset of symptoms AND

Specific brands that demonstrated sensitivity $\geq 80\%$ and have very high specificity ($\geq 97-100\%$)

We recommend against the use of saliva as a specimen for rapid antigen test in patients suspected of COVID-19 infection. (*Moderate quality of evidence; Strong recommendation*)

We recommend against the use of rapid antigen tests alone in asymptomatic patients suspected of COVID-19 infection. (*Moderate to high quality of evidence; Strong recommendation*)

Previous Consensus Issues

There were only two studies that used saliva as a specimen for rapid antigen test, which produced a pooled sensitivity of 17% (95% CI 13-23%) and a pooled specificity of 99% (95% CI 99-100%). Given the current evidence on the very low sensitivity for saliva, qualifying which specimen is used for a rapid antigen test is necessary.



What's new in this version?

- There are 124 new observational studies added to the initial studies evaluated in the previous evidence summary.
- New evidence on the use of antigen tests in special populations (children, healthcare workers), in the setting of an outbreak, as well as repeated/serial antigen testing was included in this review.
- Subgroup analysis was applied on the effect of cycle threshold values and additional test brands.
- Evidence on costs and resource implications of antigen testing was added.
- The certainty of evidence was downgraded due to risk of bias issues and inconsistencies across evaluations resulting in a change of draft statements to “*suggest*” from “*recommend*”.

Key Findings

- A total of 164 observational studies assessed the diagnostic accuracy of rapid antigen tests (RAgTs) against reverse transcriptase - polymerase chain reaction (RT-PCR) as the reference standard. Studies included different test brands, specimen types and timing of collection, symptom status, cycle threshold (CT) values, and populations, namely inpatients, children, and healthcare workers among others.
- The overall sensitivity of RAgTs is moderate at 0.71 (95% CI 0.68-0.73) while specificity is excellent at 0.995 (95% CI 0.993-0.996). This was comparable to the data of the previous evidence summary with pooled sensitivity of 0.72 (95% CI 0.64-0.78) and specificity of 0.99 (95% CI: 0.99-1.0).
- On subgroup analysis, RAgT showed higher sensitivity when used in symptomatic individuals (Sn 0.74, 95% CI 0.71-0.78), when conducted during the early phase or first week of illness (Sn 0.79, 95% CI 0.75-0.82), in positive specimens with Ct value <25 (Sn 0.94, 95% CI 0.92-0.96), and in other Ct thresholds considered as “high” viral load (Sn 0.89, 95% CI 0.85-0.92). Pooled sensitivity of commonly used specimen types falls between 65% to 79%. FDA-approved RAgT brands have pooled sensitivities ranging from 0-90% with improved performance of commonly used RAgT brands when used in symptomatic individuals.
- In outbreak settings, RAgT use remained to have an excellent specificity (Sp 0.966, 95% CI 0.997 - 0.999) with a similar sensitivity (Sn 0.68, 95% CI 0.45-0.84) but with less precise estimates.
- The overall certainty of evidence was low because of serious risk of bias in all domains (high and unclear risk in patient selection, conduct of index test and reference standard, and flow and timing) and serious inconsistency. Despite performing pre-specified subgroup analyses, significant heterogeneity was still observed. In certain subgroup analyses, such as use of RAgT in outbreak settings and saliva specimens, certainty of evidence was further downgraded to very low due to imprecision attributed to wide interval estimates.

Introduction

Reverse-transcriptase polymerase chain reaction (RT-PCR) test remains to be the gold standard in the diagnosis of COVID-19. Despite excellent diagnostic accuracy, RT-PCR-based assays are not entirely practical for all testing scenarios due to its need for additional specialized equipment, specialized training of laboratory-based staff, and high cost. On the other hand, rapid antigen tests (RAgTs) detect the presence of specific viral antigens with a faster turnaround time which may be performed at the point of care, are simple to use, requiring shorter training, and are relatively less expensive compared to RT-PCR tests.[1] If sufficiently accurate, RAgT can facilitate



timely decisions concerning the need for isolation, monitoring, treatment and contact tracing activities.[2]

The previous review by Burog et al in March 2021 showed that the pooled sensitivity and specificity of 30 studies and 10 evaluation reports on RAgTs were 0.72 (95% CI 0.64-0.78; $I^2=95.77$) and 0.99 (95% CI 0.99-1.0; $I^2=93.16$) respectively.[3] Therefore, RAgT use was only recommended strictly for symptomatic individuals during the early phase of illness using brands with at least moderate sensitivity (≥ 0.80) and high specificity ($\geq 0.97 - 1.00$).

With the addition of more RAgTs available for use in COVID-19 screening and diagnosis, this update investigated the diagnostic accuracy of RAgTs and compared it with that of the previous review. This can influence changes in the recommendations guiding current clinical practice.

Review Methods

We searched MEDLINE for studies published until September 30, 2021 using subject headings combined with free text terms related to COVID-19 or SARS-CoV-2 and rapid antigen tests/testing, with no language limits or method filters. Appendix 2 shows the detailed search.

Preprint studies were identified using the COVID-19 Living Evidence Database (https://zika.ispm.unibe.ch/assets/data/pub/search_beta/) with "antigen" as the search term. This database is updated daily and includes preprints from medRxiv and bioRxiv, as well as published articles from EMBASE and PubMed. The Cochrane COVID-19 Study Register (covid-19.cochrane.org/) was also searched using "antigen" as a search term. Search for related links and journals was also carried out. Bibliography sections of the included studies were reviewed for relevant articles that might be missed by database search.

To supplement the initial search yield, available data on RAgT from FIND SARS-CoV-2 Diagnostic pipeline (<https://www.finddx.org/covid-19/dx-data/>) was accessed. Reported sensitivity and specificity estimates from the package inserts of RAgTs approved by the Philippine FDA were also retrieved but were not included in the main analysis. Relevant clinical trials were searched on clinicaltrials.gov and the WHO International Clinical Trials Registry Platform (ICTRP). Local publications such as health technology assessments on the use of RAgTs were also sought. Methodological qualities of the diagnostic studies were assessed by independent reviewers using the QUADAS-2 instrument.

Heterogeneity was determined by visual inspection of forest plot of study evaluations and summary receiver operating characteristic (SROC) plot. Because of anticipated heterogeneity across studies, pooled sensitivity and specificity estimates were derived by stratifying studies according to test brand, CT value used, presence of symptoms, timing of specimen collection, and special populations such as children and healthcare workers. Summary estimates for sensitivity and specificity with 95% confidence intervals were derived using a random-effects bivariate binomial model [4] fitted as a generalized linear mixed effect model using the *metandi* and *gllam* commands in Stata/MP 13.0.[5] When only fewer than four studies were available for pooling, summary estimates were computed externally through a web-based app (MetaDTA v2.01; https://crsu.shinyapps.io/dta_ma/).[6,7] Results were graphically displayed in SROC curves with the summary operating points and 95% confidence regions as well as coupled forest plots.



Sensitivity analysis was performed by removing studies with low methodologic quality or with risk of bias issues in certain QUADAS-2 domains, and subsequently assessing their impact on overall diagnostic accuracy estimates.

Out of 802 titles and abstracts screened, we retrieved 198 full-text articles relevant to the key question. Removal of duplicate copies (e.g. 40 studies included in the initial RAgT review which were already available) and articles with incomplete or no data yielded 124 new studies that were appraised and included for final analysis in this update of the evidence summary.

Results

Characteristics of included studies

In addition to the 40 studies evaluated in the previous evidence review, 124 new published articles were appraised and analyzed, yielding in a total of 164 included observational studies [8-171] involving 235,546 samples. Using RT-PCR as reference standard, 25 different RAgT brands approved by the Philippine Food and Drug Administration (FDA) were evaluated. Included studies looked into screening and diagnosis of symptomatic and asymptomatic patients, employees, healthcare workers and students in both community and hospital settings. Appendix 3 shows a summary of the characteristics of included studies.

Methodological quality of included studies

The overall methodological quality of the included studies was rated as low. In < 50% of included studies, high and unclear risk of bias were seen in all domains particularly in the patient selection and conduct of both index test and reference standard. Issues on applicability, on the other hand, were documented in < 25% of studies. Certainty of evidence was downgraded to very low when only studies on outbreak settings were pooled due to the presence of imprecision. Appendix 4 shows a detailed assessment of the risk of bias of included studies.

Diagnostic accuracy of RAgT

A. Overall diagnostic accuracy

Across 164 studies, the pooled sensitivity of RAgTs was found to be moderate at 0.71 (95% CI 0.68-0.73). Pooled specificity was excellent at 0.995 (95% CI 0.994-0.996). Visual examination of forest plots and SROC plot indicated highly heterogeneous sensitivity estimates across studies. Figure 1 shows the SROC plot.

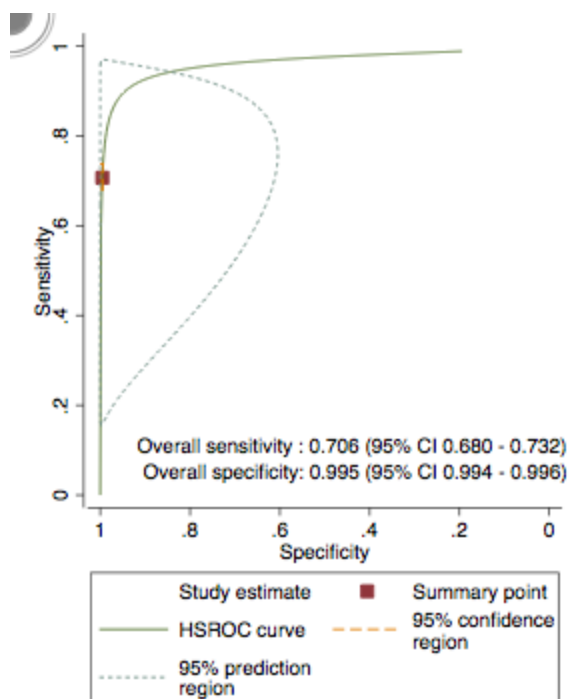


Figure 1. Summary receiver operating characteristic (SROC) plot showing the summary sensitivity and specificity point (165 studies / 272 evaluations). The 95% prediction region reflects heterogeneity in test accuracy across studies.

B. Subgroup Analysis

Table 1 shows the sensitivity of self-administered RAgTs per subgroup.



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Table 1. Subgroup Analysis for Sensitivity of RAgTs

| Variable | Current Review | | Previous Review |
|---|--------------------------------------|--------------------------|-------------------------|
| | Studies/ Evaluations (Samples) | Sensitivity (95% CI) | Sensitivity (95% CI) |
| OVERALL | 272 (235,794) | 0.71 (0.68, 0.73) | 0.72 (0.64, 0.78) |
| Special Populations | | | |
| Children | 11 (5,101) | 0.79 (0.70, 0.86) | Not evaluated |
| Healthcare workers | 5 (1,554) | 0.82 (0.64, 0.92) | Not evaluated |
| Presence of symptoms | | | |
| Symptomatic | 79 (45,523) | 0.74 (0.71, 0.78) | 0.78 (0.69, 0.86) |
| Asymptomatic | 55 (72,858) | 0.56 (0.51, 0.62) | 0.51 (0.39, 0.63) |
| Timing of testing in relation to symptoms | | | |
| Early | 54 (28,591) | 0.79 (0.75, 0.82) | 0.71 (0.44, 0.89) |
| Mixed | 44 (69,501) | 0.70 (0.65, 0.74) | 0.81 (0.76, 0.85) |
| Late | 23 (4,307) | 0.47 (0.39, 0.55) | 0.65 (0.57, 0.71) |
| Test brand and presence of symptoms | | | |
| Panbio™ Ag-RDT (Abbott) | 39 (29,764) | 0.72 (0.67, 0.77) | 0.76 (0.61, 0.86) |
| •Symptomatic | 17 (6,551) | 0.77 (0.72, 0.82) | Not evaluated |
| •Asymptomatic | 11 (2,642) | 0.52 (0.45, 0.59) | Not evaluated |
| Standard Q COVID-19 Ag Test (SD Biosensor) | 30 (24,102) | 0.72 (0.66, 0.78) | 0.77 (0.58, 0.94) |
| •Symptomatic | 12 (4,439) | 0.75 (0.68, 0.81) | Not evaluated |
| •Asymptomatic | 7 (2,903) | 0.65 (0.59, 0.70) | Not evaluated |
| Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | 16 (12,351) | 0.71 (0.61, 0.80) | 0.66 (0.55, 0.75) |
| •Symptomatic | 3 (452) | 0.83 (0.62, 0.94) | Not evaluated |
| •Asymptomatic | 6 (6,070) | 0.80 (0.57, 0.93) | Not evaluated |
| BinaxNOW SARS-CoV-2 (Abbott) | 13 (39,177) | 0.63 (0.50, 0.74) | 0.90 (0.66, 0.93) |
| •Symptomatic | 8 (3,425) | 0.81 (0.73, 0.87) | Not evaluated |



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| ● <i>Asymptomatic</i> | 5 (6,771) | 0.61 (0.52, 0.69) | Not evaluated |
| Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | 8 (7,343) | 0.75 (0.67, 0.81) | 0.70 (0.53, 0.85) |
| ● <i>Symptomatic</i> | 8 (3,798) | 0.78 (0.71, 0.84) | Not evaluated |
| ● <i>Asymptomatic</i> | 2 (2,564) | 0.58 (0.48, 0.67) | Not evaluated |
| Standard F Covid19 Ag FIA (SD Biosensor) | 7 (4,396) | 0.72 (0.41, 0.91) | 0.64 (0.40, 0.85) |
| ● <i>Symptomatic</i> | 3 (667) | 0.75 (0.43, 0.92) | Not evaluated |
| ● <i>Asymptomatic</i> | 1 (2,340) | 0.65 (0.54, 0.75) | Not evaluated |
| Biocredit COVID-19 Ag test (RapiGEN) | 7 (827) | 0.54 (0.39, 0.69) | 0.55 (0.38, 0.71) |
| ● <i>Symptomatic</i> | 2 (132) | 0.50 (0.27, 0.73) | Not evaluated |
| ● <i>Asymptomatic</i> | 1 (27) | 0.33 (0.20, 0.50) | Not evaluated |
| COVID-19 Rapid Antigen Test (BD Veritor) | 5 (5,183) | 0.68 (0.54, 0.79) | 0.76 (0.60, 0.89) |
| ● <i>Symptomatic</i> | 3 (821) | 0.83 (0.72, 0.91) | Not evaluated |
| ● <i>Asymptomatic</i> | 1 (2,317) | 0.59 (0.51, 0.66) | Not evaluated |
| Coronavirus Ag Rapid Test Cassette (Healgen Scientific) | 3 (715) | 0.84 (0.77, 0.90) | 0.77 (0.67, 0.85) |
| ● <i>Symptomatic</i> | 1 (332) | 0.79 (0.72, 0.86) | Not evaluated |
| VITROS Immunodiagnostic Products SARS-CoV-2 Antigen test (Ortho Clinical Diagnostics) | 2 (336) | 0.78 (0.72, 0.84) | 0.80 (0.74, 0.90) |
| NADAL COVID-19 Rapid Antigen Test (Nal Von Minden) | 2 (931) | 0.74 (0.45, 0.91) | Not evaluated |
| AFIAS COVID-19 Ag (AFC) (Menarini) | 2 (1,049) | 0.42 (0.38, 0.47) | Not evaluated |
| SARS-CoV-2 Antigen Rapid Test Kit (Lepu Medical) | 1 (286) | 0.46 (0.36, 0.56) | Not evaluated |
| CareStart COVID-19 Antigen Test (Access Bio) | 1 (286) | 0.46 (0.34, 0.45) | Not evaluated |
| Encode SARS-CoV-2 Antigen Rapid Test Device (Zhuhai Encode Medical Engineering) | 1 (200) | 0.74 (0.64, 0.82) | Not evaluated |
| Elecsys SARS-CoV-2 Antigen assay (Roche) | 1 (3,143) | 0.60 (0.55, 0.65) | Not evaluated |
| SIENNA COVID-19 Antigen Rapid Test Cassette | 1 (150) | 0.90 (0.82, 0.95) | Not evaluated |
| GenBody COVAG025 Rapid Antigen Test | 1 (130) | 0.90 (0.73, 0.98) | Not evaluated |



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| Humasis COVID-Ag Test COVID-19 Antigen (Humasis) | 1 (21) | 0.90 (0.70, 0.99) | Not evaluated |
| NowCheck COVID-19 Ag Test (Bionote) | 1 (1,326) | 0.56 (0.21, 0.86) | 0.89 (0.81, 0.95) |
| StrongStep Rapid Antigen Test (Liming) | 1 (19) | 0 (0, 0.34) | Not evaluated |
| Huaketai SARS-CoV-2 Rapid Antigen Test (Savant) | 1 (109) | 0.17 (0.09, 0.27) | 0.17 (0.09, 0.27) |
| SARS-CoV-2 Antigen Rapid Test Kit (JOYSBIO) | 1 (225) | 0.58 (0.47, 0.68) | Not evaluated |
| ichroma™ COVID-19 Ag (Boditech) | 1 (966) | 0.41 (0.37, 0.46) | Not evaluated |
| CT value | | | |
| Low (<25) | 40 (6,906) | 0.94 (0.92, 0.96) | Not evaluated |
| High (>25) | 35 (5,076) | 0.39 (0.38, 0.42) | Not evaluated |
| Other Ct thresholds for 'higher' viral load | 27 (14,398) | 0.89 (0.85, 0.92) | Not evaluated |
| Other Ct thresholds for 'lower' viral load | 18 (1,996) | 0.29 (0.20-0.41) | Not evaluated |
| Specimen type | | | |
| Exhaled breath | 1 (105) | 0.92 (0.64, 1.0) | Not evaluated |
| Nasopharyngeal and Saliva | 1 (343) | 0.91 (0.76, 0.98) | Not evaluated |
| Nasal (anterior nares and mid-turbinate) | 24 (51,399) | 0.79 (0.74, 0.83) | 0.84 (0.66, 0.93) |
| Nasopharyngeal | 99 (111,446) | 0.71 (0.68, 0.75) | 0.72 (0.65, 0.78) |
| Nasopharyngeal and Oropharyngeal | 35 (38,387) | 0.65 (0.59, 0.71) | 0.65 (0.47, 0.79) |
| Oropharyngeal | 3 (8,568) | 0.59 (0.42, 0.73) | Not evaluated |
| Saliva | 7 (6,148) | 0.57 (0.22, 0.86) | 0.17 (0.13, 0.23) |
| Nasal and Oropharyngeal | 1 (18,457) | 0.49 (0.47, 0.51) | Not evaluated |
| Sputum | 1 (45) | 0.11 (0.04, 0.24) | Not evaluated |
| In Outbreak Settings | | | |
| Overall | 9 (6805) | 0.68 (0.45 - 0.84) | Not evaluated |
| • Symptomatic | 3 (398) | 0.14 - 0.89 | Not evaluated |
| • Asymptomatic | 3 (1142) | 0.22 - 0.92 | Not evaluated |
| Methodological quality | | | |
| Studies with low risk of bias | 83 (73,469) | 0.75 (0.71, 0.79) | Not evaluated |



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|-----------------------------------|-------------|--------------------------|---------------|
| Studies with high risk of bias | 96 (88 252) | 0.64 (0.58, 0.69) | Not evaluated |
| Studies with unclear risk of bias | 93 (74 073) | 0.73 (0.68, 0.77) | Not evaluated |

By special population

The evidence summary update incorporated analysis of RAgT use in special populations. The pooled sensitivity of RAgT use in children was 0.79 (95% CI 0.70-0.86; n=5,101; 11 studies) while pooled sensitivity was 0.82 (95% CI 0.64-0.92; n=1,554; 5 studies) when used among healthcare workers. Though RAgT use in both populations showed high performance, more trials may be needed to substantiate recommendations for these specific populations.

By presence of symptoms

The use of RAgT in symptomatic individuals showed higher pooled sensitivity (Sn=0.74, 95% CI 0.71-0.78; n=45,523; 79 studies) compared to that of asymptomatic individuals (Sn=0.56, 95% CI 0.51-0.62; n=72 858; 55 studies). This is consistent with previous estimates showing higher sensitivity of RAgT in symptomatic individuals (Sn 0.78, 95% CI 0.69-0.86) compared to asymptomatic individuals (Sn 0.51, 95% CI 0.39-0.63).

By time of testing in relation to symptom onset

When RAgT was used either during the early (0-7 days) or late phase (>7 days) from symptom onset, the pooled sensitivity was 0.70 (95% CI 0.65-0.74; n=69,501; 44 studies). The pooled sensitivity increased to 0.79 (95% CI 0.75-0.82; n=28,591; 54 studies) when RAgT was used in the early phase of illness and decreased to 0.47 (95% CI 0.39-0.55; n=4,307; 23 studies) when RAgT was used during the late phase of the disease.

By test brand

Moderate performance was demonstrated by the five most commonly used RAgT brands, namely Abbott Panbio (Sn 0.72, 95% CI 0.65-0.77; n=29,764; 39 studies), SD biosensor Standard Q COVID-19 Ag (Sn 0.72, 95% CI 0.66-0.78; n=24,102; 30 studies), Roche SARS-CoV-2 Rapid Antigen Test (Sn 0.71, 95% CI 0.61-0.80; n=12,351; 16 studies), Abbott BinaxNOW CoVID-19 Antigen Card (Sn 0.63, 95% CI 0.50-0.74; n=39 177; 13 studies) and Quidel Sofia SARS Antigen Fluorescent Immunoassay (Sn 0.75, 95% CI 0.67-0.81, n=7,343; 8 studies).

Subgroup analysis showed improved diagnostic performance of the above RAgT brands when used with symptomatic individuals as pooled sensitivity increased in the range of 0.75 (95% CI 0.68-0.81; SD biosensor Standard Q COVID-19 Ag) to 0.83 (95% CI 0.62-0.94; Roche SARS-CoV-2 Rapid Antigen Test).

Though highest sensitivity was documented in the RAgT brands SIENNA COVID-19 Antigen Rapid Test (Sn 0.90, 95% CI 0.82-0.95; n=150; 1 study), GenBody COVAG025 Rapid Antigen Test (Sn 0.90, 95% CI 0.73- 0.98; n=130; 1 study) and Humasis COVID-Ag Test (Sn 0.90, 95% CI 0.70- 0.99; n=21; 1 study), their use in a limited number of samples warrant more trials to further validate their clinical utility.

By cycle threshold (Ct) value of RT-PCR

RAgTs demonstrated excellent performance in positive RT-PCR specimens having Ct value ≤25 with pooled sensitivity at 0.94 (95% CI 0.92-0.96; n=6,906; 40 studies). Studies that used Ct cutoff values other than 25 still showed better performance when samples had higher viral loads (i.e., lower Ct values) with a sensitivity of 0.89 (95% CI 0.85-0.92; n=14,398; 27 studies) versus 0.29 (95% CI 0.20-0.41; n=1996; 18 studies) when used in those with “low” viral load.



By specimen type

RAgT using exhaled breath samples showed the highest sensitivity (Sn 0.92, 95% CI 0.64-1.00; n=105; 1 study). This was followed by samples from combined nasopharyngeal and saliva (Sn 0.91, 95% CI 0.76-0.98; n=343; 1 study). However, the use of these specimens was limited to single studies with imprecise estimates.

The more commonly used specimen types, namely nasal, nasopharyngeal, combined nasopharyngeal and oropharyngeal, have moderate sensitivities. RAgTs had a pooled sensitivity of 0.79 (95% CI 0.74-0.82; n=51,399; 24 studies) when used in nasal specimens, 0.71 (95% CI 0.67-0.74; n=111,446; 102 studies) when used in nasopharyngeal specimens, and 0.65 (95% CI 0.59-0.71; n=38,387; 35 studies) when used in combined nasopharyngeal and oropharyngeal specimens.

Sputum as a specimen type had the lowest sensitivity (Sn 0.11, 95% CI 0.04-0.24; n=45; 1 study). Seven studies using saliva as the specimen type demonstrated poor sensitivity at 0.57 (95% CI 0.22-0.86; n=6,148). This result is consistent with the findings of the previous review.

By serial/repeat testing

Two observational studies with unclear risk of bias provided data on repeat testing. In the study of Shah et al., repeat antigen testing of anterior nasal specimen using Abbott BinaxNOW CoVID-19 Antigen Card in a community setting taken within 30 minutes from the initial evaluation showed an improved sensitivity from 0.77 (95% CI 0.72-0.82) to 0.81 (95% CI 0.77-0.86). This study tested a total of 2110 patients, 1190 (56%) of which were symptomatic. [148]

Using the same specimen type and RAgT brand, McKay and colleagues employed 3 rounds of serial testing over a 13-day period (every 4-5 days) among nursing home residents and staff using RT-PCR as reference standard. Majority of the subjects were asymptomatic (451/532 or 84.7%). Sensitivity was inconsistent across different test rounds: 1st round at 0.74 (95% CI 0.59-0.86), 2nd round at 0.63 (95% CI 0.47-0.78), and 3rd round at 0.67 (95% CI 0.41-0.88) respectively. On the other hand, specificity remained high throughout the study with a range of 0.97 to 1.0.[106]

During outbreak settings

The authors identified nine observational studies [23, 24, 27, 47, 93, 106, 112, 130, 159] that involved settings or scenarios wherein rapid antigen testing was compared to RT-PCR testing in the context of an outbreak. Six out of the nine studies consisted of a mixed population of asymptomatic and symptomatic. In studies during outbreaks and surges, rapid antigen tests had a pooled sensitivity of 0.68 (95% CI 0.45-0.84) which was similar to the overall pooled sensitivity of included studies but with less precise estimates. Pooled specificity remained excellent at 0.996 (95% CI 0.997-0.999). Certainty of evidence of the pooled studies on outbreak settings was downgraded to very low due to the presence of imprecision.

I. In Symptomatic Individuals

Three studies [23, 106, 112] provided specific data on symptomatic subjects. The three studies tested subjects within the early and late phases of illness with RT-PCR as the reference standard. A summary sensitivity estimate was not calculated due to the small number of studies and the variability in the test brands and specimens used. Sensitivity ranged from 0.14 to 0.89. When the



study with the lowest sensitivity estimate and high risk of bias [112] was excluded from analysis, sensitivity ranged from 0.76 to 0.89.

In the study by Bianco et al., 231 individuals reported at least one COVID-19 symptom and were tested using LumiraDx™ SARS-CoV-2 Antigen Test with RT-PCR as the reference standard. Sensitivity and specificity were high at 0.89 (95% CI 0.84-0.93) and 0.88 (95% CI 0.73-0.97) respectively.

Among residents and staff of a nursing home in the study by McKay et al., 79 paired specimens were evaluated using the BinaxNOW COVID-19 Ag Card, which showed moderate sensitivity of 0.76 (95% CI 0.58-0.89) but excellent specificity of 1.00 (95% CI 0.92-1.0)

Nagura-Ikeda reported the lowest sensitivity estimate of 0.14 (95% CI 0.07-0.27) and specificity of 1.00 (95% CI 0.8-1.0) from collected saliva specimens of 88 individuals using the Espline SARS-CoV2 (Fujirebio) Rapid Antigen Test.

II. In Asymptomatic Individuals

Three studies [23, 106, 112] provided data on asymptomatic subjects. A summary sensitivity estimate was not calculated due to the small number of studies and the variability in the test brands and specimens used. Sensitivity ranged from 0.22 to 0.92. When the study with the lowest sensitivity estimate [112] and high risk of bias was excluded from analysis, sensitivity ranged from 0.65 to 0.92.

The lowest sensitivity estimate of 0.22 (95% CI 0.03-0.6) was obtained from the study by Nagura-Ikeda et al. [112] which used saliva specimens and had high risk of bias. The remaining two observational studies used nasal specimens and showed higher sensitivities but had unclear risk of bias. In the study by Bianco et. al, 676 subjects underwent rapid antigen testing using LumiraDx™ SARS-CoV-2 Antigen Test with RT PCR as the reference standard. Sensitivity and specificity were high at 0.92 (95% CI 0.85-0.97) and 0.92 (95% CI 0.90-0.94) respectively. A total of 451 paired specimens from asymptomatic residents and staff of a nursing home were evaluated in a study by McKay et al. The BinaxNOW COVID-19 Ag Card was used for testing which showed poor sensitivity of 0.65 (95% CI 0.53-0.76) but excellent specificity of 0.98 (95% CI 0.96-0.99).

C. Sensitivity Analysis

RAgT showed an improved overall sensitivity when only studies with low risk of bias (high methodological quality) were included in the analysis (pooled Sn 0.75, 95% CI 0.71-0.79; n=73,469). This estimate appears to be comparable to that of studies with unclear risk of bias (Sn 0.73, 95% CI 0.68-0.77; n=74 073). In contrast, studies with high risk of bias in any of the four QUADAS-2 domains (patient selection, index test, reference standard, flow interval) produced significantly lower sensitivity estimates (pooled Sn 0.64, 95% CI 0.58-0.69; n=88,252).

Ongoing Studies on RAgT

As of November 2021, there were 24 ongoing trials [172-195] on the use of RAgTs for COVID-19 registered at ClinicalTrials.gov. Most of the trials compare the accuracy of RAgTs to RT PCR. Specimen type and brands used across the trials are varied. There are trials exploring the potential of RAgTs for screening and surveillance in various settings (i.e. workplace, airports, mass gatherings, community).



Other Considerations

With the increase in RAgT use, some studies have explored the economic advantage brought by testing. A cost-benefit study by Diel and Nienhaus in Germany have shown that the implementation of the Sofia SARS Antigen FIA test saved on average about 210 euros as compared to clinical-judgment only. Furthermore, with a high prevalence rate of 15.6% in the same study population, point-of-care COVID-19 antigen testing reduced average costs of hospitalized patients by 213 euros per tested patient.[196]

Another study has explored the economic effects of home-based RAgT in the United States of America. Despite the imperfections of home testing for COVID-19, the epidemic modeling done has shown that home-based RAgT can reduce transmission of infection and mortality at a justifiable cost.[197] No local economic evaluation study has been performed on RAgTs to date.

As there are numerous COVID-19 tests available, authorized organizations namely Philippine Food and Drug Administration (FDA) and the Research Institute for Tropical Medicine (RITM) were tasked to evaluate commercially manufactured COVID-19 tests to ensure the quality of the tests in the local setting. [198] The Philippine FDA-approved brands are constantly being updated because of this evaluation. Among the brands included in this review, Panbio™ Ag-RDT (Abbott), Sofia SARS Antigen Fluorescent Immunoassay (Quidel), and NowCheck COVID-19 Ag Test (Bionote) have passed the performance evaluation as of August 2, 2021.[199] In an advisory dated September 13, 2021, caution was advised on the use of COVID-19 Rapid Antigen Test (BD Veritor), NADAL COVID-19 Rapid Antigen Test (Nal Von Minden), and SARS-CoV-2 Antigen Rapid Test Kit (JOYSBIO) as these brands have failed the performance evaluation.[200] Standard Q COVID-19 Ag Test (SD Biosensor) was listed under the tests being recalled for the low performance in the evaluation and for failure to explain the non-conformance of the product to the specifications declared last August 2, 2021.[201] The company distributing ichroma COVID-19 Ag voluntarily surrendered their authorization to market the product as of June 18, 2021.[202]

The Department of Health (DOH) issued a memorandum dated September 1, 2021 which strictly placed a price cap of Php 960 for RAgT in all testing and clinical laboratories.[203] Unit cost of the different brands of RAgT kit in the market ranges from Php 250 to Php 1,600. Table 2 lists the available unit cost of RAgT kits locally. Automated RAgTs are much more costly than the manually interpreted RAgT.

Table 2. Unit Price of RAgT Kits

| Test Brand | Unit Cost |
|---|--------------------|
| Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | Php 1 600 (32 USD) |
| VITROS Immunodiagnostic Products SARS-CoV-2 Antigen test (Ortho Clinical Diagnostics) | Php 1 067 (21 USD) |
| COVID-19 Rapid Antigen Test (BD Veritor) | Php 907 (18 USD) |
| SIENNA COVID-19 Antigen Rapid Test Cassette | Php 750 (15 USD) |
| Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | Php 705 (14 USD) |
| GenBody COVAG025 Rapid Antigen Test | Php 503 (10 USD) |
| CareStart COVID-19 Antigen Test (Access Bio) | Php 605 (12 USD) |



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| | |
|---|------------------|
| Standard Q COVID-19 Ag Test (SD Biosensor) | Php 550 |
| Panbio™ Ag-RDT (Abbott) 1000-4975 tests (40-199 boxes) | Php 520 (13 USD) |
| NADAL COVID-19 Rapid Antigen Test (Nal Von Minden) | Php 349 (€ 6) |
| BinaxNOW SARS-CoV-2 (Abbott) | Php 250 (5 USD) |
| Coronavirus Ag Rapid Test Cassette (Healgen Scientific) | Php 680 (£ 10) |

Recommendations from Other Groups

Table 3 summarizes the recommendations from different agencies, countries, and organizations regarding the use of rapid antigen tests.



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Table 3. Summary of Recommendations from Other Groups

| Agency | Recommendation | Date |
|---|---|--------------|
| Department of Health Health Technology Assessment Council [204] | <p>The HTAC does not recommend the use of rapid antigen tests for indiscriminate use in mass screening, for return-to-work clearance and for COVID-19 diagnosis in individuals with low index of suspicion (i.e., asymptomatic and no history of exposure).</p> <p>Rapid antigen tests are currently recommended by HTAC only for very specific purposes:</p> <ul style="list-style-type: none"> • For targeted screening and diagnosis of suspected and probable cases of COVID-19 (i.e., with a high index of suspicion), meeting the clinical and/or epidemiologic criteria in the hospital or community settings. • For testing of patients in the hospital setting, where the turnaround time is critical, to guide patient cohort management to minimize transmission of COVID-19 among healthcare workers and other patients. (Hospitals are high-risk settings among healthcare workers and patients.) Otherwise, use RT-PCR in case of elective procedures; • For targeted screening and diagnosis of suspect and probable cases of COVID-19 (as defined above) in presumptive outbreaks where the result of the RT-PCR test of one suspect has not yet been released and in settings where RT-PCR is not immediately available or when delayed release of result or prolonged turnaround time is expected (i.e., more than 48 hours). • For local border screening at points of entry for individuals travelling from areas with a high daily positivity rate averaged over a seven-day period (i.e., >10%) or as reported by the DOH-Epidemiology Bureau based on its periodic updates of prevalence rate/positivity rate; and, • For international border screening at points of entry, always assume a high prevalence/positivity rate. A periodic update every month of prevalence rate/positivity rate per country is also suggested. It is recommended that the RT-PCR or RAgT test be used for screening of all incoming individuals in accordance with existing protocol and testing guidelines. Facility- or home-based quarantine shall also be implemented together with RT-PCR or Rapid Antigen testing. | Apr 30, 2021 |
| Infectious Disease Society of America [205] | <p>Recommendation 1: For symptomatic individuals suspected of having COVID-19, the IDSA panel suggests using standard NAAT (either rapid RT-PCR or laboratory-based NAAT) over rapid Ag tests (<i>conditional recommendation based on moderate certainty in test accuracy of rapid Ag test and very low certainty in comparative test accuracy of rapid RT-PCR versus rapid Ag tests</i>)</p> <p>Recommendation 2: For asymptomatic individuals with risk for exposure to SARS-CoV-2 infection, the IDSA panel suggests using a single standard NAAT (either rapid RT-PCR or laboratory-based NAAT) over a single rapid Ag test (<i>conditional recommendation based on moderate certainty in test accuracy of rapid Ag tests and very low certainty in comparative test accuracy of rapid RT-PCR versus rapid Ag tests</i>)</p> <p>Recommendation 3: For asymptomatic individuals with risk for exposure to SARS-CoV-2 infection, the IDSA panel suggests a single (i.e., one-time)</p> | May 5, 2021 |



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| | | |
|--|---|-------------|
| | <p>standard NAAT (either rapid RT-PCR or laboratory-based NAAT) rather than a strategy of two consecutive rapid Ag tests (<i>conditional recommendation based on moderate certainty in test accuracy of molecular testing and an evidence gap to inform the test accuracy of a strategies using repeat Ag testing</i>)</p> <p>Recommendation 4: In asymptomatic individuals with risk for exposure to SARS-CoV-2 infection, the IDSA panel suggests neither for nor against using single (i.e., one-time) rapid Ag testing over no testing (evidence gap to inform the utility of Ag testing compared to no testing)</p> <p>Recommendation 5: In asymptomatic individuals with risk for exposure to SARS-CoV-2 infection, the IDSA panel suggests neither for nor against using repeat rapid Ag testing over no testing (<i>evidence gap to inform the utility of a strategy of Ag testing compared to no testing</i>).</p> | |
| World Health Organization [206] | <p>WHO recommends the use of Ag-RDTs that meet minimum performance requirements of $\geq 80\%$ sensitivity and $\geq 97\%$ specificity. Ag-RDTs are less sensitive than NAAT, particularly in asymptomatic populations, but careful selection of cohorts for testing can mitigate this limitation.</p> <p>Ag-RDTs should be prioritized for use in symptomatic individuals meeting the case definition for COVID-19, and to test asymptomatic individuals at high risk of infection, including contacts and health workers, particularly in settings where NAAT testing capacity is limited.</p> | Oct 6, 2021 |
| American Academy of Pediatrics [207] | <p>Patients with symptoms consistent with COVID-19 should be tested without delay using either a NAAT (including PCR) or antigen test. The appropriate test for delayed testing of asymptomatic exposed patients is the PCR and not the rapid antigen. Testing for active SARS-CoV-2 infection using NAATs or antigen-based tests is not generally recommended for asymptomatic patients who have previously tested positive within the past 3 months.</p> | Sep 9, 2021 |



References

- [1] World Health Organization. Interim Guidance: Antigen Detection in the Diagnosis of SARS-CoV-2 Infection. Updated 6 October 2021. Accessed at <https://www.who.int/publications/i/item/antigen-detection-in-the-diagnosis-of-sars-cov-2infection-using-rapid-immunoassays> on 26 October 2021.
- [2] Dinnes J, Deeks J, Berhane S, Taylor M, Adriano A, Davenport C et al. Rapid, point-of-care antigen and molecular-based tests for diagnosis of SARS-CoV-2 infection. *Cochrane Database of Systematic Reviews*. 2021;2021(4).
- [3] Burog, AI, Bayona, HH, Cabaluna, IT, Maglente RR. Should rapid antigen tests be used in the diagnosis of COVID-19 in clinically-suspected patients? Evidence Summary. 15 March 2021. PSMID website. Accessed at <https://www.psmid.org/rapid-antigen-test-evidence-summary/> on 26 October 2021.
- [4] Reitsma JB, Glas AS, Rutjes AW, Scholten RJ, Bossuyt PM, Zwinderman AH. Bivariate analysis of sensitivity and specificity produces informative summary measures in diagnostic reviews. *J Clin Epidemiol* 2005; 58: 982-990.
- [5] Harbord RM, Whiting P. metandi: Meta-analysis of diagnostic accuracy using hierarchical logistic regression. *Stata Journal* 2009; 9: 211-229.
- [6] Patel A, Cooper N, Freeman S, Sutton A. Graphical enhancements to summary receiver operating characteristic plots to facilitate the analysis and reporting of meta-analysis of diagnostic test accuracy data. *Res Synth Methods*. 2021;12(1):34–44. <https://doi.org/10.1002/jrsm.1439>.
- [7] Freeman SC, Kerby CR, Patel A, Cooper NJ, Quinn T, Sutton AJ. Development of an interactive web-based tool to conduct and interrogate meta-analysis of diagnostic test accuracy studies: MetaDTA. *BMC Medical Research Methodology* 2019; 19: 81
- [8] Abdelrazik AM, Elshafie SM, Abdelaziz HM. Potential Use of Antigen-Based Rapid Test for SARS-CoV-2 in Respiratory Specimens in Low-Resource Settings in Egypt for Symptomatic Patients and High-Risk Contacts. *Lab Med*. 2021;52(2):e46-e49. doi:10.1093/labmed/lmaa104
- [9] Abusrewil Z, Alhudiri IM, Kaal HH, El Meshri SE, Ebrahim FO, Dalyoum T, et al. Time scale performance of rapid antigen testing for SARS-CoV-2: Evaluation of 10 rapid antigen assays. *J Med Virol*. 2021;(June):4–10. doi: 10.1002/jmv.27186
- [10] Adnan N, Khandker S, Haq A, Chaity M, Khalek A, Nazim A et al. Detection of SARS-CoV-2 by antigen ELISA test is highly swayed by viral load and sample storage condition. *Expert Review of Anti-infective Therapy*. 2021;:1-9.
- [11] Agarwal J, Das A, Pandey P, Sen M, Garg J. "David vs. Goliath": A simple antigen detection test with potential to change diagnostic strategy for SARS-CoV-2. *J Infect Dev Ctries*. 2021 Jul 31;15(7):904-909. doi: 10.3855/jidc.13925. PMID: 34343113.
- [12] Albert E, Torres I, Bueno F, et al. Field evaluation of a rapid antigen test (Panbio™ COVID-19 Ag Rapid Test Device) for COVID-19 diagnosis in primary healthcare centres. *Clin Microbiol Infect*. 2021;27(3):472.e7-472.e10. doi:10.1016/j.cmi.2020.11.004
- [13] Alemany A, Baró B, Ouchi D, et al. Analytical and clinical performance of the panbio COVID-19 antigen-detecting rapid diagnostic test. *J Infect*. 2021;82(5):186-230. doi:10.1016/j.jinf.2020.12.033
- [14] Allan-Blitz LT, Klausner JD. A Real-World Comparison of SARS-CoV-2 Rapid Antigen Testing versus PCR Testing in Florida. *J Clin Microbiol*. 2021 Sep 20;59(10):e0110721. doi: 10.1128/JCM.01107-21. Epub 2021 Aug 4. PMID: 34346715; PMCID: PMC8451433.
- [15] Amer R, Samir M, Gaber O, EL-Deeb N, Abdelmoaty A, Ahmed A et al. Diagnostic performance of rapid antigen test for COVID-19 and the effect of viral load, sampling time, subject's clinical and laboratory parameters on test accuracy. *Journal of Infection and Public Health*. 2021;14(10):1446-1453.
- [16] Andreani J, Lupo J, Germe R, Laugier C, Roccon M, Larrat S et al. Evaluation of six commercial SARS-CoV-2 rapid antigen tests in nasopharyngeal swabs: Better knowledge for better patient management?. *Journal of Clinical Virology*. 2021;143:104947.



Philippine COVID-19 Living Clinical Practice Guidelines

- [17] Asai N, Sakanashi D, Ohashi W, et al. Efficacy and validity of automated quantitative chemiluminescent enzyme immunoassay for SARS-CoV-2 antigen test from saliva specimen in the diagnosis of COVID-19. *J Infect Chemother*. 2021;27(7):1039-1042. doi:10.1016/j.jiac.2021.03.021
- [18] Baccani I, Morecchiato F, Chilleri C, Cervini C, Gori E, Matarrese D et al. Evaluation of Three Immunoassays for the Rapid Detection of SARS-CoV-2 antigens. *Diagnostic Microbiology and Infectious Disease*. 2021;101(2):115434.
- [19] Bachman C, Grant B, Anderson C, Alonzo L, Garing S, Byrnes S et al. Clinical validation of an open-access SARS-CoV-2 antigen detection lateral flow assay, compared to commercially available assays. *PLOS ONE*. 2021;16(8):e0256352.
- [20] Baro B, Rodo P, Ouchi D, Bordoy AE, Saya Amaro EN, Salsench SV, et al. Performance characteristics of five antigen-detecting rapid diagnostic test (Ag-RDT) for SARS-CoV-2 asymptomatic infection: a head-to-head benchmark comparison. *J Infect*. 2021;82(6):269–75. doi: 10.1016/j.jinf.2021.04.009.
- [21] Bello-Chavolla O, Antonio-Villa N, Fernández-Chirino L, Guerra E, Fermín-Martínez C, Márquez-Salinas A et al. Diagnostic performance and clinical implications of rapid SARS-CoV-2 antigen testing in Mexico using real-world nationwide COVID-19 registry data. *PLOS ONE*. 2021;16(8):e0256447.
- [22] Berger A, Nsoga M, Perez-Rodriguez F, Aad Y, Sattonnet-Roche P, Gayet-Ageron A et al. Diagnostic accuracy of two commercial SARS-CoV-2 antigen-detecting rapid tests at the point of care in community-based testing centers. *PLOS ONE*. 2021;16(3):e0248921.
- [23] Bianco G, Boattini M, Barbui A, Scozzari G, Riccardini F, Coggiola M et al. Evaluation of an antigen-based test for hospital point-of-care diagnosis of SARS-CoV-2 infection. *Journal of Clinical Virology*. 2021;139:104838.
- [24] Billaud G, Gaymard A, Lina B, Laboratoire de Virologie des HCL CNR des virus des infections respiratoires. Evaluation du Test Antigénique ABBOTT SARS-COV2 ABBOT. Lyon, France: SFM (French Society of Microbiology), 2020.
- [25] Blairon L, Wilmet A, Beukinga I, Tré-Hardy M. Implementation of rapid SARS-CoV-2 antigenic testing in a laboratory without access to molecular methods: Experiences of a general hospital. *Journal of Clinical Virology*. 2020;129:104472.
- [26] Bornemann L, Kaup O, Kleideiter J, Panning M, Ruprecht B, Wehmeier M. Real-life evaluation of the Sofia SARS-CoV-2 antigen assay in a large tertiary care hospital. *Journal of Clinical Virology*. 2021;140:104854.
- [27] Mboumba Bouassa R, Veyer D, Péré H, Bélec L. Analytical performances of the point-of-care SIENNA™ COVID-19 Antigen Rapid Test for the detection of SARS-CoV-2 nucleocapsid protein in nasopharyngeal swabs: A prospective evaluation during the COVID-19 second wave in France. *International Journal of Infectious Diseases*. 2021;106:8-12.
- [28] Boum Y, Fai KN, Nikolay B, Mboringong AB, Bebell LM, Ndifon M, et al. Performance and operational feasibility of antigen and antibody rapid diagnostic tests for COVID-19 in symptomatic and asymptomatic patients in Cameroon: a clinical, prospective, diagnostic accuracy study. *Lancet Infect Dis*. 2021;21(8):1089–96. [https://doi.org/10.1016/S1473-3099\(21\)00132-8](https://doi.org/10.1016/S1473-3099(21)00132-8)
- [29] Brihn A, Chang J, OYong K, Balter S, Terashita D, Rubin Z et al. Diagnostic Performance of an Antigen Test with RT-PCR for the Detection of SARS-CoV-2 in a Hospital Setting — Los Angeles County, California, June–August 2020. *MMWR Morbidity and Mortality Weekly Report*. 2021;70(19):702-706.
- [30] Bruzzone B, De Pace V, Caligiuri P, Ricucci V, Guarona G, Pennati B et al. Comparative diagnostic performance of rapid antigen detection tests for COVID-19 in a hospital setting. *International Journal of Infectious Diseases*. 2021;107:215-218.
- [31] Bulilete O, Lorente P, Leiva A, et al. Panbio™ rapid antigen test for SARS-CoV-2 has acceptable accuracy in symptomatic patients in primary health care. *J Infect*. 2021;82(3):391-398. doi:10.1016/j.jinf.2021.02.014



Philippine COVID-19 Living Clinical Practice Guidelines

- [32] Caputo V, Bax C, Colantoni L, et al. Comparative analysis of antigen and molecular tests for the detection of Sars-CoV-2 and related variants: A study on 4266 samples. *Int J Infect Dis.* 2021;108:187-189.
- [33] Caruana G, Croxatto A, Kampouri E, Kritikos A, Opota O, Foerster M, et al. Implementing SARS-CoV-2 rapid antigen testing in the emergency ward of a Swiss university hospital: The INCREASE study. *Microorganisms.* 2021;9(4):798.. doi: 10.3390/microorganisms9040798.
- [34] Caruana G, Lebrun L, Aebischer O, Opota O, Urbano L, de Rham M et al. The dark side of SARS-CoV-2 rapid antigen testing: screening asymptomatic patients. *New Microbes and New Infections.* 2021;42:100899.
- [35] Cassuto NG, Gravier A, Colin M, Theillay A, Pires-Roteira D, Pallay S, et al. Evaluation of a SARS-CoV-2 antigen-detecting rapid diagnostic test as a self-test: Diagnostic performance and usability. *J Med Virol.* 2021;93(12):6686–92. doi: 10.1002/jmv.27249.
- [36] Cento V, Renica S, Matarazzo E, Antonello M, Colagrossi L, Di Ruscio F et al. Frontline Screening for SARS-CoV-2 Infection at Emergency Department Admission by Third Generation Rapid Antigen Test: Can We Spare RT-qPCR?. *Viruses.* 2021;13(5):818.
- [37] Cerutti F, Burdino E, Milia MG, Alice T, Gregori G, Bruzzone B, et al. Urgent need of rapid tests for SARS CoV-2 antigen detection: Evaluation of the SD-Biosensor antigen test for SARS-CoV-2. *J Clin Virol.* 2020;132(104654):104654.
- [38] Chaimayo, C., Kaewnaphan, B., Tanlieng, N. et al. Rapid SARS-CoV-2 antigen detection assay in comparison with real-time RT-PCR assay for laboratory diagnosis of COVID-19 in Thailand. *Virol J* 17, 177 (2020). <https://doi.org/10.1186/s12985-020-01452-5>
- [39] Chiu R, Kojima N, Mosley G, Cheng K, Pereira D, Brobeck M et al. Evaluation of the INDICAID COVID-19 Rapid Antigen Test in Symptomatic Populations and Asymptomatic Community Testing. *Microbiology Spectrum.* 2021;9(1).
- [40] Ciotti M, Maurici M, Pieri M, Andreoni M, Bernardini S. Performance of a rapid antigen test in the diagnosis of SARS-CoV-2 infection. *J Med Virol.* 2021;93(5):2988–91. doi: 10.1002/jmv.26830. Epub 2021 Feb 9. PMID: 33527409; PMCID: PMC8014551.
- [41] Courtellemont L, Guinard J, Guillaume C, Giaché S, Rzepecki V, Seve A, et al. High performance of a novel antigen detection test on nasopharyngeal specimens for diagnosing SARS-CoV-2 infection. *J Med Virol.* 2021;93(5):3152–7. doi: 10.1002/jmv.26896.
- [42] Denina M, Giannone V, Curtoni A, Zanotto E, Garazzino S, Urbino A et al. Can we trust in Sars-CoV-2 rapid antigen testing? Preliminary results from a paediatric cohort in the emergency department. *Irish Journal of Medical Science (1971 -).* 2021;.
- [43] Diao B, Wen K, Zhang J, Chen J, Han C, Chen Y, et al. Accuracy of a nucleocapsid protein antigen rapid test in the diagnosis of SARS-CoV-2 infection. *Clin Microbiol Infect.* 2021;27(2):289.e1-289.e4. doi: 10.1016/j.cmi.2020.09.057. Epub 2020 Oct 5. PMID: 33031947; PMCID: PMC7534827.
- [44] Dierks S, Bader O, Schwanbeck J, Groß U, Weig M, Mese K et al. Diagnosing SARS-CoV-2 with Antigen Testing, Transcription-Mediated Amplification and Real-Time PCR. *Journal of Clinical Medicine.* 2021;10(11):2404.
- [45] González-Donapetry P, García-Clemente P, Bloise I, García-Sánchez C, Sánchez Castellano M, Romero M et al. Think of the Children. *Pediatric Infectious Disease Journal.* 2021;40(5):385-388.
- [46] Eleftheriou I, Dasoula F, Dimopoulou D, Lebessi E, Serafi E, Spyridis N, et al. Real-life evaluation of a COVID-19 rapid antigen detection test in hospitalized children. *J Med Virol.* 2021;93(10):6040–4. doi: 10.1002/jmv.27149
- [47] Escrivá B, Mochón M, González R, García C, Pla A, Ricart A et al. “The effectiveness of rapid antigen test-based for SARS-CoV-2 detection in nursing homes in Valencia, Spain”. *Journal of Clinical Virology.* 2021;143:104941.
- [48] Favresse J, Gillot C, Oliveira M, Cadrobbi J, Elsen M, Euchet C, Laffineur K, Rosseels C, Van Eeckhoudt S, Nicolas JB, Morimont L, Dogné JM, Douxflis J. Head-to-Head Comparison of Rapid



Philippine COVID-19 Living Clinical Practice Guidelines

- and Automated Antigen Detection Tests for the Diagnosis of SARS-CoV-2 Infection. *J Clin Med*. 2021 Jan 13;10(2):265. doi: 10.3390/jcm10020265. PMID: 33450853; PMCID: PMC7828347.
- [49] Fenollar F, Bouam A, Ballouche M, Fuster L, Prudent E, Colson P et al. Evaluation of the Panbio COVID-19 Rapid Antigen Detection Test Device for the Screening of Patients with COVID-19. *Journal of Clinical Microbiology*. 2021;59(2).
- [50] Ferté T, Ramel V, Cazanave C, Lafon M, Bébéar C, Malvy D et al. Accuracy of COVID-19 rapid antigenic tests compared to RT-PCR in a student population: The StudyCov study. *Journal of Clinical Virology*. 2021;141:104878.
- [51] Garcíá-Fiñana M, Hughes DM, Cheyne CP, Burnside G, Stockbridge M, Fowler TA, et al. Performance of the Innova SARS-CoV-2 antigen rapid lateral flow test in the Liverpool asymptomatic testing pilot: Population based cohort study. *BMJ*. 2021;374:n1367. doi: 10.1136/bmj.n1637
- [52] Ford L, Lee C, Pray I, Cole D, Bigouette J, Abedi G et al. Epidemiologic Characteristics Associated With Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Antigen-Based Test Results, Real-Time Reverse Transcription Polymerase Chain Reaction (rRT-PCR) Cycle Threshold Values, Subgenomic RNA, and Viral Culture Results From University Testing. *Clinical Infectious Diseases*. 2021;73(6):e1348-e1355.
- [53] Fourati S, Audureau E, Chevaliez S, Pawlotsky JM. Évaluation de la performance diagnostique des tests rapides d'orientation diagnostique antigéniques COVID-19. France: AP-HP Hopitaux universitaires Henri-Mondor, 2020.
- [54] Fourati S, Langendorf C, Audureau E, et al. Performance of six rapid diagnostic tests for SARS-CoV-2 antigen detection and implications for practical use. *J Clin Virol*. 2021;142:104930. doi:10.1016/j.jcv.2021.104930
- [55] Frediani JK, Levy JM, Rao A, et al. Multidisciplinary assessment of the Abbott BinaxNOW SARS-CoV-2 point-of-care antigen test in the context of emerging viral variants and self-administration. *Sci Rep*. 2021;11(1):14604. Published 2021 Jul 16. doi:10.1038/s41598-021-94055-1
- [56] Pérez-García F, Romanyk J, Moya Gutiérrez H, Labrador Ballester A, Pérez Ranz I, González Arroyo J et al. Comparative evaluation of Panbio and SD Biosensor antigen rapid diagnostic tests for COVID-19 diagnosis. *Journal of Medical Virology*. 2021;93(9):5650-5654.
- [57] Gili A, Paggi R, Russo C, Cenci E, Pietrella D, Graziani A, et al. Evaluation of Lumipulse® G SARS-CoV-2 antigen assay automated test for detecting SARS-CoV-2 nucleocapsid protein (NP) in nasopharyngeal swabs for community and population screening. *Int J Infect Dis*. 2021;105:391–6. doi: 10.1016/j.ijid.2021.02.098.
- [58] Gremmels H, Winkel BMF, Schuurman R, Rosingh A, Rigter NAM, Rodriguez O, et al. Real-life validation of the Panbio™ COVID-19 antigen rapid test (Abbott) in community-dwelling subjects with symptoms of potential SARS-CoV-2 infection. *EClinicalMedicine*. 2021;31(100677):100677.DOI: doi.org/10.1016/j.eclinm.2020.100677.
- [59] Gupta A, Khurana S, Das R, Srigan D, Singh A, Mittal A, et al. Rapid chromatographic immunoassay-based evaluation of COVID-19: A cross-sectional, diagnostic test accuracy study & its implications for COVID-19 management in India. *Indian J Med Res*. 2021;153(1 & 2):126–31. DOI: 10.4103/ijmr.IJMR_3305_20.
- [60] Halfon P, Penaranda G, Khiri H, Garcia V, Drouet H, Philibert P et al. An optimized stepwise algorithm combining rapid antigen and RT-qPCR for screening of COVID-19 patients. *PLOS ONE*. 2021;16(9):e0257817.
- [61] Hartard C, Berger S, Josse T, Schvoerer E, Jeulin H. Performance evaluation of an automated SARS-CoV-2 Ag test for the diagnosis of COVID-19 infection on nasopharyngeal swabs. *Clin Chem Lab Med*. 2021;59(12):2003–9.. doi: 10.1515/cclm-2021-0569.
- [62] Häuser, F., Sprinzl, M.F., Dreis, K.J. et al. Evaluation of a laboratory-based high-throughput SARS-CoV-2 antigen assay for non-COVID-19 patient screening at hospital admission. *Med Microbiol Immunol* 210, 165–171 (2021). <https://doi.org/10.1007/s00430-021-00706-5>



Philippine COVID-19 Living Clinical Practice Guidelines

- [63] Herrera V, Hsu V, Adewale A, Johnson L, Hendrix T, Kuhlman J et al. Testing Healthcare Workers Exposed to COVID19 using Rapid Antigen Detection. 2020;.
- [64] Hirotsu Y, Maejima M, Shibusawa M, Nagakubo Y, Hosaka K, Amemiya K, et al. Comparison of automated SARS-CoV-2 antigen test for COVID-19 infection with quantitative RT-PCR using 313 nasopharyngeal swabs, including from seven serially followed patients. *Int J Infect Dis*. 2020;99:397–402. doi: 10.1016/j.ijid.2020.08.029. Epub 2020 Aug 12. PMID: 32800855; PMCID: PMC7422837.
- [65] Holzner C, Pabst D, Anastasiou O, Dittmer U, Manegold R, Risse J et al. SARS-CoV-2 rapid antigen test: Fast-safe or dangerous? An analysis in the emergency department of an university hospital. *Journal of Medical Virology*. 2021;93(9):5323-5327.
- [66] Homza M, Zelena H, Janosek J, Tomaskova H, Jezo E, Kloudova A et al. Performance of Seven SARS-CoV-2 Self-Tests Based on Saliva, Anterior Nasal and Nasopharyngeal Swabs Corrected for Infectiousness in Real-Life Conditions: A Cross-Sectional Test Accuracy Study. *Diagnostics*. 2021;11(9):1567.
- [67] Ifko M, Skvarc M. Use of immunochromatographic sars-cov-2 antigen testing in eight long-term care facilities for the elderly. *Healthc*. 2021;9(7):2–7. doi: 10.3390/healthcare9070868
- [68] Igloi Z, Velzing J, van Beek J, van de Vijver D, Aron G, Ensing R, et al. Clinical evaluation of Roche SD Biosensor rapid antigen test for SARS-CoV-2 in municipal health service testing site, the Netherlands. *Emerg Infect Dis*. 2021;27(5):1323–9. doi: 10.3201/eid2705.204688.
- [69] Iqbal B, Khan M, Shah N, Dawood M, Jehanzeb V, Shafi M. Comparison of SARS-CoV-2 antigen electrochemiluminescence immunoassay to RT-PCR assay for laboratory diagnosis of COVID-19 in Peshawar. *Diagnosis*. 2021;0(0).
- [70] Ishii T, Sasaki M, Yamada K, Kato D, Osuka H, Aoki K, et al. Immunochromatography and chemiluminescent enzyme immunoassay for COVID-19 diagnosis. *J Infect Chemother*. 2021;27(6):915–8.. doi: 10.1016/j.jiac.2021.02.025.
- [71] Jääskeläinen AE, Ahava MJ, Jokela P, Szivovics L, Pohjala S, Vapalahti O, et al. Evaluation of three rapid lateral flow antigen detection tests for the diagnosis of SARS-CoV-2 infection. *J Clin Virol*. 2021;137(104785):104785 doi: 10.1016/j.jcv.2021.104785.
- [72] James AE, Gulley T, Kothari A, Holder K, Garner K, Patil N. Performance of the BinaxNOW coronavirus disease 2019 (COVID-19) Antigen Card test relative to the severe acute respiratory coronavirus virus 2 (SARS-CoV-2) real-time reverse transcriptase polymerase chain reaction (rRT-PCR) assay among symptomatic and asymptomatic healthcare employees. *Infect Control Hosp Epidemiol*. 2021;1–3.. doi: 10.1017/ice.2021.20.
- [73] Jegerlehner S, Suter-Riniker F, Jent P, Bittel P, Nagler M. Diagnostic accuracy of a SARS-CoV-2 rapid antigen test in real-life clinical settings: Antigen tests in real-life clinical settings. *Int J Infect Dis*. 2021;109(January):118–22. doi: <https://doi.org/10.1016/j.ijid.2021.07.01>
- [74] Jung C, Levy C, Varon E, et al. Diagnostic Accuracy of SARS-CoV-2 Antigen Detection Test in Children: A Real-Life Study. *Front Pediatr*. 2021;9:647274. Published 2021 Jul 15. doi:10.3389/fped.2021.647274
- [75] Kahn M, Schuierer L, Bartenschlager C, Zellmer S, Frey R, Freitag M et al. Performance of antigen testing for diagnosis of COVID-19: a direct comparison of a lateral flow device to nucleic acid amplification based tests. *BMC Infectious Diseases*. 2021;21(1).
- [76] Kanaujia R, Ghosh A, Mohindra R, et al. Rapid antigen detection kit for the diagnosis of SARS-CoV-2 - are we missing asymptomatic patients? [published online ahead of print, 2021 Jul 19]. *Indian J Med Microbiol*. 2021;S0255-0857(21)04168-2. doi:10.1016/j.ijmmb.2021.07.003
- [77] Karon BS, Donato LJ, Bridgeman AR, Blommel JH, Kipp B, Maus A, et al. Analytical Sensitivity and Specificity of Four Point of Care Rapid Antigen Diagnostic Tests for SARS-CoV-2 Using Real-Time Quantitative PCR, Quantitative Droplet Digital PCR, and a Mass Spectrometric Antigen Assay as Comparator Methods. *Clin Chem*. 2021;9:1–9. doi: 10.1093/clinchem/hvab138



Philippine COVID-19 Living Clinical Practice Guidelines

- [78] Kim D, Lee J, Bal J, Seo S, Chong C, Lee J et al. Development and Clinical Evaluation of an Immunochromatography-Based Rapid Antigen Test (GenBody™ COVAG025) for COVID-19 Diagnosis. *Viruses*. 2021;13(5):796.
- [79] Mathur P, Kiro V, Gupta A, Singh P, Sharad N, Khurana S et al. Evaluation of COVID-19 antigen fluorescence immunoassay test for rapid detection of SARS-CoV-2. *Journal of Global Infectious Diseases*. 2021;13(2):91.
- [80] Klein JAF, Krüger LJ, Tobian F, Gaeddert M, Lainati F, Schnitzler P, et al. Head-to-head performance comparison of self-collected nasal versus professional-collected nasopharyngeal swab for a WHO-listed SARS-CoV-2 antigen-detecting rapid diagnostic test. *Med Microbiol Immunol [Internet]*. 2021;210(4):181–6. Available from: <https://doi.org/10.1007/s00430-021-00710-9> doi: 10.1007/s00430-021-00710-9
- [81] Kobayashi R, Murai R, Moriai M, Nirasawa S, Yonezawa H, Kondoh T, et al. Evaluation of false positives in the SARS-CoV-2 quantitative antigen test. *J Infect Chemother*. 2021;27(10):1477–81. doi: 10.1016/j.jiac.2021.06.019.
- [82] Koeleman J, Brand H, de Man S, Ong D. Clinical evaluation of rapid point-of-care antigen tests for diagnosis of SARS-CoV-2 infection. *European Journal of Clinical Microbiology & Infectious Diseases*. 2021;40(9):1975-1981.
- [83] Kolwijck E, Brouwers-Boers M, Broertjes J, van Heeswijk K, Runderkamp N, Meijer A et al. Validation and implementation of the Panbio COVID-19 Ag rapid test for the diagnosis of SARS-CoV-2 infection in symptomatic hospital Trained personnels. *Infection Prevention in Practice*. 2021;3(2):100142.
- [84] Korenkov M, Poopalasingam N, Madler M, Vanshylla K, Eggeling R, Wirtz M et al. Evaluation of a Rapid Antigen Test To Detect SARS-CoV-2 Infection and Identify Potentially Infectious Individuals. *Journal of Clinical Microbiology*. 2021;59(9).
- [85] Krüger L, Gaeddert M, Köppel L, Brümmer L, Gottschalk C, Miranda I et al. Evaluation of the accuracy, ease of use and limit of detection of novel, rapid, antigen-detecting point-of-care diagnostics for SARS-CoV-2. 2020;.
- [86] Krüger L, Gaeddert M, Tobian F, Lainati F, Gottschalk C, Klein J et al. The Abbott PanBio WHO emergency use listed, rapid, antigen-detecting point-of-care diagnostic test for SARS-CoV-2—Evaluation of the accuracy and ease-of-use. *PLOS ONE*. 2021;16(5):e0247918.
- [87] Krüger L, Klein J, Tobian F, Gaeddert M, Lainati F, Klemm S et al. Evaluation of accuracy, exclusivity, limit-of-detection and ease-of-use of LumiraDx™: An antigen-detecting point-of-care device for SARS-CoV-2. *Infection*. 2021;.
- [88] Krüttgen A, Cornelissen C, Dreher M, Hornef M, Imöhl M, Kleines M. Comparison of the SARS-CoV-2 Rapid antigen test to the real star Sars-CoV-2 RT PCR kit. *Journal of Virological Methods*. 2021;288:114024.
- [89] Kurihara Y, Kiyasu Y, Akashi Y, Takeuchi Y, Narahara K, Mori S, et al. The evaluation of a novel digital immunochromatographic assay with silver amplification to detect SARS-CoV-2. *J Infect Chemother*. 2021;27(10):1493–7. doi: 10.1016/j.jiac.2021.07.006
- [90] Kweon O, Lim Y, Kim H, Choi Y, Kim M, Choi S et al. Evaluation of rapid SARS-CoV-2 antigen tests, AFIAS COVID-19 Ag and ichroma COVID-19 Ag, with serial nasopharyngeal specimens from COVID-19 patients. *PLOS ONE*. 2021;16(4):e0249972.
- [91] Kyritsi M, Vontas A, Voulgaridi I, Matziri A, Komnos A, Babalis D et al. Rapid Test Ag 2019-nCoV (PROGNOSIS, BIOTECH, Larissa, Greece); Performance Evaluation in Hospital Setting with Real Time RT-PCR. *International Journal of Environmental Research and Public Health*. 2021;18(17):9151.
- [92] Lambert-Niclot S, Cuffel A, Le Pape S, Vauloup-Fellous C, Morand-Joubert L, Roque-Afonso A-M, et al. Evaluation of a rapid diagnostic assay for detection of SARS-CoV-2 antigen in nasopharyngeal swabs. *J Clin Microbiol [Internet]*. 2020;58(8). Available from: <http://dx.doi.org/10.1128/JCM.00977-20>.



Philippine COVID-19 Living Clinical Practice Guidelines

- [93] Landaas ET, Storm ML, Tollånes MC, Barlinn R, Kran A-MB, Bragstad K, et al. Diagnostic performance of a SARS-CoV-2 rapid antigen test in a large, Norwegian cohort. *J Clin Virol.* 2021;137(104789):104789. doi: 10.1016/j.jcv.2021.104789.
- [94] Leixner G, Voill-Glaninger A, Bonner E, Kreil A, Zadnikar R, Viveiros A. Evaluation of the AMP SARS-CoV-2 rapid antigen test in a hospital setting. *International Journal of Infectious Diseases.* 2021;108:353-356.
- [95] Leli C, Di Matteo L, Gotta F, et al. Performance of a SARS-CoV-2 antigen rapid immunoassay in patients admitted to the emergency department. *Int J Infect Dis.* 2021;110:135-140. doi:10.1016/j.ijid.2021.07.043
- [96] L'Huillier A, Lacour M, Sadiku D, Gadiri M, De Siebenthal L, Schibler M et al. Diagnostic Accuracy of SARS-CoV-2 Rapid Antigen Detection Testing in Symptomatic and Asymptomatic Children in the Clinical Setting. *Journal of Clinical Microbiology.* 2021;59(9).
- [97] Linares M, Pérez-Tanoira R, Carrero A, Romanyk J, Pérez-García F, Gómez-Herruz P et al. Panbio antigen rapid test is reliable to diagnose SARS-CoV-2 infection in the first 7 days after the onset of symptoms. *Journal of Clinical Virology.* 2020;133:104659.
- [98] Lindner AK, Nikolai O, Rohardt C, Burock S, Hülso C, Bölke A, et al. Head-to-head comparison of SARS-CoV-2 antigen-detecting rapid test with professional-collected nasal versus nasopharyngeal swab. *Eur Respir J.* 2021 May 6;57(5):2004430. doi: 10.1183/13993003.04430-2020.
- [99] Lindner AK, Nikolai O, Rohardt C, Kausch F, Wintel M, Gertler M, et al.. Diagnostic accuracy and feasibility of patient self-testing with a SARS-CoV-2 antigen-detecting rapid test. *J Clin Virol.* 2021 Aug;141:104874. doi: 10.1016/j.jcv.2021.104874.
- [100] Liotti F, Menchinelli G, Lalle E, Palucci I, Marchetti S, Colavita F et al. Performance of a novel diagnostic assay for rapid SARS-CoV-2 antigen detection in nasopharynx samples. *Clinical Microbiology and Infection.* 2021;27(3):487-488.
- [101] Mak G, Cheng P, Lau S, Wong K, Lau C, Lam E et al. Evaluation of rapid antigen test for detection of SARS-CoV-2 virus. *Journal of Clinical Virology.* 2020;129:104500.
- [102] Maniscalco M, Ambrosino P, Ciullo A, Fuschillo S, Valente V, Gaudiosi C et al. A Rapid Antigen Detection Test to Diagnose SARS-CoV-2 Infection Using Exhaled Breath Condensate by A Modified Inflammacheck® Device. *Sensors.* 2021;21(17):5710.
- [103] Martín-Sánchez V, Fernández-Villa T, Carvajal Urueña A, Rivero Rodríguez A, Reguero Celada S, Sánchez Antolín G et al. Role of Rapid Antigen Testing in Population-Based SARS-CoV-2 Screening. *Journal of Clinical Medicine.* 2021;10(17):3854.
- [104] Matsuda EM, de Campos IB, de Oliveira IP, Colpas DR, Carmo AMDS, Brígido LFM. Field evaluation of COVID-19 antigen tests versus RNA based detection: Potential lower sensitivity compensated by immediate results, technical simplicity, and low cost. *J Med Virol.* 2021 Jul;93(7):4405-4410. doi: 10.1002/jmv.26985. Epub 2021 Apr 8. PMID: 33788270; PMCID: PMC8250877.
- [105] Mboma O, Rieke E, Ahmad-Nejad P, Wirth S, Aydin M. Diagnostic Performance of SARS-CoV-2 Rapid Antigen Test in a Large, German Cohort. *Children (Basel).* 2021 Aug 8;8(8):682. doi: 10.3390/children8080682. PMID: 34438573; PMCID: PMC8394520.
- [106] McKay S, Tobolowsky F, Moritz E, Hatfield K, Bhatnagar A, LaVoie S et al. Performance Evaluation of Serial SARS-CoV-2 Rapid Antigen Testing During a Nursing Home Outbreak. *Annals of Internal Medicine.* 2021;174(7):945-951.
- [107] Merino-Amador P, González-Donapetry P, Domínguez-Fernández M, González-Romo F, Sánchez-Castellano M, Seoane-Estevez A et al. Clinitest rapid COVID-19 antigen test for the diagnosis of SARS-CoV-2 infection: A multicenter evaluation study. *Journal of Clinical Virology.* 2021;143:104961.
- [108] Mertens P, De Vos N, Martiny D, Jassoy C, Mirazimi A, Cuypers L, et al. Development and potential usefulness of the COVID-19 Ag Respi-Strip diagnostic assay in a pandemic context. *Front Med (Lausanne).* 2020;7:225.. doi: 10.3389/fmed.2020.00225. PMID: 32574326; PMCID: PMC7227790.



Philippine COVID-19 Living Clinical Practice Guidelines

- [109] Möckel M, Corman V, Stegemann M, Hofmann J, Stein A, Jones T et al. SARS-CoV-2 antigen rapid immunoassay for diagnosis of COVID-19 in the emergency department. *Biomarkers*. 2021;26(3):213-220.
- [110] Fernandez-Montero A, Argemi J, Rodríguez J, Ariño A, Moreno-Galarraga L. Validation of a Rapid Antigen Test as a Screening Tool for SARS-CoV-2 Infection in Asymptomatic Populations. Sensitivity, Specificity and Predictive Values. *SSRN Electronic Journal*. 2021;.
- [111] Muhi S, Tayler N, Hoang T, et al. Multi-site assessment of rapid, point-of-care antigen testing for the diagnosis of SARS-CoV-2 infection in a low-prevalence setting: A validation and implementation study. *Lancet Reg Health West Pac*. 2021;9:100115. doi:10.1016/j.lanwpc.2021.100115
- [112] Nagura-Ikeda M, Imai K, Tabata S, Miyoshi K, Murahara N, Mizuno T, et al. Clinical Evaluation of Self-Collected Saliva by Quantitative Reverse Transcription-PCR (RT-qPCR), Direct RT-qPCR, Reverse Transcription-Loop-Mediated Isothermal Amplification, and a Rapid Antigen Test To Diagnose COVID-19. *J Clin Microbiol*. 2020 Aug 24;58(9):e01438-20. doi: 10.1128/JCM.01438-20. PMID: 32636214; PMCID: PMC7448663.
- [113] Nalumansi A, Lutalo T, Kayiwa J, Watera C, Balinandi S, Kiconco J, et al. Field evaluation of the performance of a SARS-CoV-2 antigen rapid diagnostic test in Uganda using nasopharyngeal samples. *Int J Infect Dis*. 2021;104:282–6. doi: 10.1016/j.ijid.2020.10.073. Epub 2020 Oct 30. PMID: 33130198; PMCID: PMC7836828.
- [114] Nash B, Badea A, Reddy A, Bosch M, Salcedo N, Gomez A et al. Validating and modeling the impact of high-frequency rapid antigen screening on COVID-19 spread and outcomes. 2020;.
- [115] Nikolai O, Rohardt C, Tobian F, Junge A, Corman V, Jones T et al. Anterior nasal versus nasal mid-turbinate sampling for a SARS-CoV-2 antigen-detecting rapid test: does localisation or professional collection matter?. *Infectious Diseases*. 2021;;1-6.
- [116] Nomoto H, Yamamoto K, Yamada G, Suzuki M, Kinoshita N, Takasaki J et al. Time-course evaluation of the quantitative antigen test for severe acute respiratory syndrome coronavirus 2: The potential contribution to alleviating isolation of COVID-19 patients. *Journal of Infection and Chemotherapy*. 2021;27(11):1669-1673.
- [117] Nordgren J, Sharma S, Olsson H, Jämtberg M, Falkeborn T, Svensson L et al. SARS-CoV-2 rapid antigen test: High sensitivity to detect infectious virus. *Journal of Clinical Virology*. 2021;140:104846.
- [118] Nörz D, Olearo F, Perisic S, Bauer M, Riester E, Schneider T et al. Multicenter Evaluation of a Fully Automated High-Throughput SARS-CoV-2 Antigen Immunoassay. *Infectious Diseases and Therapy*. 2021;.
- [119] Ngo Nsoga MT, Kronig I, Perez Rodriguez FJ, et al. Diagnostic accuracy of Panbio rapid antigen tests on oropharyngeal swabs for detection of SARS-CoV-2. *PLoS One*. 2021;16(6):e0253321. Published 2021 Jun 24. doi:10.1371/journal.pone.0253321
- [120] Oh S-M, Jeong H, Chang E, Choe PG, Kang CK, Park WB, et al. Clinical application of the standard Q COVID-19 Ag test for the detection of SARS-CoV-2 infection. *J Korean Med Sci*. 2021;36(14). doi: 10.3346/jkms.2021.36.e101.
- [121] Okoye NC, Barker AP, Curtis K, Orlandi RR, Snaveley EA, Wright C, et al. Performance Characteristics of BinaxNOW COVID-19 Antigen Card for Screening Asymptomatic Individuals in a University Setting. *J Clin Microbiol*. 2021 Mar 19;59(4):e03282-20. doi: 10.1128/JCM.03282-20.
- [122] Olearo F, Nörz D, Heinrich F, Sutter JP, Roedl K, Schultze A, et al.. Handling and accuracy of four rapid antigen tests for the diagnosis of SARS-CoV-2 compared to RT-qPCR. *J Clin Virol*. 2021 Apr;137:104782. doi: 10.1016/j.jcv.2021.104782.
- [123] Orsi A, Pennati BM, Bruzzzone B, Ricucci V, Ferone D, Barbera P, et al. On-field evaluation of a ultra-rapid fluorescence immunoassay as a frontline test for SARS-CoV-2 diagnostic. *J Virol Methods*. 2021;295(114201):114201.. doi: 10.1016/j.jviromet.2021.114201. Epub 2021 May 28. PMID: 34058287; PMCID: PMC8161776.



Philippine COVID-19 Living Clinical Practice Guidelines

- [124] Osmanodja B, Budde K, Zickler D, Naik M, Hofmann J, Gertler M et al. Accuracy of a Novel SARS-CoV-2 Antigen-Detecting Rapid Diagnostic Test from Standardized Self-Collected Anterior Nasal Swabs. *Journal of Clinical Medicine*. 2021;10(10):2099.
- [125] Osterman A, Baldauf HM, Eleteby M, et al. Evaluation of two rapid antigen tests to detect SARS-CoV-2 in a hospital setting. *Med Microbiol Immunol*. 2021;210(1):65-72. doi:10.1007/s00430-020-00698-8
- [126] Osterman A, Iglhaut M, Lehner A, Späth P, Stern M, Autenrieth H et al. Comparison of four commercial, automated antigen tests to detect SARS-CoV-2 variants of concern. *Medical Microbiology and Immunology*. 2021;.
- [127] Paul D, Gupta A, Rooge S, Gupta E. Performance evaluation of automated chemiluminescence immunoassay based antigen detection – Moving towards more reliable ways to predict SARS-CoV-2 infection. *Journal of Virological Methods*. 2021;298:114299.
- [128] Peña M, Ampuero M, Garcés C, Gaggero A, García P, Velasquez M et al. Performance of SARS-CoV-2 rapid antigen test compared with real-time RT-PCR in asymptomatic individuals. *International Journal of Infectious Diseases*. 2021;107:201-204.
- [129] Peña-Rodríguez M, Viera-Segura O, García-Chagollán M, Zepeda-Nuño JS, Muñoz-Valle JF, Mora-Mora J, et al. Performance evaluation of a lateral flow assay for nasopharyngeal antigen detection for SARS-CoV-2 diagnosis. *J Clin Lab Anal*. 2021;35(5):e23745. <https://doi.org/10.1002/jcla.23745>
- [130] Public Health England. Preliminary report from the Joint PHE Porton Down & University of Oxford SARS-CoV-2 test development and validation cell : Rapid evaluation of Lateral Flow Viral Antigen detection devices (LFDs) for mass community testing : 2020;(November):2–7.
- [131] Pickering S, Batra R, Merrick B, Snell L, Nebbia G, Douthwaite S et al. Comparative performance of SARS-CoV-2 lateral flow antigen tests and association with detection of infectious virus in clinical specimens: a single-centre laboratory evaluation study. *The Lancet Microbe*. 2021;2(9):e461-e471.
- [132] Pilarowski G, Marquez C, Rubio L, Peng J, Martinez J, Black D, et al. Field performance and public health response using the BinaxNOW TM Rapid SARS-CoV-2 antigen detection assay during community-based testing. *Clin Infect Dis*. 2020;73(9):e3098–101.
- [133] Pollock NR, Jacobs JR, Tran K, Cranston AE, Smith S, O’Kane CY, et al. Performance and implementation evaluation of the Abbott BinaxNOW rapid antigen test in a high-throughput drive-through community testing site in Massachusetts. *J Clin Microbiol*. 2021;59(5). Available from: <http://dx.doi.org/10.1128/JCM.00083-21>
- [134] Pollock NR, Tran K, Jacobs JR, Cranston AE, Smith S, O’Kane CY, et al. Performance and operational evaluation of the Access Bio CareStart rapid Antigen test in a high-throughput drive-through community testing site in Massachusetts. *Open Forum Infect Dis*. 2021;8(7):ofab243. <https://doi.org/10.1093/ofid/ofab243>
- [135] Porte L, Legarraga P, Vollrath V, Aguilera X, Munita J, Araos R et al. Evaluation of a novel antigen-based rapid detection test for the diagnosis of SARS-CoV-2 in respiratory samples. *International Journal of Infectious Diseases*. 2020;99:328-333.
- [136] Porte L, Legarraga P, Iruretagoyena M, Vollrath V, Pizarro G, Munita J et al. Evaluation of two fluorescence immunoassays for the rapid detection of SARS-CoV-2 antigen—new tool to detect infective COVID-19 patients. *PeerJ*. 2021;9:e10801.
- [137] Pray IW, Ford L, Cole D, Lee C, Bigouette JP, Abedi GR, et al. Performance of an antigen-based test for asymptomatic and symptomatic SARS-CoV-2 testing at two university campuses - Wisconsin, September-October 2020. *MMWR Morb Mortal Wkly Rep*. 2021;69(5152):1642–7 doi: <http://dx.doi.org/10.15585/mmwr.mm695152a3>
- [138] Prince-Guerra JL, Almendares O, Nolen LD, Gunn JKL, Dale AP, Buono SA, et al. Evaluation of Abbott BinaxNOW rapid antigen test for SARS-CoV-2 infection at two community-based testing sites - Pima County, Arizona, November 3-17, 2020. *MMWR Morb Mortal Wkly Rep*. 2021;70(3):100–5. doi: <http://dx.doi.org/10.15585/mmwr.mm7003e3>



Philippine COVID-19 Living Clinical Practice Guidelines

- [139] Ristić M, Nikolić N, Čabarkapa V, Turkulov V, Petrović V. Validation of the STANDARD Q COVID-19 antigen test in Vojvodina, Serbia. *PLoS One*. 2021;16(2 February):1–13. doi: 10.1371/journal.pone.0247606
- [140] Rottenstreich A, Zarbiv G, Kabiri D, Porat S, Sompolinsky Y, Reubinoff B, et al. Rapid antigen detection testing for universal screening for severe acute respiratory syndrome coronavirus 2 in women admitted for delivery. *Am J Obstet Gynecol*. 2021;224(5):539–40. doi: 10.1016/j.ajog.2021.01.002
- [141] Carbonell-Sahuquillo S, Lázaro-Carreño MI, Camacho J, Barrés-Fernández A, Albert E, Torres I, et al. Evaluation of a rapid antigen detection test (Panbio™ COVID-19 Ag Rapid Test Device) as a point-of-care diagnostic tool for COVID-19 in a pediatric emergency department. *J Med Virol*. 2021;93(12):6803–7. doi: 10.1002/jmv.27220
- [142] Salvagno G, Gianfilippi G, Fiorio G, Pighi L, De Nitto S, Henry B et al. Performance of Fujirebio Espline SARS-CoV-2 Rapid Antigen Test for Identifying Potentially Infectious Individuals. *SSRN Electronic Journal*. 2021;.
- [143] Schildgen V, Demuth S, Lüsebrink J, Schildgen O. Limits and opportunities of SARS-CoV-2 antigen rapid tests: An experienced-based perspective. *Pathogens*. 2021;10(1):38. doi: Schildgen V, Demuth S, Lüsebrink J, Schildgen O. Limits and opportunities of SARS-CoV-2 antigen rapid tests: An experienced-based perspective. *Pathogens*. 2021;10(1):38.
- [144] Schuit E, Veldhuijzen IK, Venekamp RP, et al. Diagnostic accuracy of rapid antigen tests in asymptomatic and presymptomatic close contacts of individuals with confirmed SARS-CoV-2 infection: cross sectional study. *BMJ*. 2021;374:n1676. Published 2021 Jul 27. doi:10.1136/bmj.n1676
- [145] Scohy A, Anantharajah A, Bodéus M, Kabamba-Mukadi B, Verroken A, Rodriguez-Villalobos H. Low performance of rapid antigen detection test as frontline testing for COVID-19 diagnosis. *J Clin Virol*. 2020;129(104455):104455. doi: doi.org/10.1016/j.jcv.2020.104455
- [146] Seitz T, Schindler S, Winkelmeier P, Zach B, Wenisch C, Zoufaly A, et al. Evaluation of rapid antigen tests based on saliva for the detection of SARS-CoV-2. *J Med Virol*. 2021;93(7):4161–2. doi: 10.1002/jmv.26983
- [147] Seynaeve Y, Heylen J, Fontaine C, et al. Evaluation of Two Rapid Antigenic Tests for the Detection of SARS-CoV-2 in Nasopharyngeal Swabs. *J Clin Med*. 2021;10(13):2774. Published 2021 Jun 24. doi:10.3390/jcm10132774
- [148] Shah M, Salvatore P, Ford L, Kamitani E, Whaley M, Mitchell K et al. Performance of Repeat BinaxNOW Severe Acute Respiratory Syndrome Coronavirus 2 Antigen Testing in a Community Setting, Wisconsin, November 2020–December 2020. *Clinical Infectious Diseases*. 2021;73(Supplement_1):S54-S57.
- [149] Shaikh N, Friedlander E, Tate P, Liu H, Chang C, Wells A et al. Performance of a Rapid SARS-CoV-2 Antigen Detection Assay in Symptomatic Children. *Pediatrics*. 2021;148(3):e2021050832.
- [150] Shrestha B, Neupane AK, Pant S, Shrestha A, Bastola A, Rajbhandari B, et al. Sensitivity and specificity of lateral flow antigen test kits for COVID-19 in asymptomatic population of Quarantine centre of Province 3. *Kathmandu Univ Med J (KUMJ)*. 2020;18(2):36–9.
- [151] Soleimani R, Deckers C, Huang T, Bogaerts P, Evrard S, Wallemme I et al. Rapid COVID-19 antigenic tests: Usefulness of a modified method for diagnosis. *Journal of Medical Virology*. 2021;93(9):5655-5659.
- [152] Sood N, Shetgiri R, Rodriguez A, Jimenez D, Treminino S, Daflos A, et al. Evaluation of the Abbott BinaxNOW rapid antigen test for SARS-CoV-2 infection in children: Implications for screening in a school setting. *PLoS One*. 2021;16(4):e0249710. doi: 10.1371/journal.pone.0249710.
- [153] Stohr JJM, Zwart VF, Goderski G, Meijer A, Nagel-Imming CRS, Kluytmans-van den Bergh MFQ, et al. Self-testing for the detection of SARS-CoV-2 infection with rapid antigen tests for people with suspected COVID-19 in the community. *Clin Microbiol Infect [Internet]*. 2021; Available from: <http://dx.doi.org/10.1016/j.cmi.2021.07.039>.



Philippine COVID-19 Living Clinical Practice Guidelines

- [154] Stokes NL, Reed KA, Berbari EF, Vetter S, Binnicker MJ. Evaluation of the BinaxNOW COVID-19 rapid antigen test in an asymptomatic patient population undergoing pre-procedural screening. *J Clin Microbiol.* 2021;JCM0165021.
- [155] Stokes W, Berenger BM, Portnoy D, Scott B, Szelewicki J, Singh T, et al. Clinical performance of the Abbott Panbio with nasopharyngeal, throat, and saliva swabs among symptomatic individuals with COVID-19. *Eur J Clin Microbiol Infect Dis.* 2021;40(8):1721–6. doi: 10.1007/s10096-021-04202-9.
- [156] Takeda Y, Mori M, Omi K. SARS-CoV-2 qRT-PCR Ct value distribution in Japan and possible utility of rapid antigen testing kit [Internet]. *bioRxiv.* 2020. Available from: <http://dx.doi.org/10.1101/2020.06.16.20131243>
- [157] Thakur P, Saxena S, Manchanda V, Rana N, Goel R, Arora R. Utility of Antigen-Based Rapid Diagnostic Test for Detection of SARS-CoV-2 Virus in Routine Hospital Settings. *Laboratory Medicine.* 2021;.
- [158] Torres I, Poujois S, Albert E, Colomina J, Navarro D. Evaluation of a rapid antigen test (Panbio™ COVID-19 Ag rapid test device) for SARS-CoV-2 detection in asymptomatic close contacts of COVID-19 patients. *Clin Microbiol Infect.* 2021 Apr;27(4):636.e1-636.e4. doi: 10.1016/j.cmi.2020.12.022.
- [159] Tsai S, Lee W, Chen P, Hung S. Real world clinical performance of SARS-CoV-2 rapid antigen tests in suspected COVID-19 cases in Taiwan. *Journal of the Formosan Medical Association.* 2021;120(11):2042-2043.
- [160] Turcato G, Zaboli A, Pfeifer N, Ciccariello L, Sibilio S, Tezza G, et al. Clinical application of a rapid antigen test for the detection of SARS-CoV-2 infection in symptomatic and asymptomatic patients evaluated in the emergency department: A preliminary report. *J Infect.* 2021;82(3):e14–6.
- [161] Uwamino Y, Nagata M, Aoki W, Nakagawa T, Inose R, Yokota H, et al. Accuracy of rapid antigen detection test for nasopharyngeal swab specimens and saliva samples in comparison with RT-PCR and viral culture for SARS-CoV-2 detection. *J Infect Chemother.* 2021;27(7):1058–62.
- [162] Van der Moeren N, Zwart VF, Lodder EB, Van den Bijllaardt W, Van Esch HRJM, Stohr JJM, et al. Evaluation of the test accuracy of a SARS-CoV-2 rapid antigen test in symptomatic community dwelling individuals in the Netherlands. *PLoS One* [Internet]. 2021;16(5 May):1–11. Available from: <http://dx.doi.org/10.1371/journal.pone.0250886>
- [163] Van Honacker E, Van Vaerenbergh K, Boel A, De Beenhouwer H, Leroux-Roels I, Cattoir L. Comparison of five SARS-CoV-2 rapid antigen detection tests in a hospital setting and performance of one antigen assay in routine practice: a useful tool to guide isolation precautions?. *Journal of Hospital Infection.* 2021;114:144-152.
- [164] Veyrenche N, Bolloré K, Pisoni A, Bedin A-S, Mondain A-M, Ducos J, et al. Diagnosis value of SARS-CoV-2 antigen/antibody combined testing using rapid diagnostic tests at hospital admission [Internet]. *bioRxiv.* 2020. Available from: <http://dx.doi.org/10.1101/2020.09.19.20197855>
- [165] Von Ahnen T, von Ahnen M, Wirth U, Schardey H, Herdtle S. Evaluation of a rapid-antigen test for COVID-19 in an asymptomatic collective. *Wiener Medizinische Wochenschrift.* 2021;.
- [166] Wachinger J, Olaru ID, Horner S, Schnitzler P, Heeg K, Denking CM. The potential of SARS-CoV-2 antigen-detection tests in the screening of asymptomatic persons [published online ahead of print, 2021 Jul 26]. *Clin Microbiol Infect.* 2021;S1198-743X(21)00412-2. doi:10.1016/j.cmi.2021.07.020
- [167] Wagenhäuser I, Knies K, Rauschenberger V, Eisenmann M, McDonogh M, Petri N, et al. Clinical performance evaluation of SARS-CoV-2 rapid antigen testing in point of care usage in comparison to RT-qPCR. *EBioMedicine.* 2021;69(103455):103455. *EBioMedicine.* 2021 Jul;69:103455. doi: 10.1016/j.ebiom.2021.103455. Epub 2021 Jun 26. PMID: 34186490; PMCID: PMC8234263.
- [168] Weitzel T, Legarraga P, Iruretagoyena M, Pizarro G, Vollrath V, Araos R, et al. Head-to-head comparison of four antigen-based rapid detection tests for the diagnosis of SARS-CoV-2 in respiratory samples [Internet]. *bioRxiv.* 2020. Available from: <http://dx.doi.org/10.1101/2020.05.27.119255>



Philippine COVID-19 Living Clinical Practice Guidelines

- [169] Yokota I, Sakurazawa T, Sugita J, Iwasaki S, Yasuda K, Yamashita N, Fujisawa S, Nishida M, Konno S, Teshima T. Performance of Qualitative and Quantitative Antigen Tests for SARS-CoV-2 Using Saliva. *Infect Dis Rep*. 2021 Aug 24;13(3):742-747. doi: 10.3390/idr13030069.
- [170] Young S, Taylor SN, Cammarata CL, Roger-Dalbert C, Montano A, Griego-Fullbright C, et al. Clinical evaluation of BD Veritor™ SARS-CoV-2 point-of-care test performance compared to PCR-based testing and versus the Sofia® 2 SARS Antigen point-of-care test [Internet]. *bioRxiv*. 2020. Available from: <http://dx.doi.org/10.1101/2020.09.01.20185777>.
- [171] Study to Evaluate the Performance of the Thermo COVID-19 Rapid Antigen Test for Detection of SARS-CoV-2 [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04878068?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=2&rank=1>
- [172] Clinical Evaluation of a Point-of-Care (POC), COVID-19 Rapid Antigen Test (CovIDx™) - Full Text View - *ClinicalTrials.Gov* [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04750629?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=2&rank=2>
- [173] Diagnostic Accuracy of Rapid Antigen Test Based on Anterior Nasal Swab Compared With RT-PCR for SARS-CoV-2 Detection [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT05045846?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=2&rank=4>
- [174] Clinical Performance Evaluation of KnowNow SARS-CoV-2 Test for the Detection of COVID-19 Antigen [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04733170?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=2&rank=5>
- [175] Sensitivity and Specificity of SARS-CoV-2 Rapid Antigen Test Compared to RT-PCR Test [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04689399?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=2&rank=6>
- [176] Rapid Antigen Testing for SARS-CoV-2 Among Healthcare Workers to Prevent Spread of COVID-19 [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04716088?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=2&rank=7>
- [177] Evaluation of a COVID-19 Rapid Diagnostic Test in ER Departments in Mexico: A Multi-center Study [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04894760?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=2&rank=8>
- [178] Somerset and South Essex Coronavirus Antigen Testing - Full Text View - *ClinicalTrials.Gov* [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04403906?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=2&rank=9>
- [179] Dry Versus Wet Nasopharyngeal Rapid Test for the Detection of COVID-19. - Full Text View - *ClinicalTrials.Gov* [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04839094?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=2&rank=10>
- [180] An Approach to Screening for COVID-19 at Vancouver Airport [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04665193?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=3&rank=11>
- [181] COVAG - Covid-19 Antigen Study - the Diagnostic Efficacy of SARS-CoV-2 Rapid Detection Tests - Full Text View - *ClinicalTrials.Gov* [Internet]. *Clinicaltrials.gov*. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT05074017?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=3&rank=12>



Philippine COVID-19 Living Clinical Practice Guidelines

- [182] A Clinical Evaluation of COVID-19 Rapid Point of Care Antigen Tests [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04568356?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=3&rank=13>
- [183] COVID-19 Rapid Testing for Self-Administration Among an Asymptomatic Sample [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04896710?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=3&rank=15>
- [184] The Role of Frequent Point-of-care Molecular Workplace Surveillance for Miners [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04977050?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=3&rank=14>
- [185] The Impact of SARS-CoV-2 Rapid Antigen Testing Kit Screening in Bangkok Community [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT05047900?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=3&rank=16>
- [186] Performance Evaluation of SARS-COV-2 (Covid-19) Antigen Rapid Test [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04808921?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=3&rank=17>
- [187] COVID-19 Diagnostic Self-testing Using Virtual Point-of-care [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04348864?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=3&rank=18>
- [188] Sensitivity of Frequent SARS-CoV-2 (COVID-19) Rapid Antigen Testing Regimen [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04805840?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=3&rank=20>
- [189] Antigen Rapid Test Screening to Prevent SARS-CoV-2 Transmission (COVID-19) at Mass Gathering Events [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04898127?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=4&rank=26>
- [190] Performance Study of SONA Saliva C-19 Rapid Test [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04877002?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=4&rank=27>
- [191] Open Label, Single-Center Study Utilizing BIOZEK COVID-19 Antigen Rapid Test [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04926779?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=4&rank=28>
- [192] Open Label, Single-Center Study Utilizing BIOZEK COVID-19 Antigen Rapid Test [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04805892?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=4&rank=30>
- [193] COVID-19 Antigen Rapid Test Kit - Full Text View - ClinicalTrials.Gov [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04889365?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=5&rank=32>
- [194] Study on Prevention of SARS-CoV-2 Transmission During a Large Indoor Gathering Event - Full Text View - ClinicalTrials.Gov [Internet]. Clinicaltrials.gov. [cited 2021 Nov 4]. Available from:



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- <https://clinicaltrials.gov/ct2/show/NCT04872075?term=rapid+antigen+tests&cond=COVID-19&intr=rapid+antigen+test&draw=5&rank=33>
- [195] Diel R, Nienhaus A. Point-of-care COVID-19 antigen testing in German emergency rooms – a cost-benefit analysis. *Pulmonology*. 2021;. <https://doi.org/10.1016/j.pulmoe.2021.06.009>
- [196] Paltiel AD, Zheng A, Sax PE. Clinical and economic effects of widespread rapid testing to decrease SARS-CoV-2 transmission. *Ann Intern Med*. 2021;174(6):803–10. doi: <https://doi.org/10.7326/M21-0510>
- [197] EVALUATION OF RAPID Ag TEST KITS [Internet]. Gov.ph. [cited 2021 Nov 6]. Available from: <https://ritm.gov.ph/covid-19-kit-evaluation/completed-evaluations/sars-cov-2-rapid-antigen-ag-test/>
- [198] FDA Advisory No.2021-2094 [Internet]. Gov.ph. [cited 2021 Nov 6]. Available from: <https://www.fda.gov.ph/fda-advisory-no-2021-2094-list-of-covid-19-test-kits-with-fda-special-certification-and-performance-validation-conducted-and-or-recommended-by-the-research-institute-for-tropical-medicine-ritm/>
- [199] FDA Advisory No. 2021-2452 [Internet]. Gov.ph. [cited 2021 Nov 6]. Available from: <https://www.fda.gov.ph/fda-advisory-no-2021-2454-caution-on-the-purchase-and-use-of-certain-covid-19-test-kits-following-the-performance-validation-conducted-by-the-ritm/>
- [200] FDA advisory no.2021-1896 [Internet]. Gov.ph. 2021 [cited 2021 Nov 6]. Available from: <https://www.fda.gov.ph/fda-advisory-no-2021-1896-recall-of-covid-19-test-kits-with-low-results-of-performance-validation-conducted-by-the-research-institute-of-tropical-medicine-ritm/>
- [201] FDA advisory no.2021-1338 [Internet]. Gov.ph. 2021 [cited 2021 Nov 6]. Available from: <https://www.fda.gov.ph/fda-advisory-no-2021-1338-delisted-companies-with-issued-fda-special-certification-for-covid-19-test-kits/>
- [202] Department of Health. Price Cap for COVID-19 Rapid Antigen Testing – Asian Preparedness Partnership (APP) [Internet]. Adpc.net. [cited 2021 Nov 5]. Available from: <https://app.adpc.net/resources/price-cap-for-covid-19-rapid-antigen-testing/>
- [203] Department of Health Health Technology Assessment Council. Evidence Summary for Rapid Antigen Test - Recommendations. 29 April 2021. Accessed https://hta.doh.gov.ph/wp-content/uploads/2021/05/2021_Evidence_Summary_for_Rapid_Antigen_Test-Recommendation.pdf on 26 October 2021.
- [204] Hanson KE, Altayar O, Caliendo AM, Arias CA, Englund JA, Hayden MK, Lee MJ, Loeb M, Patel R, El Alayli A, Sultan S, Falck-Ytter Y, Lavergne V, Mansour R, Morgan RL, Murad MH, Patel P, Bhimraj A, Mustafa RA. Infectious Diseases Society of America Guidelines on the Diagnosis of COVID-19: Antigen Testing. *Infectious Diseases Society of America* 2021; Version 1.0.0. Available at <https://www.idsociety.org/practice-guideline/covid-19-guideline-antigen-testing/>. Accessed 5 November 2021.
- [205] World Health Organization. Antigen Detection in the Diagnosis of SARS-CoV-2 infection [Internet]. Who.int. [cited 2021 Nov 5]. Available from: <https://www.who.int/publications/i/item/antigen-detection-in-the-diagnosis-of-sars-cov-2infection-using-rapid-immunoassays>
- [206] American Academy of Pediatrics. COVID-19 Testing Guidance [Internet]. Aap.org. [cited 2021 Nov 5]. Available from: <https://www.aap.org/en/pages/2019-novel-coronavirus-covid-19-infections/clinical-guidance/covid-19-testing-guidance>



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Appendix 1: Evidence to Decision

| FACTORS | | JUDGEMENT | | | | RESEARCH EVIDENCE/ADDITIONAL CONSIDERATIONS | |
|--|--|---|--|---|----------------------|---|--|
| Problem | No | Yes (8) | | | | | |
| Certainty of Evidence | High | Moderate (3) | Low (6) | Very low | | | Overall, the studies included are of low methodological quality due to risk of bias issues and high heterogeneity among included evaluations. |
| Accuracy | Very Accurate | Accurate (7) | Inaccurate (2) | Very Inaccurate | | | Across 164 studies, the pooled sensitivity of RAgTs was found to be moderate at 0.71 (95%CI: 0.68-0.73). Pooled specificity was excellent at 0.995 (95%CI: 0.993-0.996). |
| Values | Important uncertainty or variability (1) | Possibly important uncertainty or variability (7) | Possibly NO important uncertainty or variability (1) | No important uncertainty or variability | | | |
| Resources Required | Uncertain | Large cost (1) | Moderate Cost (6) | Negligible cost | Moderate savings (1) | Large savings | Unit cost of the different brands of rapid antigen test kit in the market ranges from Php 250 to Php 1600. |
| Certainty of evidence of required resources | No included studies (1) | Very low (1) | Low (5) | Moderate (1) | High (1) | | In the Philippines, the Department of Health issued a memorandum dated September 1, 2021 strictly placing a price cap of Php 960 for rapid antigen testing in all testing and clinical laboratories. |
| Cost effectiveness | No included studies (6) | Favors RT-PCR (1) | Does not favor either RAgT or RT-PCR | Favors RAgT (2) | | | No local economic evaluation studies are available as of press time on comparing rapid antigen tests and RT-PCR. |
| Equity | Uncertain (4) | Reduced | Probably no impact | Increased (5) | | | |
| Acceptability | Uncertain (1) | No (1) | Yes (7) | | | | |
| Feasibility | Uncertain | No | Yes (9) | | | | |



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Appendix 2: Search Yield and Results

| Search | Query | Results | Time |
|--------|---|------------|----------|
| #10 | Search #1 AND #8 Filters: from 1000/1/1 - 2021/9/30 Sort by: Most Recent | 802 | 04:57:47 |
| #9 | Search #1 AND #8 Sort by: Most Recent | 874 | 04:56:30 |
| #8 | Search #7 OR #2 Sort by: Most Recent | 18,609 | 04:56:20 |
| #7 | Search #5 OR #6 Sort by: Most Recent | 18,608 | 04:56:13 |
| #6 | Search: rapid antigen test* OR “rapid antigen detection test” OR radt OR radts OR rdt OR rdts OR (antigen* n3 detect*) Sort by: Most Recent | 18,555 | 04:56:06 |
| #5 | Search #3 and #4 Sort by: Most Recent | 171 | 04:55:58 |
| #4 | Search: (test OR tests OR detect* OR diagnos* OR kit OR kits OR assay*) Sort by: Most Recent | 11,628,285 | 04:55:41 |
| #3 | Search: ((rapid OR point-of-care OR “point of care” OR poc OR popt) n3 antigen)) Sort by: Most Recent | 204 | 04:55:27 |
| #2 | Search: “COVID-19 Ag Respi-Strip” OR “BIOCREDIT COVID-19 Ag” OR “STANDARD F COVID-19 Ag” OR “STANDARD Q COVID-19 Ag” OR “Bioeasy 2019-nCoV Ag” Sort by: Most Recent | 25 | 04:55:19 |
| #1 | Search: (“Coronavirus Infections”[Mesh] OR novel coronavirus OR NCOV OR “COVID-19”[Supplementary Concept] OR covid19 OR covid 19 OR covid-19 OR “severe acute respiratory syndrome coronavirus 2”[Supplementary Concept] OR severe acute respiratory syndrome coronavirus 2 OR SARS2 OR SARS 2 OR SARS COV2 OR SARS COV 2 OR SARS-COV-2) Sort by: Most Recent | 205,257 | 04:55:11 |



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Appendix 3. Characteristics of Included Studies

| Study | | Setting | RAgT Brand | Test Specimen | Population | Sample Size | Reference Standard | |
|-------|--------------------|-------------|---|---------------|---|-------------|--------------------|----------|
| | | | | | | | Test | Specimen |
| 1 | Okoye 2021 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | NP | Asymptomatic university students | 2638 | RT PCR | Nasal |
| 2 | Oh 2021 | Korea | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Patients admitted in the study hospital | 118 | RT PCR | NP |
| 3 | Lindner 202105 | Germany | STANDARD Q COVID19 Ag (SD Biosensor) | MT + NP | Symptomatic patients at ambulatory testing facility | 146 | RT PCR | NP + OP |
| 4 | Lindner 202104 | Germany | STANDARD Q COVID19 Ag (SD Biosensor) | MT | Symptomatic patients at ambulatory testing facility | 287 | RT PCR | NP + OP |
| 5 | Ishii 2021 | Japan | ESPLINE SARS-CoV-2 (Fujirebio) | NP + Saliva | COVID and non-COVID patients admitted | 271 | RT PCR | NP |
| 6 | Baro 2021 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Unexposed asymptomatics during third wave in Spain | 286 | RT PCR | NP |
| 7 | Stokes 2021 | Canada | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Symptomatic individuals | 1786 | RT PCR | NP |
| 8 | Torres 202101 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Asymptomatic close contacts | 634 | RT PCR | NP |
| 9 | Torres 202102 | Spain | CLINITEST Rapid COVID-19 Antigen Test (Siemens) | NP | Symptomatic individuals and asymptomatic close contacts | 270 | RT PCR | NP |
| 10 | Torres 202102 | Netherlands | COVID-19 Rapid Antigen Test (BD Veritor) | NP | Non-hospitalized symptomatic patients | 351 | RT PCR | NP |
| 11 | Yokota 2021 | Japan | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | NP + Saliva | Inpatients and Outpatients | 343 | RT PCR | NP |
| 12 | Igloi 2021 | Netherlands | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | NP | Symptomatic or close contact | 970 | RT PCR | NP + OP |
| 13 | Courtellemont 2021 | France | COVID-VIRO Antigen Rapid Test (AAZ) | NP | Asymptomatic and hospitalized | 248 | RT PCR | NP |
| 14 | Olearo 2021 | Germany | STANDARD Q COVID19 Ag (SD Biosensor) | NP + OP | Asymptomatic and hospitalized | 184 | RT PCR | NP + OP |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | | | |
| | | | SARS-CoV-2 Antigen Rapid Test (MedSan GmbH) | | | | | |
| | | | CLINITEST Rapid COVID-19 Antigen Test (Siemens) | | | | | |
| 15 | Jaaskelainen 2021 | Finland | Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | NP | Symptomatic | 188 | RT-PCR | NP |



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| | | | | | | | | |
|----|---------------------|------------|---|-------|---|-------|--------|------------|
| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | 198 | | |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | 190 | | |
| 16 | Adnan 2021 | Bangladesh | Rapid In-house ELISA (Bangladesh) | NP | No data | 339 | RT-PCR | NP |
| 17 | Pollock 2021a | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | Nasal | Symptomatic patients | 2,482 | RT-PCR | Nasal |
| 18 | Pollock 2021b | USA | CareStart COVID-19 Antigen Test (Access Bio) | Nasal | Symptomatic patients | 1,603 | RT-PCR | Nasal |
| 19 | Peña-Rodríguez 2021 | Mexico | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Symptomatic, asymptomatic, exposed contacts (3-5 days) | 369 | RT-PCR | NP + OP |
| 20 | Chiu 2021 | USA | INDICAID COVID-19 rapid antigen test (PHASE) | Nasal | Symptomatic participants | 698 | RT PCR | Nasal |
| | | HongKong | | | Asymptomatic participants | 22994 | RT PCR | Nasal + OP |
| 21 | Shaikh 2021 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | Nasal | Symptomatic children | 199 | RT PCR | Nasal |
| 22 | Laandas 2021 | Norway | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Community and outbreak setting | 4,857 | RT-PCR | NP |
| 23 | Boum 2021 | Cameroon | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Hospitalized, asymptomatic volunteers, exposed contacts | 1,195 | RT-PCR | NP |
| 24 | Pickering 2021 | UK | Innova Rapid SARS-CoV-2 Antigen Test (Xiamen Biotime Biotechnology) | NP | Trained personnel; community settings, inpatients and outpatients | 200 | RT PCR | NP |
| | | | Spring Healthcare SARS-CoV-2 Antigen Rapid Test Cassette (Shanghai ZJ Bio-Tech) | | | | | |
| | | | E25Bio Rapid Diagnostic Test (E25Bio) | | | | | |
| | | | Encode SARS-CoV-2 Antigen Rapid Test Device (Zhuhai Encode Medical Engineering) | | | | | |
| | | | COVID-19 Coronavirus Rapid Antigen Test Cassette (Surescreen) | | | | | |
| | | | COVID-19 Coronavirus Rapid Antigen Test Cassette (Surescreen) | | | | | |
| 25 | Sood 2021 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | Nasal | Children less than 18 years | 783 | RT-PCR | OP |



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| 26 | Matsuda 2021 | Mexico | Ecotest COVID-19 Rapid Antigen Test (Verify) | NP | Symptomatic patients | 112 | RT-PCR | NP + OP |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | | | |
| 27 | Bello-Chavolla 2021 | Mexico | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Community setting: COVID suspects | 193824 | RT-PCR | NP |
| 28 | Kyritsi 2021 | Greece | Rapid Test Ag 2019-nCoV (PROGNOSIS, BIOTECH) | NP | Hospital setting | 624 | RT-PCR | NP |
| 29 | Iqbal 2021 | Pakistan | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | NP | Hospital setting: COVID suspects | 170 | RT-PCR | NP |
| 30 | Nikolai 2021 | Germany | STANDARD Q COVID19 Ag (SD Biosensor) | Nasal + MT | Community setting: COVID suspects from ambulatory facility | 228 | RT-PCR | NP |
| 31 | Caruana 2021 | Switzerland | Exdia COVID-19 Antigen Test (Precision-Bio) | NP | Symptomatic and asymptomatic ER patients | 532 | RT-PCR | NP |
| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | | | |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | | | |
| | | | COVID-19 Rapid Antigen Test (BD Veritor) | | | | | |
| 32 | Martin-Sanchez 2021 | Spain | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | NP | Community setting: population-based screening of asymptomatic individuals | 881 | RT-PCR | NP |
| 33 | Maniscalco 2021 | Italy | Inflamcheck device (Exhalation technology LTD) | Exhaled breath condensate | Hospital setting: COVID suspects, convalescent patients, asymptomatic with high risk of COVID, asymptomatic with low risk of COVID | 105 | RT-PCR | NP |
| 34 | Merino-Amador 2021 | Spain | CLINITEST Rapid COVID-19 Antigen Test (Siemens) | NP | Hospital setting: COVID suspects with exposure < 7 days or early symptoms | 450 | RT-PCR | NP |
| 35 | Hauser 2021 | Germany | LIAISON SARS-CoV-2 Antigen Assay (DiaSorin) | NP | Hospitalized patients | 196 | RT-PCR | NP |
| 36 | Osterman 2021 | Germany | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | NP | Hospital setting: patients from ER or clinics | 410 | RT-PCR | NP |
| | | | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | | | | | |
| | | | LIAISON SARS-CoV-2 Antigen Assay (DiaSorin) | | | | | |



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| | | | SARS-CoV-2 Ag ELISA (Euroimmun) | | | | | |
| 37 | Andreani 2021 | France | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Community settings: patients and personnels in a diagnostic facility | 239 | RT-PCR | NP |
| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | | | |
| | | | Certest SARS-CoV-2 one step card test (Theradiag) | | | | | |
| | | | Orient Gene Coronavirus Ag rapid cassette (Menarini) | | | | | |
| | | | Espline SARS-CoV-2 (Fujirebio) | | | | | |
| | | | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | | | | | |
| 38 | Salvagno 2021 | Italy | Espline SARS-CoV-2 (Fujirebio) | NP | Community settings: patients with confirmed COVID | 174 | RT-PCR | NP |
| 39 | Lhuillier 2021 | Switzerland | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Community and hospital setting: Children | 822 | RT-PCR | NP |
| 40 | Korenkov 2021 | Germany | STANDARD Q COVID19 Ag (SD Biosensor) | NP + OP | Hospital setting: COVID suspect and Trained personnel screening | 2028 | RT-PCR | NP + OP |
| 41 | Bachman 2021 | USA | Open-access lateral flow assay | Anterior nares | Community settings: COVID suspects in clinics | 170 | RT PCR | Anterior nares + NP |
| | | | Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | | | | | |
| | | | BinaxNOW COVID-19 Antigen Card (Abbott) | | | | | |
| | | | Meso Scale Discovery MESO Quick-Plex SQ 120 (MSD) | | | | | |
| | | | Open-access lateral flow assay | | | | | |
| | | | Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | | | | | |
| | | | BinaxNOW COVID-19 Antigen Card (Abbott) | | | | | |
| | | | Meso Scale Discovery MESO Quick-Plex SQ 120 (MSD) | | | | | |
| 42 | Nomoto 2021 | Japan | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | NP | Hospital setting: inpatients | 100 | RT-PCR | NP |



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| 43 | Kruger 2021 | Germany | LumiraDx SARS-CoV-2 Antigen (LumiraDx) | MT | Community settings: COVID suspects in Drive-in testing site and ambulatory care facility | 761 | RT-PCR | NP |
| 44 | Kahn 2021 | Germany | Standard F Covid19 Ag FIA (SD Biosensor) | OP | Community settings; Individuals at point-of-care facilities | 3110 | RT-PCR | OP |
| 45 | Escriva 2021 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Community settings: nursing home residents and employees | 448 | RT-PCR | NP |
| 46 | Norz 2021 | Germany | Elecsys SARS-CoV-2 Antigen assay (Roche) | NP + OP | Hospital settings: multicenter routine diagnostic | 3139 | RT-PCR | NP and OP |
| 47 | Mboma 2021 | Germany | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Hospitalized patients and accompanying relatives | 3,686 | RT-PCR | NP |
| 48 | Stohr 2021 | The Netherlands | COVID-19 Rapid Antigen Test (BD Veritor) | NP | Community settings | 3,201 | RT PCR | NP and OP |
| | | | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | | | | | |
| 49 | Allan-Blitz 2021 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | AN + OP | Community setting | 15,304 | RT-PCR | NP + OP + AN |
| 50 | Cassuto 2021 | France | COVID-VIRO Antigen Rapid Test (AAZ) | NP | Symptomatic adult volunteers | 234 | RT-PCR | NP |
| 51 | Agarwal 2021 | India | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Hospitalized patients and asymptomatic contacts | 467 | RT-PCR | NP |
| 52 | Hartard 2021 | France | LIAISON SARS-CoV-2 Antigen Assay (DiaSorin) | NP | Mass screening of hospitalized patients | 378 | RT-PCR | NP |
| 53 | Kobayashi 2021 | Japan | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | NP + saliva | Samples from hospitals and public health centers in Japan | 10,422 | RT-PCR | NP |
| 54 | Wagenhauser 2021 | Germany | NADAL COVID-19 Rapid Antigen Test (Nal Von Minden) | OP | Mass screening of all hospitalized patients | 5,056 | RT-PCR | OP |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | | | |
| | | | SARS-CoV-2 Antigen Rapid Test (MedSan GmbH) | | | | | |
| 55 | Amer 2021 | Egypt | STANDARD Q COVID19 Ag (SD Biosensor) | NP + OP | Community and hospital setting: COVID suspects Trained personnels | 83 | RT-PCR | NP + OP |
| 56 | Montero 2021 | Spain | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | NP + OP | Community setting: asymptomatic adults in a semi-closed community | 2543 | RT-PCR | NP + OP |



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| 57 | Ferte 2021 | France | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Community setting: symptomatic and asymptomatic students | 688 | RT-PCR | NP |
| 58 | Leixner 2021 | Austria | AMP Rapid Test SARS-CoV-2 Ag (AMP) | NP | Hospital setting: symptomatic patients presenting at the ER | 392 | RT-PCR | NP + OP |
| 59 | Soleimani 2021 | Belgium | BioSpeedia COVID19Speed-Antigen Test (Institut Pasteur) | NP | Community settings: COVID suspects or with exposure | 401 | RT-PCR | NP |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | | | |
| 60 | Kiro 2021 | India | Standard F Covid19 Ag FIA (SD Biosensor) | NP | Hospital setting: admitted patients and OPD patients from COVID screening | 354 | RT-PCR | NP |
| 61 | Garcia 2021 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Community settings | 356 | RT-PCR | NP |
| | | | Standard F Covid19 Ag FIA (SD Biosensor) | | | | | |
| 62 | Seynaeve 2021 | Belgium | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP | Hospitalized patients | 100 | RT PCR | NP |
| | | | Coronavirus Ag Rapid Test Cassette (Healgen Scientific) | | | | | |
| 63 | Nsoga 2021 | Nsoga | Panbio COVID-19 Ag Rapid Test (Abbott) | OP | Hospitalized patients | 402 | RT-PCR | NP |
| 64 | Eleftheriou 2021 | Greece | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | hospitalized pediatric patients (< 16 years old) symptomatic and high risk for infection based on regional epidemiologic data | 744 | RT-PCR | NP |
| 65 | Fiñana 2021 | UK | Innova Rapid SARS-CoV-2 Antigen Test (Xiamen Biotime Biotechnology) | NP + OP | Adults >18 years old; asymptomatic: community setting | 5504 | RT-PCR | NP + OP |
| 66 | Kurihara 2021 | Japan | Quick Chaser Auto SARS-CoV-2 (Mizuhomedy) | NP | community: individuals that possibly contracted SARS-Cov2; referred by clinics, health centers and healthcare workers for testing | 1401 | RT PCR | NP |
| 67 | Ilko 2021 | Slovenia | NADAL COVID-19 Rapid Antigen Test (Nal Von Minden) | NP | Long-term healthcare facility: Elderly 65 - 95 yrs old with at least one symptom of COVID | 125 | RT-PCR | NP |



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| 68 | Karon 2021 | USA | COVID-19 Rapid Antigen Test (BD Veritor) | NP | residual phosphate buffered saline samples of symptomatic and asymptomatic patients | 347 | RT-PCR | NP |
| 69 | Abusrewil 2021 | Libya | 10 brands: Fluorecare, Espline, Rapigen, Assut, Orient Gene, AMP, Acon, Abbot, Certest Biotec, Bioperfectus | NP | community setting: clinical features suggestive of COVID-19 or a history of close contact with COVID-19 positive patients | 231 | RT-PCR | NP |
| 70 | Orsi 2021 | Italy | FREND Antigen Test (NanoEntek) | NP | Hospitalized patients | 110 | RT-PCR | NP |
| 71 | Brihn 2021 | USA | Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | Anterior Nares | Symptomatic and asymptomatic patients | 2039 | RT-PCR | NP |
| 72 | Osmanodja 2021 | Germany | Dräger Antigen Test SARS-CoV-2 (Dräger) | Anterior Nares | Community setting | 379 | RT-PCR | NP + OP |
| 73 | Bornemann 2021 | Germany | Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | NP | Community setting | 1404 | RT-PCR | NP |
| 74 | Holzner 2021 | Germany | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Community setting | 2375 | RT-PCR | NP |
| 75 | Pena 2021 | Chile | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Community setting | 842 | RT-PCR | NP |
| 76 | Donapetry 2021 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Children | 440 | RT-PCR | NP |
| 77 | Cento 2021 | Italy | LumiraDx™ SARS-CoV-2 Antigen Test | NP | Community setting | 960 | RT-PCR | NP |
| 78 | Klein 2021 | Germany | Panbio COVID-19 Ag Rapid Test (Abbott) | NP + MT | community: symptomatic adults and high risk contact of confirmed case in SARS-Cov-2-drive in testing center: | 290 | RT-PCR | NP |
| 79 | Frediani 2021 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | Anterior Nares | community: community-based and hospital based testing center: age > 7 yrs old (adults and pedia) symptomatic within 7 days from onset | 341 | RT-PCR | NP |
| 80 | Schuit 2021 | Netherlands | COVID-19 Rapid Antigen Test (BD Veritor) | NP + OP | Close contacts, asymptomatic individuals requesting for a COVID-19 test | 2678 | RT-PCR | NP + OP |
| | | | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | | | | | |
| 81 | Wachinger | Germany | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Asymptomatic patients in hospitals, presenting for elective surgeries, OPD | 1596 | RT-PCR | Unspecified |



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| 82 | Fourati 2021 | France | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP | COVID-19 patients | 626 | RT-PCR | NP |
| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | | | |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | | | |
| | | | COVID-VIRO Antigen Rapid Test (AAZ) | | | | | |
| | | | NG-test SARS-CoV-2 Ag (NG-Biotech) | | | | | |
| | | | BIOSYNEX COVID-19 Ag BSS- (Biosynex SA) | | | | | |
| 83 | Leli 2021 | Italy | LumiraDx™ SARS-CoV-2 Antigen Test | Nasal | Hospital (presenting in emergency department) | 792 | RT-PCR | NP |
| 84 | Sahuquillo 2021 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Hospital setting: pediatric ER; Symptomatic children aged 0-14 yrs | 357 | RT-PCR | NP |
| 85 | Kanauja 2021 | India | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP | hospital setting (adult communicable disease ward); symptomatic adults and asymptomatic contacts | 484 | RT-PCR | NP |
| 86 | Dierks 2021 | Germany | LumiraDx™ SARS-CoV-2 Antigen Test | NP | Hospital setting: employee of University Medical Center | 444 | RT-PCR | NP |
| 87 | Jung 2021 | France | BIOSYNEX COVID-19 Ag BSS (Biosynex SA) | NP | hospital setting: ER & primary pediatric care center; age < 18 | 308 | RT-PCR | NP |
| 88 | Tsai 2021 | Taiwan | Vstrip COVID-19 Antigen Rapid Test (Panion & BF Biotech) | NP | Hospital setting: COVID suspects but no mention whether symptomatic or asymptomatic | 61 | RT-PCR | NP |
| 89 | Jegerlehner 2021 | Switzerland | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | NP | community setting: COVID testing facility; adults>18 yrs symptomatic and those suspected of exposure to infected | 1462 | RT-PCR | NP |
| 90 | Thakur 2021 | India | Immune-chromatographic lateral flow assay PathoCatch/ACCUCARE (Lab Care Diagnostics) | NP | Community settings (Preoperative) | 677 | RT-PCR | NP + OP |
| 91 | Kim 2021 | South Korea | | NP | | 130 | RT-PCR | NP |



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| | | India | GenBody COVAG025 Rapid Antigen Test | | Hospital setting: patients visited or admitted in the hospital | 200 | | |
| 92 | Bruzzone 2021 | Italy | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Hospital setting: retrospective analysis of swab samples in laboratory | 321 | RT-PCR | NP |
| | | | Humasis COVID-Ag Test COVID-19 Antigen (Humasis) | | | | | |
| | | | Rapid Test Prima Professional (Prima Lab) | | | | | |
| | | | BIOCREDIT COVID-19 Ag test (RapiGEN, Inc) | | | | | |
| | | | Standard F Covid19 Ag FIA (SD Biosensor) | | | | | |
| | | | LumiraDx™ SARS-CoV-2 Antigen Test | | | | | |
| | | | FREND Antigen Test (NanoEntek) | | | | | |
| 93 | Nordgren 2021 | Sweden | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Community setting: Symptomatic individuals | 332 | RT-PCR | NP |
| | | | Coronavirus Ag Rapid Test Cassette (Healgen Scientific) | | | | | |
| 94 | Bianco 2021 | Italy | LumiraDx™ SARS-CoV-2 Antigen Test | Nasal | Hospital setting: patients in adult and pedia ER and Trained personnels | 907 | RT-PCR | NP |
| 95 | Kolwijck 2021 | Netherlands | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Hospital setting: Trained personnel at least 16 years old | 433 | RT-PCR | NP and OP |
| 96 | Ford 2021 | USA | Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | MT | Community setting: students, faculty and employee of University | 1051 | RT-PCR | MT |
| 97 | Kweon 2021 | Korea | AFIAS COVID-19 Ag (AFC) (Menarini; Florence, Italy) | NP | Hospital setting: Mixed COVID19 and non-COVID19 patients | 322 | RT-PCR | NP |
| | | | ichroma™ COVID-19 Ag (Boditech) | | | | | |
| 98 | Berger 2021 | Switzerland | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Hospital setting: COVID suspects at least 16 years old consulting in the hospital | 535 | RT-PCR | NP |
| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | 529 | | |
| 99 | Van Honacker 2021 | Belgium | BIOSYNEX COVID-19 Ag BSS (Biosynex SA) | NP | Hospital setting: COVID suspect and non-COVID with other respiratory isolates | 100 | RT-PCR | NP |
| | | | Biotical SARS-CoV-2 Ag Card (Biotical Health) | | | 100 | | |



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| | | | Coronavirus AG Rapid test cassette (Zhejiang Orient Gene Biotech Co., Zhejiang, China) | | | 98 | | |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | 97 | | |
| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | 98 | | |
| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | 4207 | | |
| 100 | Bouassa 2021 | France | SIENNA COVID-19 Antigen Rapid Test Cassette | NP | Hospital setting: retrospective study of archived NP swabs | 150 | RT-PCR | NP |
| 101 | Caruana 2021 | Switzerland | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Hospital setting: asymptomatic adults hospitalized in medical and surgical wards | 116 | RT-PCR | NP |
| 102 | Baccani 2021 | Italy | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | NP | Hospital setting: inpatients in 3 Tuscan hospitals | 201 | RT-PCR | NP |
| | | | Standard F Covid19 Ag FIA (SD Biosensor) | | | 93 | | |
| | | | AFIAS COVID-19 Ag (AFC) (Menarini; Florence, Italy) | | | 81 | | |
| 103 | Koeleman 2021 | Netherlands | Certest SARS-CoV-2 (Certest Biotec S.L., Spain) | NP and OP | Hospital setting: symptomatic ER patients, nursing home residents and Trained personnels | 80 | RT-PCR | NP and OP |
| | | | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | | | 80 | | |
| | | | Romed Coronavirus Ag Rapid Test (Romed, The Netherlands) | | | 80 | | |
| | | | Romed Coronavirus Ag Rapid Test (Romed, The Netherlands) | | | 900 | | |
| 104 | Kruger 2021 | Germany | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Community setting: patients in drive-in testing site and ambulatory testing facility | 1108 | RT-PCR | NP and OP |
| 105 | Denina 2021 | Italy | LumiraDx™ SARS-CoV-2 Antigen Test | Nasal | Hospital setting: children admitted at the ER and hospital | 191 | RT-PCR | NP |
| 107 | Stokes 202109 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | Anterior nares | Hospital setting: Asymptomatic pre-surgery | 997 | RT-PCR | NP |



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| 107 | Halfon 2021 | France | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Community setting: patients screened in the laboratory | 200 | RT-PCR | NP |
| 108 | Paul 2021 | India | VITROS Immunodiagnostic Products SARS-CoV-2 Antigen test (Ortho Clinical Diagnostics) | NP + OP | Hospital setting: symptomatic patients in OPD | 148 | RT-PCR | NP and OP |
| | | | Angcard® COVID-19 rapid Antigen Test kit (Angstrom® Biotech Pvt. Ltd., Alwar, Rajasthan, India) | NP | | | | |
| 109 | Von Ahnen 2021 | Germany | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | NP | Hospital setting: screening of all employees | 919 | RT-PCR | NP |
| 110 | Shah 2021 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | Anterior nares | Community setting: patients recruited from community testing site | 2110 | RT-PCR | Anterior nares |
| 111 | McKay 2021 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | Anterior Nares | Community setting: residents and employees of nursing home | 532 | RT-PCR | Mixed |
| 112 | Abdelrazik 2021 | Egypt | BIOCREDIT COVID-19 Ag test (RapiGEN, Inc) | NP | Patients and HCW, patient contacts | 310 | RT-PCR | NP |
| 113 | Albert 2020 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP or OP | COVID 19 test centre | 497 | RT-PCR | NP |
| 114 | Aleman 2020 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP or OP | Symptomatic patients, close contacts, preventive screening of unexposed asymptomatics | 1406 | RT-PCR | NP or MT |
| 115 | Asai 2021 | Japan | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | Saliva | Suspected patients | 305 | RT-PCR | Saliva |
| 116 | Billaud 2020 | France | Panbio COVID-19 Ag Rapid Test (Abbott) | NP or OP | Contacts | 462 | RT-PCR | NP or OP |
| 117 | Blairon 2020 | Belgium | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP or OP | Laboratory-based | 56 | RT-PCR | NP |
| 118 | Bulilete 2021 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP | Symptomatic patients and close contacts | 1369 | RT-PCR | NP |
| 119 | Caputo 2021 | Italy | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | NP | Clinical suspect, screening or contacts | 4266 | RT-PCR | NP |
| 120 | Cerutti 2020 | Italy | STANDARD Q COVID19 Ag (SD Biosensor) | NP | Mixed | 256 | RT-PCR | NP |
| 121 | Chaimayo 2020 | Thailand | STANDARD Q COVID19 Ag (SD Biosensor) | NP + OP | Covid-19 suspects | 454 | RT-PCR | NP + OP |
| 122 | Ciotti 2021 | Italy | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP | Covid-19 suspects | 50 | RT-PCR | NP |



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| 123 | Diao 2020 | China | 2019-nCoV Ag Fluorescence Rapid Test Kit (Bioeasy) | NP or OP | Unclear | 239 | RT-PCR | NP |
| 124 | Favresse 2021 | Belgium | Biotical SARS-CoV-2 Ag Card (Biotical Health) | NP | Symptomatic and asymptomatic subjects | 188 | RT-PCR | NP |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | | | |
| | | | Coronavirus Ag Rapid Test Cassette (Healgen Scientific) | | | | | |
| | | | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | | | | | |
| | | | VITROS Immunodiagnostic Products SARS-CoV-2 Antigen test (Ortho Clinical Diagnostics) | | | | | |
| 125 | Fenollar 2020 | France | Panbio COVID-19 Ag Rapid Test (Abbott) | NP or OP | Symptomatic patients | 182 | RT-PCR | NP |
| | | | | | Asymptomatic contacts | 159 | | |
| 126 | Fourati 2020 | France | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP or OP | Laboratory-based | 629 | RT-PCR | NP |
| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | 628 | | |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | 632 | | |
| | | | BIOSYNEX COVID-19 Ag BSS (Biosynex SA) | | | 634 | | |
| | | | COVID-VIRO Antigen Rapid Test (AAZ) | | | 632 | | |
| 127 | Gili 2021 | Italy | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | NP | Screening population | 1340 | RT-PCR | NP |
| 128 | Gremmels 2020 | Netherlands | Panbio COVID-19 Ag Rapid Test (Abbott) | NP or OP | COVID-19 test centre | 1367 | RT-PCR | NP |
| 129 | Gupta 2020 | India | STANDARD Q COVID19 Ag (SD Biosensor) | NP or OP | COVID-19 test centre | 330 | RT-PCR | NP + OP |
| 130 | Herrera 2020 | USA | Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | NP | Symptomatic and asymptomatic healthcare workers | 1172 | RT-PCR | NP |
| 131 | Hirotsu 2020 | Japan | Lumipulse SARS-CoV-2 Antigen Kit (Fujirebio) | NP | Hospital setting: infected and noninfected patients | 313 | RT-PCR | NP |
| 132 | Homza 2021 | Czech Republic | SARS-CoV-2 Antigen Rapid Test Kit (JOYSBIO [Tianjin Biotechnology Co., China]) | NP | COVID-19 test centre | 225 | RT-PCR, viral culture | NP |
| | | | Ecotest COVID-19 Antigen Rapid Test (Assure Tech, Hangzhou, China) | | | 179 | | |



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| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | 72 | | |
| | | | Immupass VivaDiag SARS-CoV-2 Ag Rapid Test (VivaChek Biotech [Hangzhou] Co., China) | | | 268 | | |
| | | | ND COVID-19 Ag test (NDFOS, Eumseong, Korea) | | | 91 | | |
| 133 | James 2021 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | NP | Symptomatic and asymptomatic patients | 2339 | RT-PCR | Nasal swab |
| 134 | Kruger 2020 | Germany | 2019-nCoV Ag Fluorescence Rapid Test Kit (Bioeasy) | NP + OP | COVID-19 test centre | 712 | RT-PCR | NP + OP |
| | | | COVID-19 Ag Respi-Strip (Coris Bioconcept) | | | 409 | | |
| | | | STANDARD Q COVID19 Ag (SD Biosensor) | | | 1216 | | |
| 135 | Kruttgen 2020 | Germany | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | NP | Admitted patients in the hospital | 150 | RT-PCR | NP |
| 136 | Lambert-Niclot 2020 | France | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP or OP | Laboratory-based | 138 | RT-PCR | NP |
| 137 | Linares 2020 | Spain | Panbio COVID-19 Ag Rapid Test (Abbott) | NP or OP | Hospital A&E | 255 | RT-PCR | NP |
| 138 | Liotti 2020 | Italy | Standard F Covid19 Ag FIA (SD Biosensor) | NP or OP | Laboratory-based | 359 | RT-PCR | NP |
| 139 | Mak 2020 | China | BIOCREDIT COVID-19 Ag test (RapiGEN, Inc) | Mixed respiratory samples | COVID-confirmed patients | 368 | RT-PCR | and respiratory sam |
| 140 | Mertens 2020 | Belgium | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP or OP | Laboratory-based | 328 | RT-PCR | NP |
| 141 | Mockel 2021 | Germany | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | NP + OP | COVID19 suspect adults in the emergency department | 271 | RT-PCR | NP |
| 142 | Muhi 2021 | Australia | Panbio COVID-19 Ag Rapid Test (Abbott) | Nasal | Hospital | 2602 | RT-PCR + Viral culture | NP + OP |
| 143 | Nagura-Ikeda 2020 | Japan | Espline SARS-CoV-2 (Fujirebio) | Saliva | Mixed | 103 | RT-PCR | Saliva |
| 144 | Nalumansi 2020 | Uganda | STANDARD Q COVID19 Ag (SD Biosensor) | NP | COVID-19 suspects and low-risk volunteers | 262 | RT-PCR | NP |
| 145 | Nash 2020 | USA | E25Bio Rapid Diagnostic Test (E25Bio) | NP or OP | Laboratory-based | 190 | RT-PCR | NP + OP |
| 146 | Osterman 022021 | Germany | STANDARD Q COVID19 Ag (SD Biosensor) | NP + OP | Symptomatic patients in the ER | 681 | RT-PCR | NP + OP |
| | | | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | | | 771 | | |



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| | | | | | | | | |
|-----|--------------------|---------|--|----------------|--|------|--------|----------------|
| 147 | Porte 2020a | Chile | 2019-nCoV Ag Fluorescence Rapid Test Kit (Bioeasy) | NP + OP | Hospital A&E | 127 | RT-PCR | NP + OP |
| 148 | Porte 2020b | Chile | Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | NP + OP | COVID-19 test centre | 64 | RT-PCR | NP + OP |
| | | | Standard F Covid19 Ag FIA (SD Biosensor) | NP + OP | COVID-19 test centre | 64 | RT-PCR | |
| 149 | Pray 2021 | USA | Sofia SARS Antigen Fluorescent Immunoassay (Quidel) | Anterior Nares | students, faculty & staff of university campus | 1098 | RT-PCR | Anterior Nares |
| 150 | Prince-Guerra 2021 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | Anterior Nares | 10 years and older receiving testing for COVID 19 | 4904 | RT-PCR | NP |
| 151 | Ristic 2021 | Serbia | STANDARD Q COVID19 Ag (SD Biosensor) | NP | triage ambulance of primary and tertiary outpatients healthcare facility; 14 - 91 yrs old | 120 | RT-PCR | NP |
| 152 | Rottenstreich 2021 | Israel | NowCheck COVID-19 Ag Test (Bionote) | NP | Asymptomatic women admitted for delivery | 1326 | RT-PCR | NP |
| 153 | Schildgen 2020 | Germany | BIOCREDIT COVID-19 Ag test (RapiGEN, Inc) | Mixed | Laboratory based: specimens with confirmed PCR results | 73 | RT-PCR | Mixed |
| | | | Panbio COVID-19 Ag Rapid Test (Abbott) | | | | | |
| | | | Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics) | | | | | |
| 154 | Scohy 2020 | Belgium | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP | Laboratory-based: NP specimen for COVID testing with available data on symptomatology of some specimen | 148 | RT-PCR | NP |
| 155 | Seitz 2021 | Austria | COVID- 19 Antigen Test Cassette [hypersensitive colloidal gold] (Xiamen Zhongsheng Langjie Biotechnology Co., Ltd) | Saliva | healthy citizens of Vienna, Austria were invited to participate in a voluntary SARS-CoV-2 mass screening | 40 | RT-PCR | gargle sample |
| 156 | Shrestha 2020 | Nepal | BIOCREDIT COVID-19 Ag test (RapiGEN, Inc) | NP or OP | close contacts of confirmed COVID 19 | 113 | RT-PCR | NP |
| 157 | Takeda 2020 | Japan | Espline SARS-CoV-2 (Fujirebio) | NP or OP | Laboratory-based: specimens for COVID testing | 162 | RT-PCR | NP |
| 158 | Turcato 2020 | Italy | STANDARD Q COVID19 Ag (SD Biosensor) | NP | ER consults (COVID related and non-COVID complains) and contacts of COVID infected persons | 3410 | RT-PCR | NP |



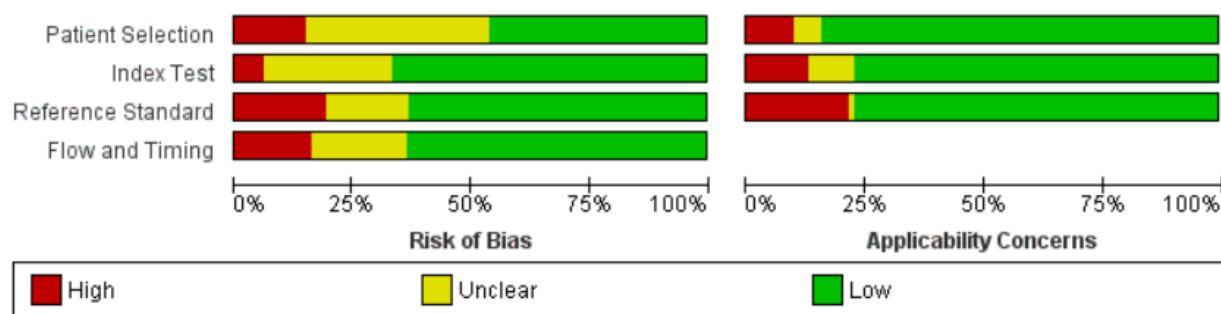
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| | | | | | | | | |
|-----|-----------------|----------------|---|----------------|---|------|--------|----------------|
| 159 | Uwamino 2021 | Japan | Espline SARS-CoV-2 (Fujirebio) | NP and saliva | Adults for COVID testing | 117 | RT-PCR | NP and saliva |
| 160 | Veyrenche 2020 | France | COVID-19 Ag Respi-Strip (Coris Bioconcept) | NP or OP | Inpatients, symptomatic adults and healthcare worker | 65 | RT-PCR | NP |
| 161 | Weitzel 2020 | Chile | BIOCREREDIT COVID-19 Ag test (RapiGEN, Inc) | NP + OP | symptomatic individuals | 109 | RT-PCR | NP + OP |
| | | | StrongStep Rapid Antigen Test (Liming) | | | 19 | | |
| | | | Huaketai SARS-CoV-2 Rapid Antigen Test (Savant) | | | 109 | | |
| | | | 2019-nCoV Ag (Shenzhen Bioeasy Biotech) | | | 111 | | |
| | | | COVID-19 Rapid Antigen Test (BD Veritor) | | | 251 | | |
| 162 | Young 2020 | USA | COVID-19 Rapid Antigen Test (BD Veritor) | NP or OP | 18 years old and above with symptoms | 251 | RT-PCR | NP or OP |
| 163 | Pilarowski 2020 | USA | BinaxNOW COVID-19 Antigen Card (Abbott) | Anterior Nares | symptomatic and asymptomatic individuals | 3302 | RT-PCR | Anterior Nares |
| 164 | PHE 2020 | United Kingdom | Innova Rapid SARS-CoV-2 Antigen Test (Xiamen Biotime Biotechnology) | NP + OP | Hospital in-patient; contacts; patients in COVID-19 test center | 3168 | RT-PCR | NP + OP |

RT PCR: Reverse Transcriptase – Polymerase Chain Reaction; NP: Nasopharyngeal; OP: Oropharyngeal.



Appendix 4. Risk of Bias and Applicability Concerns of Included Studies





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Appendix 5. Characteristics of Ongoing Studies

| NCT Number | Title | Population | Interventions | Comparator | Outcomes |
|-------------|---|---|---|-------------------|--|
| NCT04878068 | Study to Evaluate the Performance of the Thermo COVID-19 Rapid Antigen Test for Detection of SARS-CoV-2 | 12 years old and above at COVID-19 testing centre | Device: Thermo COVID-19 Rapid Antigen Test; Saliva sample | RT PCR | Establish Performance of Thermo COVID-19 Rapid Antigen Test, Participant Feedback, User Feedback |
| NCT04750629 | Clinical Evaluation of a Point-of-Care (POC), COVID-19 Rapid Antigen Test (CovIDx™) | 1 year old and above suspected of SARS-CoV-2 | Device: POC CovIDx™ Rapid Antigen Test | SARS-CoV-2 RT-PCR | Sensitivity of SARS-CoV-2 antigen in nasal swab as compared to a high-sensitive SARS-CoV-2 RT-PCR granted Emergency Use Authorization (EUA) by the FDA |
| NCT05045846 | Diagnostic Accuracy of Rapid Antigen Test Based on Anterior Nasal Swab Compared With RT-PCR for SARS-CoV-2 Detection. | 18 years old and above who have self-booked appointment at COVID testing centre | Rapid Antigen Test Based on Anterior Nasal Swab | RT-PCR | Accuracy of anterior nasal swab in rapid antigen (Ag)-tests |
| NCT04733170 | Clinical Performance Evaluation of KnowNow SARS-CoV-2 Test for the Detection of COVID-19 Antigen | 18 years old and above suspected with COVID-19 | Diagnostic Test: KnowNow SARS-CoV-2 Rapid Antigen Test | RT-PCR | Assess clinical diagnostic performance of the KnowNow SARS-CoV-2 Rapid Antigen Test, Efficacy to assess the test compared to the reference test method; Usability Questionnaire to evaluate the use of the test with 2 saliva collection methods |
| NCT04689399 | Sensitivity and Specificity of SARS-CoV-2 Rapid Antigen Test Compared to RT-PCR Test | 18 years old above with appointment at COVID-19 testing center | Diagnostic Test: Standard Q COVID-19 Ag - test, produced by SD Biosensor INC. | RT PCR | Sensitivity and specificity of the rapid antigen test of COVID-19 Economic analyses PCR analysis on nasopharyngeal swabs |
| NCT04716088 | Rapid Antigen Testing for SARS-CoV-2 Among Healthcare Workers to Prevent Spread of COVID-19 | 18 years old and above healthcare workers | Repeated Rapid antigen test for SARS CoV2 | RT PCR | SARS CoV2 infection |
| NCT04894760 | Evaluation of a COVID-19 Rapid Diagnostic Test in ER | 18 years old and above with | Diagnostic Test: Rapid Antigen Test (PanBio Ag test) | RT PCR | Diagnostic accuracy |



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| | | | | | |
|-------------|--|--|--|--|---|
| | Departments in Mexico: a Multi-center Study | respiratory symptoms visiting the ER | | | |
| NCT04403906 | Somerset and South Essex Coronavirus Antigen Testing | 18 years old and above with clinical indication for COVID testing | Diagnostic Test: PCL COV05 - COVID 19 Ag Rapid FIA test (Rapid Antigen Test) | RT PCR | To compare the result of SARS-COV2 PCR test to PCL rapid antigen test Number of technically failed samples due to test issues. Time taken for PCL Antigen test result |
| NCT04839094 | Dry Versus Wet Nasopharyngeal Rapid Test for the Detection of COVID-19. | 18 years old and above hospitalized with a confirmed SARS-Cov-2 infection | Diagnostic Test: rapid antigen testing; nasopharyngeal vs saliva samples | RT PCR | Diagnostic assessment Limit of detection determination. |
| NCT04665193 | An Approach to Screening for COVID-19 at Vancouver Airport | Adults with ticket to board a flight from Vancouver International Airport | Rapid Antigen Test | none | COVID-19 status; effectiveness and feasibility of rapid antigen test in airport screening |
| NCT05074017 | COVAG - Covid-19 Antigen Study - the Diagnostic Efficacy of SARS-CoV-2 Rapid Detection Tests | 18 years old and above at testing centers in whom RT- PCR testing for SARS-CoV-2 is medically indicated or requested | 2 rapid antigen tests | RT PCR | Sensitivity of the rapid antigen tests |
| NCT04568356 | A Clinical Evaluation of COVID-19 Rapid Point of Care Antigen Tests | 18 years old and above suspected with COVID | Diagnostic Test: Direct Antigen Tests for COVID-19; nasopharyngeal, nasal and saliva samples | RT PCR | Percent Positive Agreement and Negative Percent Agreement |
| NCT04896710 | COVID-19 Rapid Testing for Self-Administration Among an Asymptomatic Sample | 16 Years to 80 Years Asymptomatic | Device: SD Biosensor RAT; self-administered | professional administered SD Biosensor RAT | Concordance Ability of rapid antigen test to detect COVID-19 positive Acceptability of self-administration |



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| | | | | | |
|-------------|--|---|---|--|---|
| NCT04977050 | The Role of Frequent Point-of-care Molecular Workplace Surveillance for Miners | 18 Years and above miners | Diagnostic Test: Quidel quickvue antigen test for COVID-19 | RT PCR | Screening test (molecular) Diagnostic test (RT-PCR) Serologic antibody test |
| NCT05047900 | The Impact of SARS-CoV-2 Rapid Antigen Testing Kit Screening in Bangkok Community | 10 years and older in any Bangkok community | Routine Use of Rapid antigen testing kit | Did not routinely use Rapid antigen testing kit | Incidence rate of COVID19 infection Incidence rate of severe COVID19 infection Incidence of COVID19 infection in COVID19 vaccinated and unvaccinated people sensitivity and specificity of rapid antigen testing kit |
| NCT04808921 | Performance Evaluation of SARS-COV-2 (Covid-19) Antigen Rapid Test | Child and adults clinically suspected of COVID 19 | Diagnostic Test: Xiamen Wiz Biotech Co., Ltd. SARS-CoV-2 Antigen Rapid Test | RT PCR | Percent Positive Agreement and Negative Percent Agreement |
| NCT04348864 | COVID-19 Diagnostic Self-testing Using Virtual Point-of-care | 18 years old and above suspected or clinically diagnosed with COVID | Diagnostic Test: COVID-19 Antigen/Antibody Rapid Testing, mobile device image capture and telemedicine support Self-test interpretation | RT PCR; Expert Clinical Interpretation of results | Clinical accuracy of the antibody and antigen rapid tests compared to LAMP/PCR-based test result Clinical accuracy of the antibody and antigen rapid tests based on Clinical diagnosis Self-test interpretation of result vs expert clinical image interpretation of result Ease of self-testing procedure |
| NCT04805840 | Sensitivity of Frequent SARS-CoV-2 (COVID-19) Rapid Antigen Testing Regimen | 18 years old and above healthy volunteers | Frequent testing of CoV-SCAN rapid COVID-19 antigen test | RT PCR | Sensitivity of CoV-SCAN daily testing regimen Specificity of CoV-SCAN daily testing regimen Time from CoV-SCAN positive to PCR positive result |
| NCT04898127 | Antigen Rapid Test Screening to Prevent SARS-CoV-2 Transmission (COVID-19) at Mass Gathering Events. | 18 - 45 years old volunteers | Diagnostic Test: SARS-CoV-2 antigen rapid test Rapid test before access to mass gathering event (concert), provided the test result is negative. | RT PCR | SARS-CoV-2 infection Clinical COVID-19 disease Hospital admissions False positive rapid test |



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| | | | | | |
|-------------|--|--|--|---|--|
| NCT04877002 | Performance Study of SONA Saliva C-19 Rapid Test | 18 years old and above presenting with COVID symptoms at the ER | Diagnostic Test: Sona Saliva C-19 Rapid Test | RT PCR | Percent Positive Agreement and Negative Percent Agreement |
| NCT04926779 | Open Label, Single-Center Study Utilizing BIOZEK COVID-19 Antigen Rapid Test | 18 years old and above with COVID-19 symptoms and close contacts | Diagnostic Test: Biozek Covid-19 Antigen Rapid Test (Saliva) | RT PCR | Sensitivity and Specificity of Biozek Covid-19 Antigen Rapid Test (Saliva) Sensitivity and Specificity of Biozek Covid-19 Antigen Rapid Test (Nasopharyngeal Swab) |
| NCT04805892 | Open Label, Single-Center Study Utilizing BIOZEK COVID-19 Antigen Rapid Test. | 18 years old and above with COVID-19 symptoms and close contacts | Diagnostic Test: BIOZEK COVID-19 Antigen Rapid Test; professional vs sel-collected | RT PCR | Sensitivity and Specificity of BIOZEK COVID-19 Antigen Rapid Test on a sample collected by healthcare professionals. Sensitivity and Specificity of BIOZEK COVID-19 Antigen Rapid Test on self-collected sample. |
| NCT04889365 | COVID-19 Antigen Rapid Test Kit | 18 years old and above with or without COVID-19 symptoms | Diagnostic Test: SG Diagnostics COVID-19 Antigen Rapid Test Kit | Diagnostic Test: Polymerase chain reaction (PCR) test | Sensitivity of SG Diagnostics COVID-19 Antigen Rapid Test Kit Specificity of SG Diagnostics COVID-19 Antigen Rapid Test Kit |
| NCT04872075 | Study on Prevention of SARS-CoV-2 Transmission During a Large Indoor Gathering Event | 18 to 45 years old healthy volunteers | Concert Attendees; Diagnostic Test: Rapid nasopharyngeal antigen test for Sars-Cov-2 Diagnostic Test: Saliva Sample | RT PCR; People staying at home | Number of participants with a positive salivary RT-PCR at day 7 after the date of the concert Number of participants in each group with a positive salivary RT-PCR the day of the concert Molecular analysis of transmission cluster |



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Appendix 6 GRADE Evidence Profile

A. OVERALL

Should rapid antigen test be used to diagnose COVID-19 in patients suspected to have COVID-19?

Patient or population: patients suspected to have COVID-19

Pooled sensitivity: 0.71 (95% CI: 0.68 to 0.73)

Pooled specificity: 0.99 (95% CI: 0.99 to 1.00)

| Outcomes | No of studies (No of patients) | Study design | Factors that may decrease certainty of evidence | | | | | Effect per 1,000 patients tested | | | Test Accuracy CoE |
|--|-----------------------------------|--|---|--------------|----------------------|-------------|------------------|----------------------------------|-----------------------------|-----------------------------|-------------------|
| | | | Risk of bias | Indirectness | Inconsistency | Imprecision | Publication Bias | Pre-test probability of 5% | Pre-test probability of 10% | Pre-test probability of 15% | |
| True positives (patients with COVID-19) | 164 studies (43,943 patients) | Cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^b | not serious | none | 36 (34 to 37) | 71 (68 to 73) | 107 (102 to 110) | ⊕⊕○○ Low |
| False negatives (patients incorrectly classified as not having COVID-19) | | | | | | | | 14 (13 to 16) | 29 (27 to 32) | 43 (40 to 48) | |
| True negatives (patients without COVID-19) | 164 studies (191,573 patients) | Cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^b | not serious | none | 945 (943 to 946) | 896 (894 to 896) | 846 (844 to 847) | ⊕⊕○○ Low |
| False positives (patients incorrectly classified as having COVID-19) | | | | | | | | 5 (4 to 7) | 4 (4 to 6) | 4 (3 to 6) | |

CI: confidence interval

Explanations

a. High and unclear risk on all domains (patient selection, conduct of index test and reference standard and patient flow and timing)

b. High heterogeneity across studies ($I^2 = 99\%$ [99-100%])



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B. SYMPTOMATIC

Question: Should rapid antigen test be used to diagnose COVID-19 in symptomatic individuals suspected of COVID-19?

| | |
|-------------|-----------------------------|
| Sensitivity | 0.74 (95% CI: 0.71 to 0.78) |
| Specificity | -- (95% CI: -- to --) |

| | | | |
|-------------|-----|-----|-----|
| Prevalences | 5 % | 10% | 15% |
|-------------|-----|-----|-----|

| Outcome | No of studies (No of patients) | Study design | Factors that may decrease certainty of evidence | | | | | Effect per 1,000 patients tested | | | Test accuracy CoE |
|---|--------------------------------|--|---|--------------|------------------------|-------------|------------------|----------------------------------|-----------------------------|-----------------------------|-------------------|
| | | | Risk of bias | Indirectness | Inconsistency | Imprecision | Publication bias | pre-test probability of 5% | pre-test probability of 10% | pre-test probability of 15% | |
| True positives (patients with COVID-19) | 79 studies 45523 patients | cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^{a,b} | not serious | none | 37 (36 to 39) | 74 (71 to 78) | 111 (107 to 117) | ⊕⊕○○ Low |
| False negatives (patients incorrectly classified as not having COVID-19) | | | | | | | | 13 (11 to 14) | 26 (22 to 29) | 39 (33 to 43) | |
| True negatives (patients without COVID-19) | 0 studies patients | | | | | | | 0 (0 to 0) | 0 (0 to 0) | 0 (0 to 0) | - |
| False positives (patients incorrectly classified as having COVID-19) | | | | | | | | 950 (950 to 950) | 900 (900 to 900) | 850 (850 to 850) | |

Explanations

- a. Unclear and high risk of bias in all domains (e.g. patient selection, conduct of index test and reference stand, and patient flow)
b. High heterogeneity (different timing of testing, test brands, specimen type)



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Should rapid antigen test be used to diagnose COVID-19 in symptomatic individuals suspected of COVID-19?

Patient or population: symptomatic individuals suspected of COVID-19

Setting:

New test: rapid antigen test | **Cut-off value:**

Pooled sensitivity: 0.74 (95% CI: 0.71 to 0.78) | **Pooled specificity:** not calculated

| Test result | Number of results per 1,000 patients tested (95% CI) | | | Number of participants (studies) | Certainty of the Evidence (GRADE) |
|-----------------|--|-------------------------------------|-------------------------------------|----------------------------------|-----------------------------------|
| | Prevalence 5% Typically seen in | Prevalence 10% Typically seen in | Prevalence 15% Typically seen in | | |
| True positives | 37 (36 to 39) | 74 (71 to 78) | 111 (107 to 117) | 45523 (79) | ⊕⊕○○ Low ^{a,b} |
| False negatives | 13 (11 to 14) | 26 (22 to 29) | 39 (33 to 43) | | |
| True negatives | | | | | |
| False positives | | | | | |

CI: confidence interval

Explanations

a. Unclear and high risk of bias in all domains (e.g. patient selection, conduct of index test and reference stand, and patient flow)

b. High heterogeneity ($I^2 = 99\%$ [99-100%])



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C. ASYMPTOMATIC

Question: Should rapid antigen test be used to diagnose COVID-19 in asymptomatic individuals suspected of COVID-19?

| | |
|-------------|-----------------------------|
| Sensitivity | 0.56 (95% CI: 0.51 to 0.62) |
| Specificity | -- (95% CI: -- to --) |

| | | | |
|-------------|-----|-----|-----|
| Prevalences | 5 % | 10% | 15% |
|-------------|-----|-----|-----|

| Outcome | No of studies (No of patients) | Study design | Factors that may decrease certainty of evidence | | | | | Effect per 1,000 patients tested | | | Test accuracy CoE |
|---|--------------------------------|--|---|--------------|----------------------|-------------|------------------|----------------------------------|-----------------------------|-----------------------------|-------------------|
| | | | Risk of bias | Indirectness | Inconsistency | Imprecision | Publication bias | pre-test probability of 5% | pre-test probability of 10% | pre-test probability of 15% | |
| True positives (patients with COVID-19) | 55 studies 72858 patients | cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^b | not serious | none | 28 (26 to 31) | 56 (51 to 62) | 84 (77 to 93) | ⊕⊕○○ Low |
| False negatives (patients incorrectly classified as not having COVID-19) | | | | | | | | 22 (19 to 24) | 44 (38 to 49) | 66 (57 to 73) | |

Explanations

- a. High and unclear risk of bias in all domains (e.g. patient selection, conduct of index test and reference standard, and patient flow and timing)
- b. High heterogeneity (different test brands, specimen types, timing of testing) ($I^2 = 99\%$ [99-100%])



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Should rapid antigen test be used to diagnose COVID-19 in asymptomatic individuals suspected of COVID-19?

Patient or population: asymptomatic individuals suspected of COVID-19

Setting:

New test: rapid antigen test | **Cut-off value:** variable

Pooled sensitivity: 0.56 (95% CI: 0.51 to 0.62) | **Pooled specificity:** not computed

| Test result | Number of results per 1,000 patients tested (95% CI) | | | Number of participants (studies) | Certainty of the Evidence (GRADE) |
|-----------------|--|-------------------------------------|-------------------------------------|----------------------------------|-----------------------------------|
| | Prevalence 5% Typically seen in | Prevalence 10% Typically seen in | Prevalence 15% Typically seen in | | |
| True positives | 28 (26 to 31) | 56 (51 to 62) | 84 (77 to 93) | 72858 (55) | ⊕⊕○○ Low ^{a,b} |
| False negatives | 22 (19 to 24) | 44 (38 to 49) | 66 (57 to 73) | | |
| True negatives | | | | | |
| False positives | | | | | |

CI: confidence interval

Explanations

a. High and unclear risk of bias in all domains (e.g. patient selection, conduct of index test and reference standard, and patient flow and timing)

b. High heterogeneity ($I^2 = 99\%$ [99-100%])



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D. EARLY PHASE

Question: Should rapid antigen test be used to diagnose COVID-19 in in the early phase of illness in individuals suspected of COVID-19??

| | |
|-------------|-----------------------------|
| Sensitivity | 0.79 (95% CI: 0.75 to 0.82) |
| Specificity | -- (95% CI: -- to --) |

| | | | |
|-------------|-----|-----|-----|
| Prevalences | 5 % | 10% | 15% |
|-------------|-----|-----|-----|

| Outcome | No of studies (No of patients) | Study design | Factors that may decrease certainty of evidence | | | | | Effect per 1,000 patients tested | | | Test accuracy CoE |
|--|--------------------------------|---|---|--------------|----------------------|-------------|------------------|----------------------------------|-----------------------------|-----------------------------|-------------------|
| | | | Risk of bias | Indirectness | Inconsistency | Imprecision | Publication bias | pre-test probability of 5% | pre-test probability of 10% | pre-test probability of 15% | |
| True positives (patients with COVID-19) | 54 studies 28591 patients | cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^b | not serious | none | 40 (38 to 41) | 79 (75 to 82) | 119 (112 to 123) | ⊕⊕○○ Low |
| False negatives (patients incorrectly classified as not having COVID-19) | | | | | | | | 10 (9 to 12) | 21 (18 to 25) | 31 (27 to 38) | |

Explanations

- a. High and unclear risk of bias in all domains (e.g. patient selection, conduct of index test and reference standard, and patient flow and timing)
b. High heterogeneity (different test brands, symptom status, specimen) ($I^2 = 99\%$ [99-100%])



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Should rapid antigen test be used to diagnose COVID-19 in in the early phase of illness in individuals suspected of COVID-19??

Patient or population: in the early phase of illness in individuals suspected of COVID-19?

Setting:

New test: rapid antigen test | **Cut-off value:** variable

Pooled sensitivity: 0.79 (95% CI: 0.75 to 0.82) | **Pooled specificity:** not calculated

| Test result | Number of results per 1,000 patients tested (95% CI) | | | Number of participants (studies) | Certainty of the Evidence (GRADE) |
|-----------------|--|-------------------------------------|-------------------------------------|----------------------------------|-----------------------------------|
| | Prevalence 5% Typically seen in | Prevalence 10% Typically seen in | Prevalence 15% Typically seen in | | |
| True positives | 40 (38 to 41) | 79 (75 to 82) | 119 (112 to 123) | 28591 (54) | ⊕⊕○○ Low ^{a,b} |
| False negatives | 10 (9 to 12) | 21 (18 to 25) | 31 (27 to 38) | | |
| True negatives | | | | (0) | - |
| False positives | | | | | |

CI: confidence interval

Explanations

a. High and unclear risk of bias in all domains (e.g. patient selection, conduct of index test and reference standard, and patient flow and timing)

b. High heterogeneity ($I^2 = 99\%$ [99-100%])



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E. SALIVA SPECIMEN

Question: Should rapid antigen test using saliva as specimen be used to diagnose COVID-19 in individuals suspected of COVID-19?

| | |
|-------------|-----------------------------|
| Sensitivity | 0.57 (95% CI: 0.22 to 0.86) |
| Specificity | Not calculated |

| | | | |
|-------------|-----|-----|-----|
| Prevalences | 5 % | 10% | 15% |
|-------------|-----|-----|-----|

| Outcome | No of studies (No of patients) | Study design | Factors that may decrease certainty of evidence | | | | | Effect per 1,000 patients tested | | | Test accuracy CoE | Importance |
|---|--------------------------------|--|---|--------------|----------------------|----------------------|------------------|----------------------------------|-----------------------------|-----------------------------|-------------------|------------|
| | | | Risk of bias | Indirectness | Inconsistency | Imprecision | Publication bias | pre-test probability of 5% | pre-test probability of 10% | pre-test probability of 15% | | |
| True positives (patients with COVID-19) | 7 studies 6148 patients | cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^b | serious ^c | none | 28 (11 to 43) | 57 (22 to 86) | 85 (33 to 129) | Very low | |
| False negatives (patients incorrectly classified as not having COVID-19) | | | | | | | | 22 (7 to 39) | 43 (14 to 78) | 65 (21 to 117) | | |

Explanations

- a. High and unclear risk of bias in all domains (e.g. patient selection, conduct of index test and reference standard, and patient flow and timing)
- b. High heterogeneity ($I^2 = 99\%$ [99-100%])
- c. Very wide estimates



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Should rapid antigen test using saliva as specimen be used to diagnose COVID-19 in individuals suspected of COVID-19?

Patient or population: individuals suspected of COVID-19

Setting:

New test: rapid antigen test – saliva-based | **Cut-off value:** variable

Pooled sensitivity: 0.57 (95% CI: 0.22 to 0.86) | **Pooled specificity:** not calculated

| Test result | Number of results per 1,000 patients tested (95% CI) | | | Number of participants (studies) | Certainty of the Evidence (GRADE) |
|-----------------|--|-------------------------------------|-------------------------------------|----------------------------------|-----------------------------------|
| | Prevalence 5% Typically seen in | Prevalence 10% Typically seen in | Prevalence 15% Typically seen in | | |
| True positives | 28 (11 to 43) | 57 (22 to 86) | 85 (33 to 129) | 6148 (7) | ⊕○○○ Very low ^{a,b,c} |
| False negatives | 22 (7 to 39) | 43 (14 to 78) | 65 (21 to 117) | | |
| True negatives | | | | | |
| False positives | | | | | |

CI: confidence interval

Explanations

a. High and unclear risk of bias in all domains (e.g. patient selection, conduct of index test and reference standard, and patient flow and timing)

b. High heterogeneity ($I^2 = 99\%$ [99-100%])

c. Very wide estimates



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F. REPEAT TESTING

Question: Should repeat testing using rapid antigen tests be used to diagnose COVID-19 in individuals suspected of COVID-19?

| | |
|-------------|----------------|
| Sensitivity | 0.63 to 0.81 |
| Specificity | Not calculated |

| | | | |
|-------------|-----|-----|-----|
| Prevalences | 5 % | 10% | 15% |
|-------------|-----|-----|-----|

| Outcome | No of studies (No of patients) | Study design | Factors that may decrease certainty of evidence | | | | | Effect per 1,000 patients tested | | | Test accuracy CoE |
|--|--------------------------------|---|---|--------------|----------------------|----------------------|------------------|----------------------------------|-----------------------------|-----------------------------|----------------------|
| | | | Risk of bias | Indirectness | Inconsistency | Imprecision | Publication bias | pre-test probability of 5% | pre-test probability of 10% | pre-test probability of 15% | |
| True positives (patients with COVID-19) | 2 studies 4752 patients | cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^b | serious ^c | none | 32 to 41 | 63 to 81 | 95 to 122 | ⊕○○ ○ Very low |
| False negatives (patients incorrectly classified as not having COVID-19) | | | | | | | | 9 to 18 | 19 to 37 | 28 to 55 | |

Explanations

- a. Unclear risk of bias in patient selection and conduct of reference standard
- b. Heterogenous studies with inconsistent sensitivity values ($I^2 = 99\%$ [99-100%])
- c. Wide estimates seen especially in the study of McKay et al.



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Should repeat testing using rapid antigen tests be used to diagnose COVID-19 in individuals suspected of COVID-19?

Patient or population: individuals suspected of COVID-19

Setting:

New test: repeat antigen testing | **Cut-off value:**

Range of sensitivities: 0.63 to 0.81 | **Range of specificities** not calculated

| Test result | Number of results per 1,000 patients tested (95% CI) | | | Number of participants (studies) | Certainty of the Evidence (GRADE) |
|-----------------|--|-------------------------------------|-------------------------------------|----------------------------------|-----------------------------------|
| | Prevalence 5% Typically seen in | Prevalence 10% Typically seen in | Prevalence 15% Typically seen in | | |
| True positives | 32 to 41 | 63 to 81 | 95 to 122 | 4752 (2) | ⊕○○○ Very low ^{a,b,c} |
| False negatives | 9 to 18 | 19 to 37 | 28 to 55 | | |
| True negatives | 0 to 0 | 0 to 0 | 0 to 0 | (0) | - |
| False positives | 950 to 950 | 900 to 900 | 850 to 850 | | |

CI: confidence interval

Explanations

a. Unclear risk of bias in patient selection and conduct of reference standard

b. Heterogenous studies with inconsistent sensitivity values

c. Wide estimates seen especially in the study of McKay et al.



Philippine COVID-19 Living Clinical Practice Guidelines

F. CHILDREN

Question: Should rapid antigen test be used to diagnose COVID-19 in children suspected of COVID-19?

| | |
|-------------|-----------------------------|
| Sensitivity | 0.79 (95% CI: 0.70 to 0.86) |
| Specificity | 0.99 (95% CI: 0.96 to 1.00) |

| | | | |
|-------------|-----|-----|-----|
| Prevalences | 5 % | 10% | 15% |
|-------------|-----|-----|-----|

| Outcome | No of studies (No of patients) | Study design | Factors that may decrease certainty of evidence | | | | | Effect per 1,000 patients tested | | | Test accuracy CoE |
|---|--------------------------------|--|---|--------------|----------------------|-------------|------------------|----------------------------------|-----------------------------|-----------------------------|-------------------|
| | | | Risk of bias | Indirectness | Inconsistency | Imprecision | Publication bias | pre-test probability of 5% | pre-test probability of 10% | pre-test probability of 15% | |
| True positives (patients with COVID-19) | 11 studies 5101 patients | cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^b | not serious | none | 40 (35 to 43) | 79 (70 to 86) | 119 (105 to 129) | ⊕⊕○○ Low |
| False negatives (patients incorrectly classified as not having COVID-19) | | | | | | | | 10 (7 to 15) | 21 (14 to 30) | 31 (21 to 45) | |
| True negatives (patients without COVID-19) | 0 studies patients | cross-sectional (cohort type accuracy study) | serious ^a | not serious | not serious | not serious | none | 945 (916 to 949) | 896 (868 to 899) | 846 (819 to 849) | ⊕⊕⊕○ Moderate |
| False positives (patients incorrectly classified as having COVID-19) | | | | | | | | 5 (1 to 34) | 4 (1 to 32) | 4 (1 to 31) | |

Explanations

- a. High and unclear risk of bias in all domains (e.g. patient selection, conduct of index test and reference standard, and patient flow and timing)
b. High heterogeneity ($I^2 = 99\%$ [99-100%])



Philippine COVID-19 Living Clinical Practice Guidelines

Should rapid antigen test be used to diagnose COVID-19 in children suspected of COVID-19?

Patient or population: children suspected of COVID-19

Setting:

New test: rapid antigen tests | **Cut-off value:**

Pooled sensitivity: 0.79 (95% CI: 0.70 to 0.86) | **Pooled specificity:** 0.99 (95% CI: 0.96 to 1.00)

| Test result | Number of results per 1,000 patients tested (95% CI) | | | Number of participants (studies) | Certainty of the Evidence (GRADE) |
|-----------------|--|-------------------------------------|-------------------------------------|----------------------------------|-----------------------------------|
| | Prevalence 5% Typically seen in | Prevalence 10% Typically seen in | Prevalence 15% Typically seen in | | |
| True positives | 40 (35 to 43) | 79 (70 to 86) | 119 (105 to 129) | 5101 (11) | ⊕⊕○○ Low ^{a,b} |
| False negatives | 10 (7 to 15) | 21 (14 to 30) | 31 (21 to 45) | | |
| True negatives | 945 (916 to 949) | 896 (868 to 899) | 846 (819 to 849) | 5101 (11) | ⊕⊕⊕○ Moderate ^a |
| False positives | 5 (1 to 34) | 4 (1 to 32) | 4 (1 to 31) | | |

CI: confidence interval

Explanations

a. High and unclear risk of bias in all domains (e.g. patient selection, conduct of index test and reference standard, and patient flow and timing)

b. High heterogeneity



Philippine COVID-19 Living Clinical Practice Guidelines

G. OUTBREAK SETTINGS

Should rapid antigen test be used to diagnose COVID-19 in individuals suspected of COVID-19 during outbreaks?

Patient or population: individuals suspected to have COVID-19 (includes both symptomatic and asymptomatic patients)

Range of sensitivity: 0.14 to 0.89

Range of specificity: 0.88 to 1.0

| Outcomes | No of studies (No of patients) | Study design | Factors that may decrease certainty of evidence | | | | | Effect per 1,000 patients tested | | | Test Accuracy CoE |
|--|-----------------------------------|---|---|--------------|----------------------|----------------------|------------------|----------------------------------|-----------------------------|-----------------------------|-------------------|
| | | | Risk of bias | Indirectness | Inconsistency | Imprecision | Publication Bias | Pre-test probability of 5% | Pre-test probability of 10% | Pre-test probability of 15% | |
| True positives (patients with COVID-19) | 3 studies (398 patients) | Cross-sectional (cohort type accuracy study) | very serious ^a | not serious | serious ^b | serious ^c | none | 7 to 45 | 14 to 89 | 21 to 134 | ⊕○○○ Very low |
| False negatives (patients incorrectly classified as not having COVID-19) | | | | | | | | 5 to 43 | 11 to 86 | 16 to 129 | |
| True negatives (patients without COVID-19) | 3 studies (398) | Cross-sectional (cohort type accuracy study) | very serious ^a | not serious | serious ^b | not serious | none | 836 to 950 | 792 to 900 | 748 to 850 | ⊕⊕○○ Low |
| False positives (patients incorrectly classified as having COVID-19) | | | | | | | | 0 to 114 | 0 to 108 | 0 to 102 | |

CI: confidence interval

Explanations

- Majority of included studies have high risk of bias most notably in the patient selection, conduct of reference standard, and patient flow and timing
- High heterogeneity across studies (different timing of testing, test brands, specimen type) - $I^2 = 99\%$
- Wide confidence intervals



Philippine COVID-19 Living Clinical Practice Guidelines

Should rapid antigen test be used to diagnose COVID-19 in asymptomatic individuals during outbreaks?

Patient or population: asymptomatic individuals in an outbreak setting

Range sensitivity: 0.64 to 0.92

Range specificity: 0.92 to 0.98

| Outcomes | No of studies (No of patients) | Study design | Factors that may decrease certainty of evidence | | | | | Effect per 1,000 patients tested | | | Test Accuracy CoE |
|--|-----------------------------------|---|---|--------------|----------------------|----------------------|------------------|----------------------------------|-----------------------------|-----------------------------|----------------------|
| | | | Risk of bias | Indirectness | Inconsistency | Imprecision | Publication Bias | Pre-test probability of 5% | Pre-test probability of 10% | Pre-test probability of 15% | |
| True positives (patients with COVID-19) | 2 studies (1,127 patients) | Cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^a | serious ^a | none | 32 to 46 | 64 to 92 | 96 to 138 | ⊕○○○ VeryLow |
| False negatives (patients incorrectly classified as not having COVID-19) | | | | | | | | 4 to 18 | 8 to 36 | 12 to 54 | |
| True negatives (patients without COVID-19) | 2 studies (1,127 patients) | Cross-sectional (cohort type accuracy study) | serious ^a | not serious | serious ^a | not serious | none | 874 to 931 | 828 to 882 | 782 to 833 | ⊕⊕○○ Low |
| False positives (patients incorrectly classified as having COVID-19) | | | | | | | | 19 to 76 | 18 to 72 | 17 to 68 | |

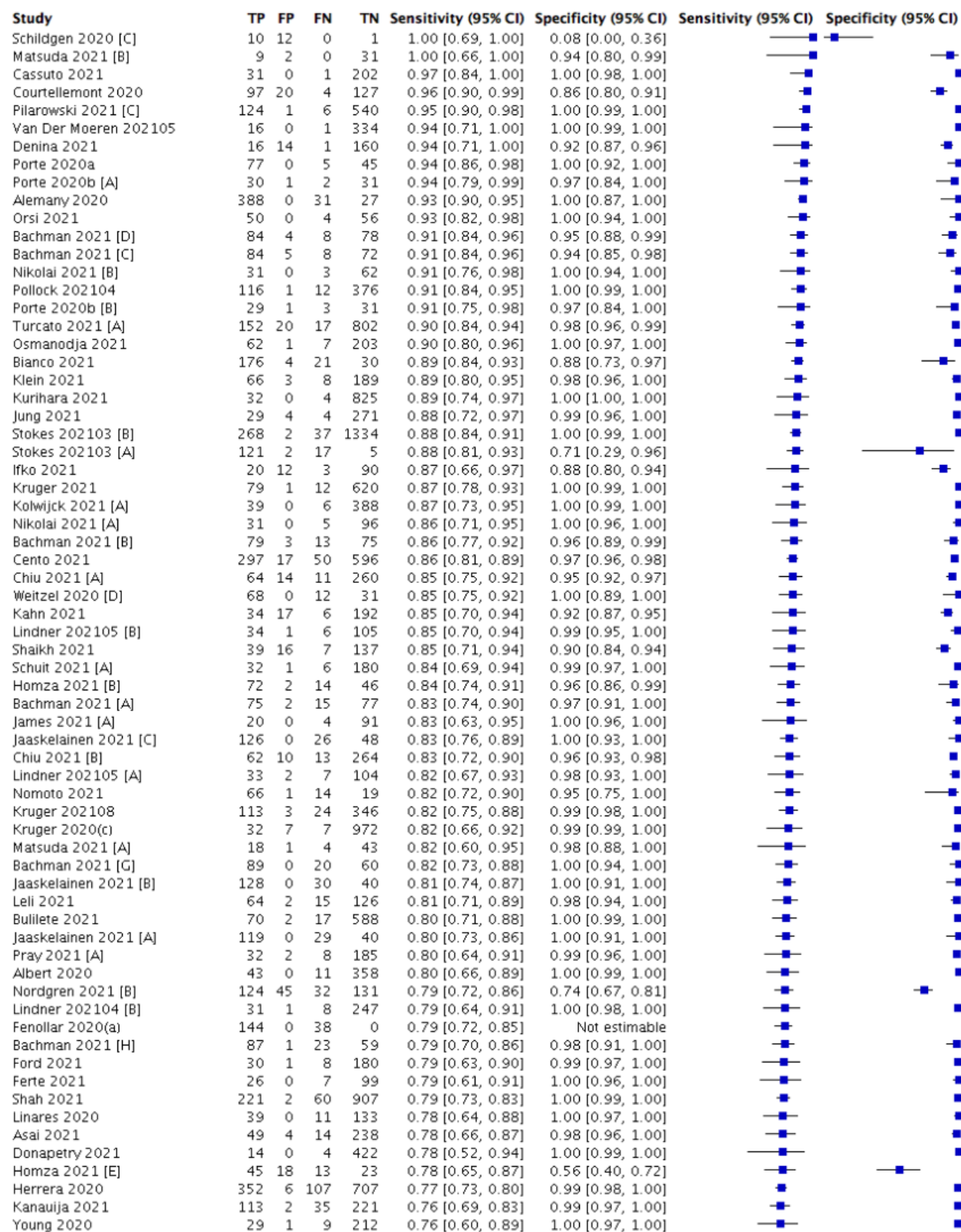
CI: confidence interval

Explanations

- a. Unclear issues on patient selection domain
- b. High heterogeneity across studies (different test brands, different timing of testing)
- c. few studies, wide confidence interval



Appendix 7. Forest Plots





Philippine COVID-19 Living Clinical Practice Guidelines

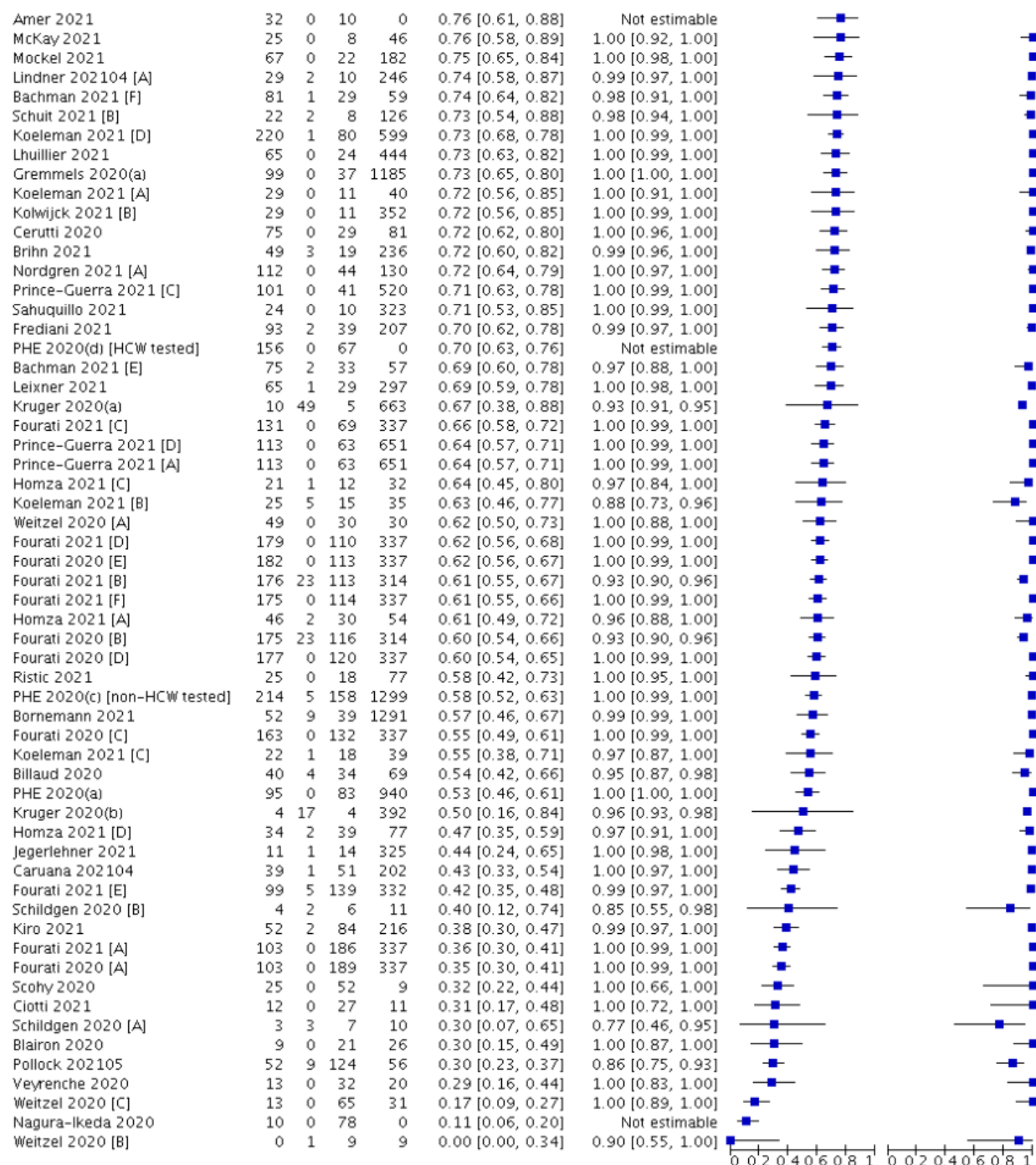


Figure 1. Sensitivity and specificity of RAgT in symptomatic individuals



Philippine COVID-19 Living Clinical Practice Guidelines

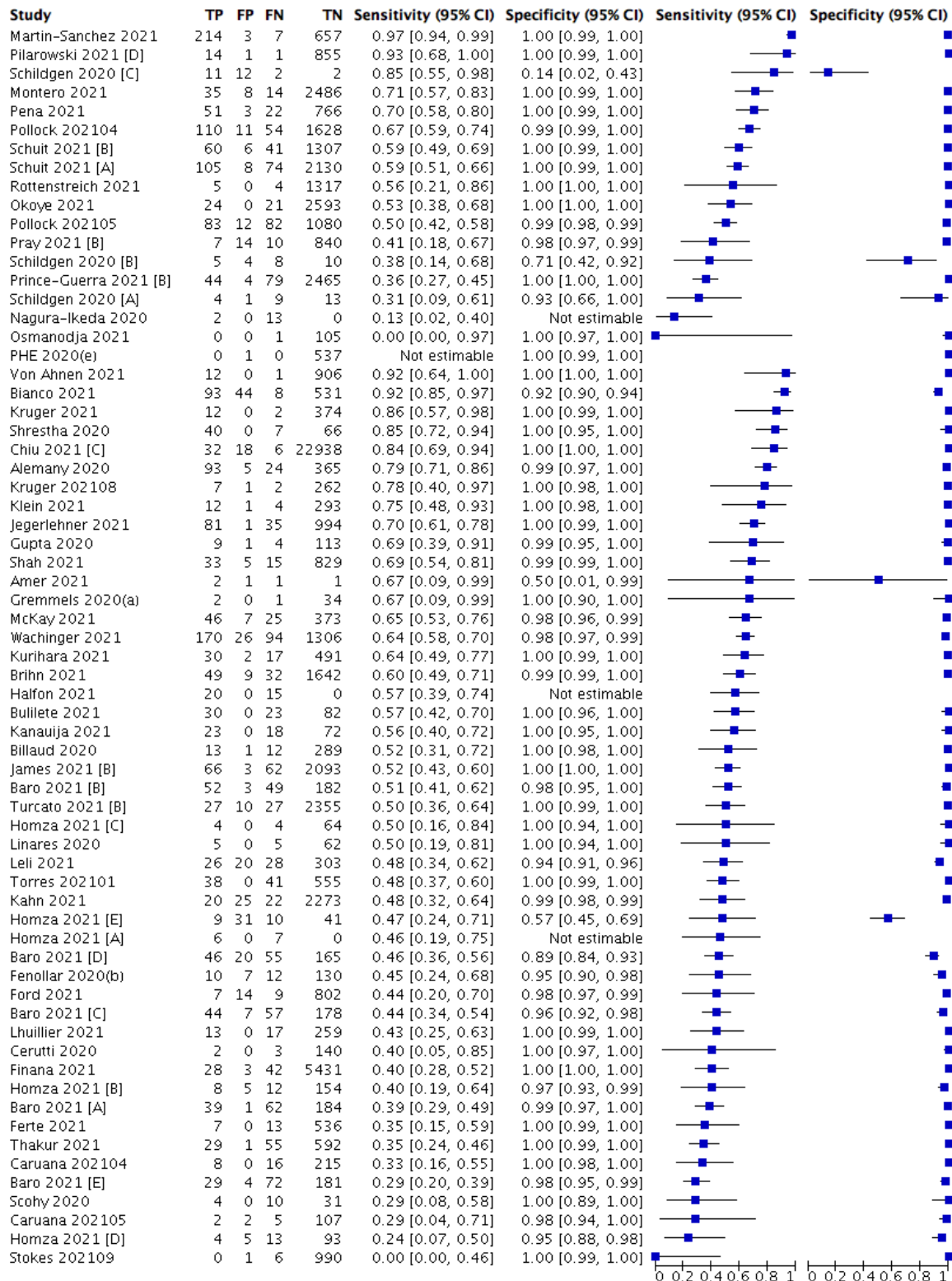


Figure 2. Sensitivity and specificity of RAgT in asymptomatic individuals



Philippine COVID-19 Living Clinical Practice Guidelines

| Study | TP | FP | FN | TN | Sensitivity (95% CI) | Specificity (95% CI) | Sensitivity (95% CI) | Specificity (95% CI) |
|-----------------------|-----|-----|----|------|----------------------|----------------------|----------------------|----------------------|
| Gili 2021 | 90 | 130 | 0 | 1518 | 1.00 [0.96, 1.00] | 0.92 [0.91, 0.93] | | |
| Ishii 2021 [D] | 8 | 1 | 0 | 123 | 1.00 [0.63, 1.00] | 0.99 [0.96, 1.00] | | |
| Kobayashi 2021 [A] | 304 | 0 | 1 | 4615 | 1.00 [0.98, 1.00] | 1.00 [1.00, 1.00] | | |
| Chaimayo 2021 | 59 | 5 | 1 | 389 | 0.98 [0.91, 1.00] | 0.99 [0.97, 1.00] | | |
| Kobayashi 2021 [B] | 111 | 3 | 3 | 5272 | 0.97 [0.93, 0.99] | 1.00 [1.00, 1.00] | | |
| Courtellemont 2021 | 117 | 0 | 4 | 127 | 0.97 [0.92, 0.99] | 1.00 [0.97, 1.00] | | |
| Kim 2021 [B] | 94 | 0 | 6 | 100 | 0.94 [0.87, 0.98] | 1.00 [0.96, 1.00] | | |
| Sood 2021 | 127 | 99 | 9 | 539 | 0.93 [0.88, 0.97] | 0.84 [0.81, 0.87] | | |
| Merino-Amador 2021 | 179 | 2 | 13 | 256 | 0.93 [0.89, 0.96] | 0.99 [0.97, 1.00] | | |
| Maniscalco 2021 | 12 | 1 | 1 | 91 | 0.92 [0.64, 1.00] | 0.99 [0.94, 1.00] | | |
| Alemayehu 2020 | 872 | 5 | 79 | 450 | 0.92 [0.90, 0.93] | 0.99 [0.97, 1.00] | | |
| Ishii 2021 [C] | 22 | 1 | 2 | 460 | 0.92 [0.73, 0.99] | 1.00 [0.99, 1.00] | | |
| Yokota 2021 | 31 | 2 | 3 | 307 | 0.91 [0.76, 0.98] | 0.99 [0.98, 1.00] | | |
| Seynaeve 2021 [C] | 41 | 0 | 4 | 50 | 0.91 [0.79, 0.98] | 1.00 [0.93, 1.00] | | |
| Ishii 2021 [A] | 10 | 0 | 1 | 260 | 0.91 [0.59, 1.00] | 1.00 [0.99, 1.00] | | |
| Bianco 2021 | 269 | 48 | 29 | 561 | 0.90 [0.86, 0.93] | 0.92 [0.90, 0.94] | | |
| Bouassa 2021 | 90 | 0 | 10 | 50 | 0.90 [0.82, 0.95] | 1.00 [0.93, 1.00] | | |
| Kim 2021 [A] | 27 | 2 | 3 | 98 | 0.90 [0.73, 0.98] | 0.98 [0.93, 1.00] | | |
| Van Honacker 2021 [A] | 52 | 21 | 6 | 18 | 0.90 [0.79, 0.96] | 0.46 [0.30, 0.63] | | |
| Agarwal 2021 | 26 | 2 | 3 | 436 | 0.90 [0.73, 0.98] | 1.00 [0.98, 1.00] | | |
| Pilarowski 2021 [A] | 211 | 3 | 26 | 3062 | 0.89 [0.84, 0.93] | 1.00 [1.00, 1.00] | | |
| Berger 2021 [B] | 170 | 1 | 21 | 337 | 0.89 [0.84, 0.93] | 1.00 [0.98, 1.00] | | |
| Pickering 2021 [A] | 89 | 1 | 11 | 99 | 0.89 [0.81, 0.94] | 0.99 [0.95, 1.00] | | |
| Osmanodja 2021 | 62 | 1 | 8 | 308 | 0.89 [0.79, 0.95] | 1.00 [0.98, 1.00] | | |
| Karon 2021 [D] | 174 | 0 | 23 | 150 | 0.88 [0.83, 0.92] | 1.00 [0.98, 1.00] | | |
| Schildgen 2020 [C] | 37 | 25 | 5 | 6 | 0.88 [0.74, 0.96] | 0.19 [0.07, 0.37] | | |
| Seynaeve 2021 [D] | 44 | 0 | 6 | 50 | 0.88 [0.76, 0.95] | 1.00 [0.93, 1.00] | | |
| Andreani 2021 [F] | 109 | 32 | 15 | 83 | 0.88 [0.81, 0.93] | 0.72 [0.63, 0.80] | | |
| Baccani 2021 [A] | 29 | 7 | 4 | 161 | 0.88 [0.72, 0.97] | 0.96 [0.92, 0.98] | | |
| Karon 2021 [C] | 164 | 0 | 23 | 150 | 0.88 [0.82, 0.92] | 1.00 [0.98, 1.00] | | |
| Kruger 2021 | 92 | 1 | 14 | 1001 | 0.87 [0.79, 0.93] | 1.00 [0.99, 1.00] | | |
| Caputo 2021 | 436 | 102 | 67 | 3661 | 0.87 [0.83, 0.90] | 0.97 [0.97, 0.98] | | |
| Klein 2021 | 78 | 4 | 12 | 486 | 0.87 [0.78, 0.93] | 0.99 [0.98, 1.00] | | |
| Berger 2021 [A] | 106 | 0 | 18 | 411 | 0.85 [0.78, 0.91] | 1.00 [0.99, 1.00] | | |
| Kyritsi 2021 | 141 | 1 | 24 | 458 | 0.85 [0.79, 0.90] | 1.00 [0.99, 1.00] | | |
| Iglol 2021 | 158 | 4 | 28 | 780 | 0.85 [0.79, 0.90] | 0.99 [0.99, 1.00] | | |
| Hartard 2021 | 39 | 2 | 7 | 330 | 0.85 [0.71, 0.94] | 0.99 [0.98, 1.00] | | |
| Escriva 2021 | 99 | 0 | 18 | 331 | 0.85 [0.77, 0.91] | 1.00 [0.99, 1.00] | | |
| Tsai 2021 | 5 | 1 | 1 | 54 | 0.83 [0.36, 1.00] | 0.98 [0.90, 1.00] | | |
| Favresse 2021 [E] | 80 | 0 | 16 | 92 | 0.83 [0.74, 0.90] | 1.00 [0.96, 1.00] | | |
| Van Honacker 2021 [C] | 48 | 3 | 10 | 37 | 0.83 [0.71, 0.91] | 0.93 [0.80, 0.98] | | |
| Van Honacker 2021 [E] | 48 | 0 | 10 | 40 | 0.83 [0.71, 0.91] | 1.00 [0.91, 1.00] | | |
| Eleftheriou 2021 | 42 | 0 | 9 | 693 | 0.82 [0.69, 0.92] | 1.00 [0.99, 1.00] | | |
| Kruger 202108 | 120 | 4 | 26 | 611 | 0.82 [0.75, 0.88] | 0.99 [0.98, 1.00] | | |
| Gupta 2020 | 63 | 1 | 14 | 252 | 0.82 [0.71, 0.90] | 1.00 [0.98, 1.00] | | |
| Nsoga 2021 | 136 | 2 | 32 | 232 | 0.81 [0.74, 0.87] | 0.99 [0.97, 1.00] | | |
| Gremmels 2020(b) | 51 | 0 | 12 | 145 | 0.81 [0.69, 0.90] | 1.00 [0.97, 1.00] | | |
| Takeda 2020 | 50 | 0 | 12 | 100 | 0.81 [0.69, 0.90] | 1.00 [0.96, 1.00] | | |
| Holzner 2021 | 367 | 8 | 89 | 1816 | 0.80 [0.77, 0.84] | 1.00 [0.99, 1.00] | | |
| Nash 2020 | 80 | 8 | 20 | 82 | 0.80 [0.71, 0.87] | 0.91 [0.83, 0.96] | | |
| Lindner 202104 [B] | 31 | 1 | 8 | 247 | 0.79 [0.64, 0.91] | 1.00 [0.98, 1.00] | | |
| Van Honacker 2021 [D] | 45 | 0 | 12 | 40 | 0.79 [0.66, 0.89] | 1.00 [0.91, 1.00] | | |
| Bruzzzone 2021 [A] | 199 | 0 | 54 | 68 | 0.79 [0.73, 0.84] | 1.00 [0.95, 1.00] | | |
| Salvagno 2021 | 47 | 1 | 13 | 113 | 0.78 [0.66, 0.88] | 0.99 [0.95, 1.00] | | |
| Amer 2021 | 54 | 5 | 15 | 9 | 0.78 [0.67, 0.87] | 0.64 [0.35, 0.87] | | |
| Karon 2021 [B] | 153 | 4 | 44 | 146 | 0.78 [0.71, 0.83] | 0.97 [0.93, 0.99] | | |
| Pollock 202104 | 226 | 12 | 66 | 2004 | 0.77 [0.72, 0.82] | 0.99 [0.99, 1.00] | | |
| Muhi 2021 | 17 | 1 | 5 | 2580 | 0.77 [0.55, 0.92] | 1.00 [1.00, 1.00] | | |
| Shah 2021 | 258 | 7 | 76 | 1769 | 0.77 [0.72, 0.82] | 1.00 [0.99, 1.00] | | |
| Favresse 2021 [C] | 74 | 3 | 22 | 89 | 0.77 [0.67, 0.85] | 0.97 [0.91, 0.99] | | |
| Pickering 2021 [B] | 77 | 2 | 23 | 98 | 0.77 [0.68, 0.85] | 0.98 [0.93, 1.00] | | |
| Abusrewil 2021 [A] | 83 | 0 | 25 | 123 | 0.77 [0.68, 0.84] | 1.00 [0.97, 1.00] | | |
| Pena-Rodriguez 2021 | 79 | 0 | 25 | 265 | 0.76 [0.67, 0.84] | 1.00 [0.99, 1.00] | | |
| Torres 202102 | 88 | 0 | 28 | 154 | 0.76 [0.67, 0.83] | 1.00 [0.98, 1.00] | | |
| Homza 2021 [B] | 81 | 7 | 26 | 204 | 0.76 [0.66, 0.83] | 0.97 [0.93, 0.99] | | |
| Pickering 2021 [C] | 75 | 14 | 25 | 86 | 0.75 [0.65, 0.83] | 0.86 [0.78, 0.92] | | |
| Soleimani 2021 [B] | 174 | 0 | 58 | 169 | 0.75 [0.69, 0.80] | 1.00 [0.98, 1.00] | | |
| Kurihara 2021 | 62 | 2 | 21 | 1316 | 0.75 [0.64, 0.84] | 1.00 [0.99, 1.00] | | |
| Landaas 2021 | 186 | 3 | 64 | 3738 | 0.74 [0.69, 0.80] | 1.00 [1.00, 1.00] | | |
| Pickering 2021 [D] | 74 | 0 | 26 | 100 | 0.74 [0.64, 0.82] | 1.00 [0.96, 1.00] | | |
| Paul 2021 [A] | 72 | 0 | 26 | 50 | 0.73 [0.64, 0.82] | 1.00 [0.93, 1.00] | | |
| Linares 2020 | 44 | 0 | 16 | 195 | 0.73 [0.60, 0.84] | 1.00 [0.98, 1.00] | | |
| Gremmels 2020(a) | 101 | 0 | 38 | 1228 | 0.73 [0.64, 0.80] | 1.00 [1.00, 1.00] | | |
| Iqbal 2021 | 92 | 1 | 35 | 42 | 0.72 [0.64, 0.80] | 0.98 [0.88, 1.00] | | |
| Halfon 2021 | 72 | 1 | 28 | 99 | 0.72 [0.62, 0.81] | 0.99 [0.95, 1.00] | | |
| Kanaujia 2021 | 136 | 2 | 53 | 293 | 0.72 [0.65, 0.78] | 0.99 [0.98, 1.00] | | |
| Bullete 2021 | 100 | 2 | 40 | 670 | 0.71 [0.63, 0.79] | 1.00 [0.99, 1.00] | | |
| Kruttgen 2020 | 53 | 3 | 22 | 72 | 0.71 [0.59, 0.81] | 0.96 [0.89, 0.99] | | |



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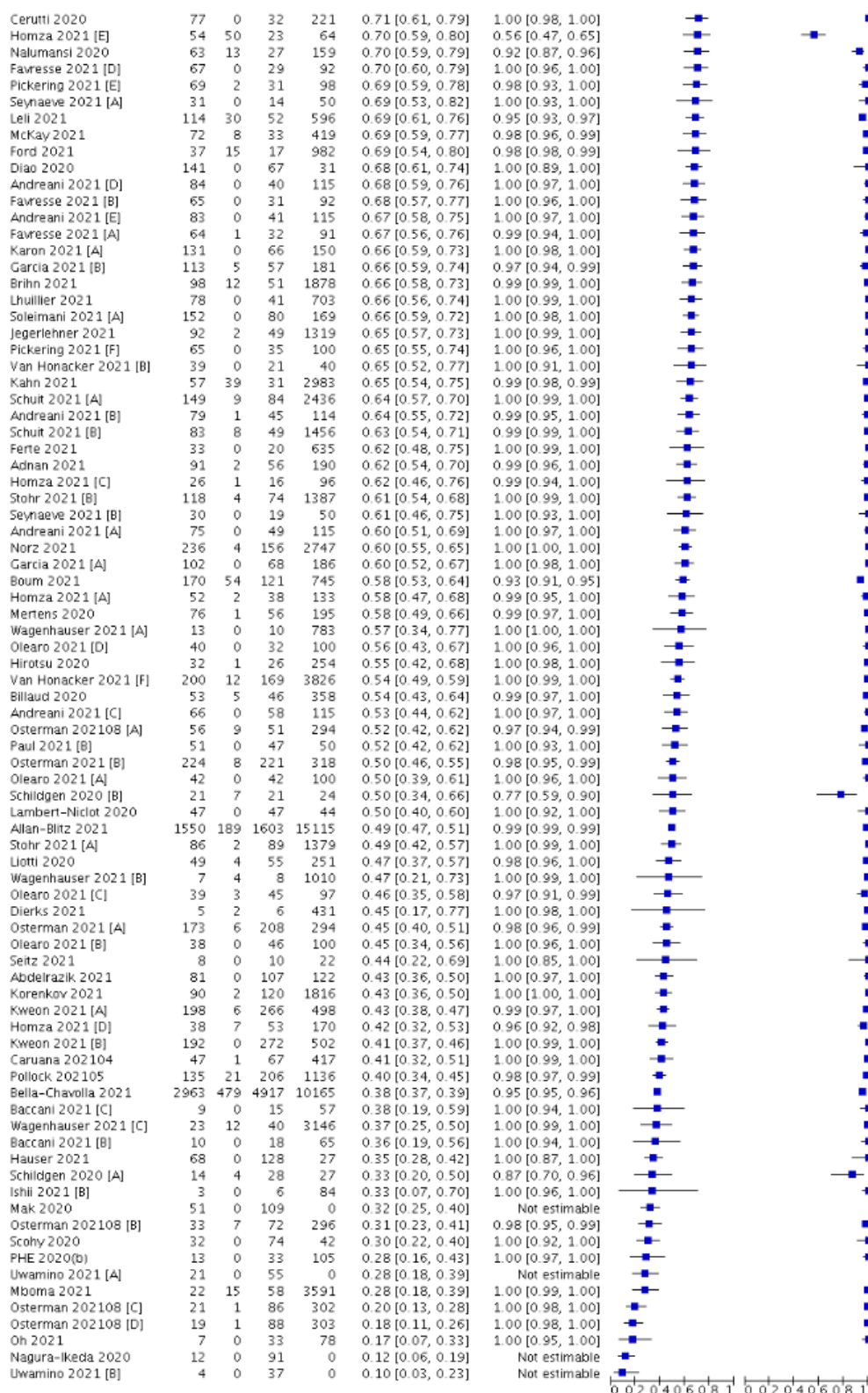


Figure 3. Sensitivity and specificity of RAgtT in individuals with mixed symptoms or unreported



Philippine COVID-19 Living Clinical Practice Guidelines

| Study | TP | FP | FN | TN | Sensitivity (95% CI) | Specificity (95% CI) | Sensitivity (95% CI) | Specificity (95% CI) |
|------------------------|-----|----|----|------|----------------------|----------------------|----------------------|----------------------|
| Kim 2021 [A] | 7 | 0 | 0 | 0 | 1.00 [0.59, 1.00] | Not estimable | —■ | —■ |
| Muhi 2021 | 15 | 0 | 0 | 156 | 1.00 [0.78, 1.00] | 1.00 [0.98, 1.00] | —■ | —■ |
| Cassuto 2021 | 31 | 0 | 1 | 202 | 0.97 [0.84, 1.00] | 1.00 [0.98, 1.00] | —■ | —■ |
| Courtellemont 2021 | 117 | 0 | 4 | 127 | 0.97 [0.92, 0.99] | 1.00 [0.97, 1.00] | —■ | —■ |
| Kim 2021 [B] | 64 | 0 | 3 | 0 | 0.96 [0.87, 0.99] | Not estimable | —■ | —■ |
| Porte 2020a | 72 | 0 | 4 | 42 | 0.95 [0.87, 0.99] | 1.00 [0.92, 1.00] | —■ | —■ |
| Denina 2021 | 16 | 14 | 1 | 160 | 0.94 [0.71, 1.00] | 0.92 [0.87, 0.96] | —■ | —■ |
| Porte 2020b [A] | 30 | 1 | 2 | 31 | 0.94 [0.79, 0.99] | 0.97 [0.84, 1.00] | —■ | —■ |
| Pollock 202104 | 104 | 0 | 7 | 335 | 0.94 [0.87, 0.97] | 1.00 [0.99, 1.00] | —■ | —■ |
| Nomoto 2021 | 44 | 0 | 3 | 4 | 0.94 [0.82, 0.99] | 1.00 [0.40, 1.00] | —■ | —■ |
| Merino-Amador 2021 | 179 | 2 | 13 | 256 | 0.93 [0.89, 0.96] | 0.99 [0.97, 1.00] | —■ | —■ |
| Bachman 2021 [C] | 84 | 5 | 8 | 72 | 0.91 [0.84, 0.96] | 0.94 [0.85, 0.98] | —■ | —■ |
| Bachman 2021 [D] | 84 | 4 | 8 | 78 | 0.91 [0.84, 0.96] | 0.95 [0.88, 0.99] | —■ | —■ |
| Seynaeve 2021 [C] | 41 | 0 | 4 | 50 | 0.91 [0.79, 0.98] | 1.00 [0.93, 1.00] | —■ | —■ |
| Kruger 2021 | 69 | 1 | 7 | 533 | 0.91 [0.82, 0.96] | 1.00 [0.99, 1.00] | —■ | —■ |
| Porte 2020b [B] | 29 | 1 | 3 | 31 | 0.91 [0.75, 0.98] | 0.97 [0.84, 1.00] | —■ | —■ |
| Garcia 2021 [B] | 66 | 0 | 7 | 0 | 0.90 [0.81, 0.96] | Not estimable | —■ | —■ |
| Bianco 2021 | 269 | 48 | 29 | 561 | 0.90 [0.86, 0.93] | 0.92 [0.90, 0.94] | —■ | —■ |
| Kweon 2021 [A] | 135 | 0 | 16 | 0 | 0.89 [0.83, 0.94] | Not estimable | —■ | —■ |
| Berger 2021 [B] | 170 | 1 | 21 | 337 | 0.89 [0.84, 0.93] | 1.00 [0.98, 1.00] | —■ | —■ |
| Osmanodja 2021 | 62 | 1 | 8 | 308 | 0.89 [0.79, 0.95] | 1.00 [0.98, 1.00] | —■ | —■ |
| Kweon 2021 [B] | 135 | 0 | 18 | 0 | 0.88 [0.82, 0.93] | Not estimable | —■ | —■ |
| Seynaeve 2021 [D] | 44 | 0 | 6 | 50 | 0.88 [0.76, 0.95] | 1.00 [0.93, 1.00] | —■ | —■ |
| Stokes 202103 [B] | 268 | 2 | 37 | 1334 | 0.88 [0.84, 0.91] | 1.00 [0.99, 1.00] | —■ | —■ |
| Stokes 202103 [A] | 121 | 2 | 17 | 5 | 0.88 [0.81, 0.93] | 0.71 [0.29, 0.96] | —■ | —■ |
| Garcia 2021 [A] | 64 | 0 | 9 | 0 | 0.88 [0.78, 0.94] | Not estimable | —■ | —■ |
| Ifko 2021 | 20 | 12 | 3 | 90 | 0.87 [0.66, 0.97] | 0.88 [0.80, 0.94] | —■ | —■ |
| Kolwijck 2021 [A] | 39 | 0 | 6 | 388 | 0.87 [0.73, 0.95] | 1.00 [0.99, 1.00] | —■ | —■ |
| Linares 2020 | 32 | 0 | 5 | 846 | 0.86 [0.71, 0.95] | 1.00 [1.00, 1.00] | —■ | —■ |
| Kruger 202108 | 102 | 2 | 16 | 303 | 0.86 [0.79, 0.92] | 0.99 [0.98, 1.00] | —■ | —■ |
| Gupta 2020 | 49 | 0 | 8 | 134 | 0.86 [0.74, 0.94] | 1.00 [0.97, 1.00] | —■ | —■ |
| Bachman 2021 [B] | 79 | 3 | 13 | 75 | 0.86 [0.77, 0.92] | 0.96 [0.89, 0.99] | —■ | —■ |
| McKay 2021 | 48 | 0 | 8 | 0 | 0.86 [0.74, 0.94] | Not estimable | —■ | —■ |
| Berger 2021 [A] | 106 | 0 | 18 | 411 | 0.85 [0.78, 0.91] | 1.00 [0.99, 1.00] | —■ | —■ |
| Kyritsi 2021 | 141 | 1 | 24 | 458 | 0.85 [0.79, 0.90] | 1.00 [0.99, 1.00] | —■ | —■ |
| Chiu 2021 [A] | 64 | 14 | 11 | 260 | 0.85 [0.75, 0.92] | 0.95 [0.92, 0.97] | —■ | —■ |
| Lindner 202105 [B] | 34 | 1 | 6 | 105 | 0.85 [0.70, 0.94] | 0.99 [0.95, 1.00] | —■ | —■ |
| Weitzel 2020 [D] | 68 | 0 | 12 | 31 | 0.85 [0.75, 0.92] | 1.00 [0.89, 1.00] | —■ | —■ |
| Shaikh 2021 | 39 | 16 | 7 | 137 | 0.85 [0.71, 0.94] | 0.90 [0.84, 0.94] | —■ | —■ |
| Escriva 2021 | 99 | 0 | 18 | 331 | 0.85 [0.77, 0.91] | 1.00 [0.99, 1.00] | —■ | —■ |
| Amer 2021 | 31 | 1 | 6 | 0 | 0.84 [0.68, 0.94] | 0.00 [0.00, 0.97] | —■ | —■ |
| Bachman 2021 [A] | 75 | 2 | 15 | 77 | 0.83 [0.74, 0.90] | 0.97 [0.91, 1.00] | —■ | —■ |
| Jaaskelainen 2021 [C] | 126 | 0 | 26 | 38 | 0.83 [0.76, 0.89] | 1.00 [0.91, 1.00] | —■ | —■ |
| Halfon 2021 | 29 | 0 | 6 | 0 | 0.83 [0.66, 0.93] | Not estimable | —■ | —■ |
| Chiu 2021 [B] | 62 | 10 | 13 | 264 | 0.83 [0.72, 0.90] | 0.96 [0.93, 0.98] | —■ | —■ |
| Lindner 202105 [A] | 33 | 2 | 7 | 104 | 0.82 [0.67, 0.93] | 0.98 [0.93, 1.00] | —■ | —■ |
| Shah 2021 | 199 | 2 | 44 | 684 | 0.82 [0.76, 0.87] | 1.00 [0.99, 1.00] | —■ | —■ |
| Bachman 2021 [G] | 89 | 0 | 20 | 60 | 0.82 [0.73, 0.88] | 1.00 [0.94, 1.00] | —■ | —■ |
| Jaaskelainen 2021 [B] | 128 | 0 | 30 | 40 | 0.81 [0.74, 0.87] | 1.00 [0.91, 1.00] | —■ | —■ |
| Jaaskelainen 2021 [A] | 119 | 0 | 29 | 40 | 0.80 [0.73, 0.86] | 1.00 [0.91, 1.00] | —■ | —■ |
| Kruger 2020(c) | 28 | 7 | 7 | 907 | 0.80 [0.63, 0.92] | 0.99 [0.98, 1.00] | —■ | —■ |
| Albert 2020 | 43 | 0 | 11 | 358 | 0.80 [0.66, 0.89] | 1.00 [0.99, 1.00] | —■ | —■ |
| Ristic 2021 | 19 | 0 | 5 | 39 | 0.79 [0.58, 0.93] | 1.00 [0.91, 1.00] | —■ | —■ |
| Bachman 2021 [H] | 87 | 1 | 23 | 59 | 0.79 [0.70, 0.86] | 0.98 [0.91, 1.00] | —■ | —■ |
| Asai 2021 | 49 | 4 | 14 | 238 | 0.78 [0.66, 0.87] | 0.98 [0.96, 1.00] | —■ | —■ |
| Donapetry 2021 | 14 | 0 | 4 | 422 | 0.78 [0.52, 0.94] | 1.00 [0.99, 1.00] | —■ | —■ |
| Young 2020 | 29 | 1 | 9 | 212 | 0.76 [0.60, 0.89] | 1.00 [0.97, 1.00] | —■ | —■ |
| Torres 202102 | 88 | 0 | 28 | 154 | 0.76 [0.67, 0.83] | 1.00 [0.98, 1.00] | —■ | —■ |
| Gremmels 2020(a) | 75 | 0 | 26 | 846 | 0.74 [0.65, 0.82] | 1.00 [1.00, 1.00] | —■ | —■ |
| Bachman 2021 [F] | 81 | 1 | 29 | 59 | 0.74 [0.64, 0.82] | 0.98 [0.91, 1.00] | —■ | —■ |
| Kolwijck 2021 [B] | 29 | 0 | 11 | 352 | 0.72 [0.56, 0.85] | 1.00 [0.99, 1.00] | —■ | —■ |
| Fourati 2021 [B] | 143 | 0 | 57 | 0 | 0.71 [0.65, 0.78] | Not estimable | —■ | —■ |
| Prince-Guerra 2021 [C] | 101 | 0 | 41 | 520 | 0.71 [0.63, 0.78] | 1.00 [0.99, 1.00] | —■ | —■ |
| Fourati 2020 [E] | 142 | 0 | 58 | 0 | 0.71 [0.64, 0.77] | Not estimable | —■ | —■ |
| Fourati 2021 [D] | 142 | 0 | 58 | 0 | 0.71 [0.64, 0.77] | Not estimable | —■ | —■ |
| Fourati 2020 [B] | 141 | 0 | 58 | 0 | 0.71 [0.64, 0.77] | Not estimable | —■ | —■ |
| Sahuquillo 2021 | 24 | 0 | 10 | 323 | 0.71 [0.53, 0.85] | 1.00 [0.99, 1.00] | —■ | —■ |
| Frediani 2021 | 93 | 2 | 39 | 207 | 0.70 [0.62, 0.78] | 0.99 [0.97, 1.00] | —■ | —■ |
| Bachman 2021 [E] | 75 | 2 | 33 | 57 | 0.69 [0.60, 0.78] | 0.97 [0.88, 1.00] | —■ | —■ |
| Seynaeve 2021 [A] | 31 | 0 | 14 | 50 | 0.69 [0.53, 0.82] | 1.00 [0.93, 1.00] | —■ | —■ |



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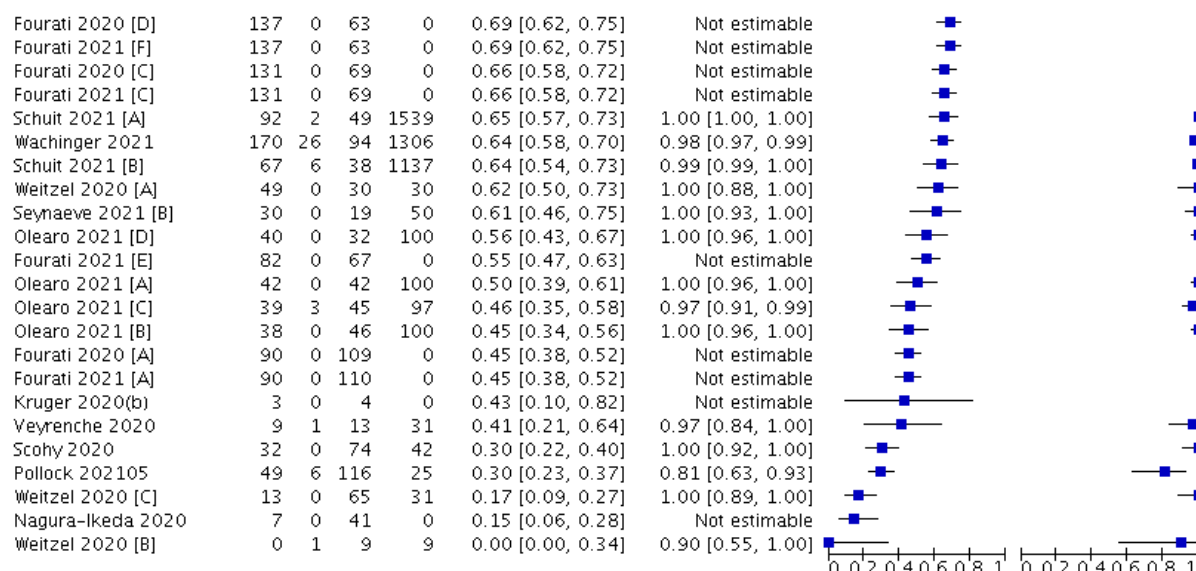


Figure 4. Sensitivity and specificity of RAgT in early testing (0-7 days after symptom onset)

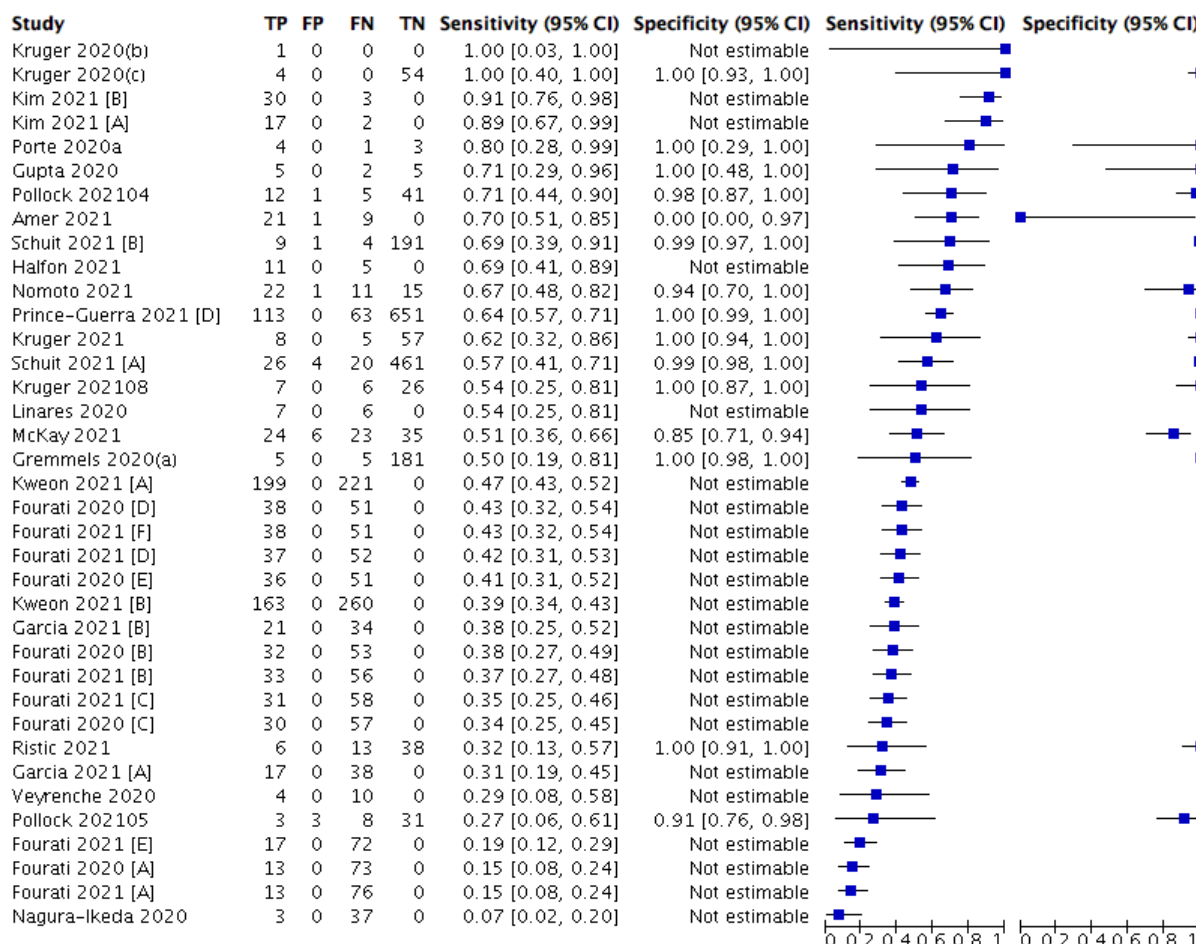


Figure 5. Sensitivity and specificity of RAgT in late testing (> 7 days)



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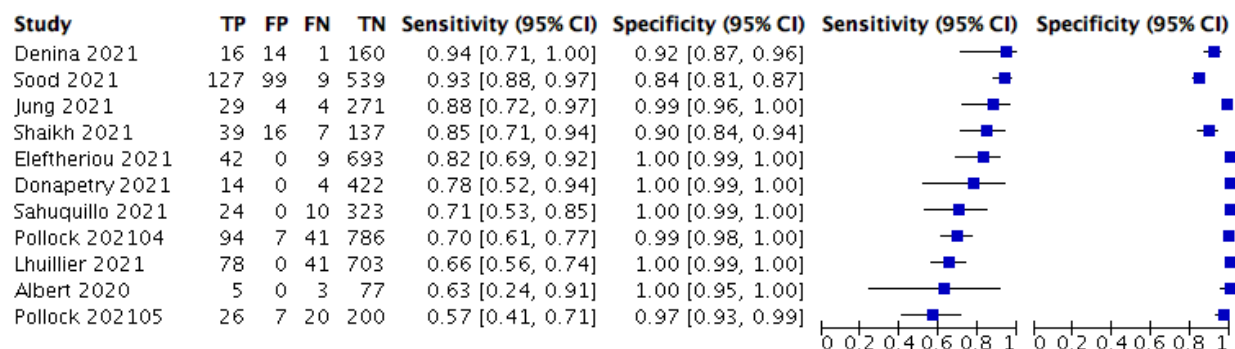


Figure 6. Sensitivity and specificity of RAgT in special population - children

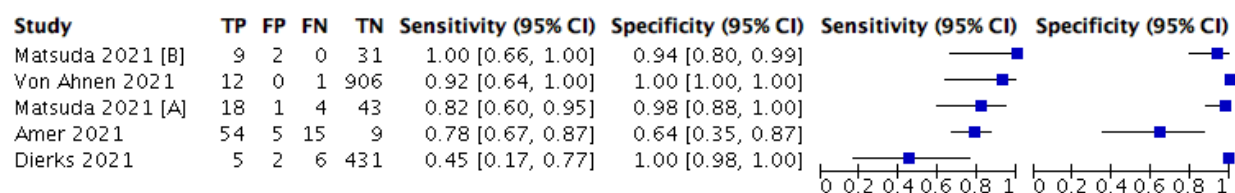


Figure 7. Sensitivity and specificity of RAgT in special population - healthcare workers

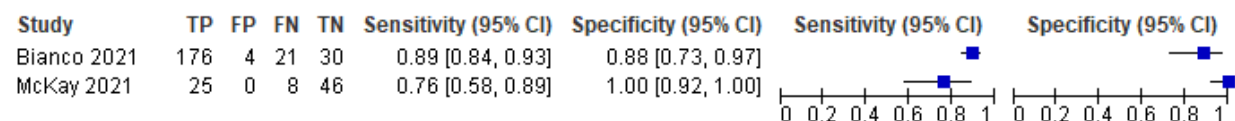


Figure 8. Sensitivity and specificity of RAgT use in symptomatic individuals in an outbreak settings

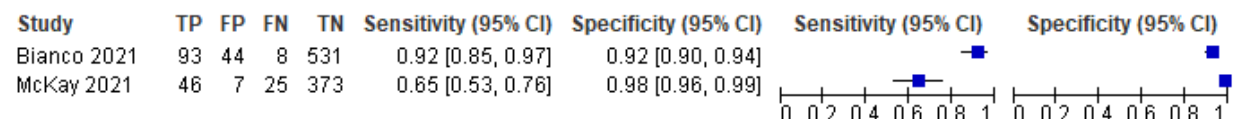


Figure 9. Sensitivity and specificity of RAgT use in asymptomatic individuals in an outbreak settings



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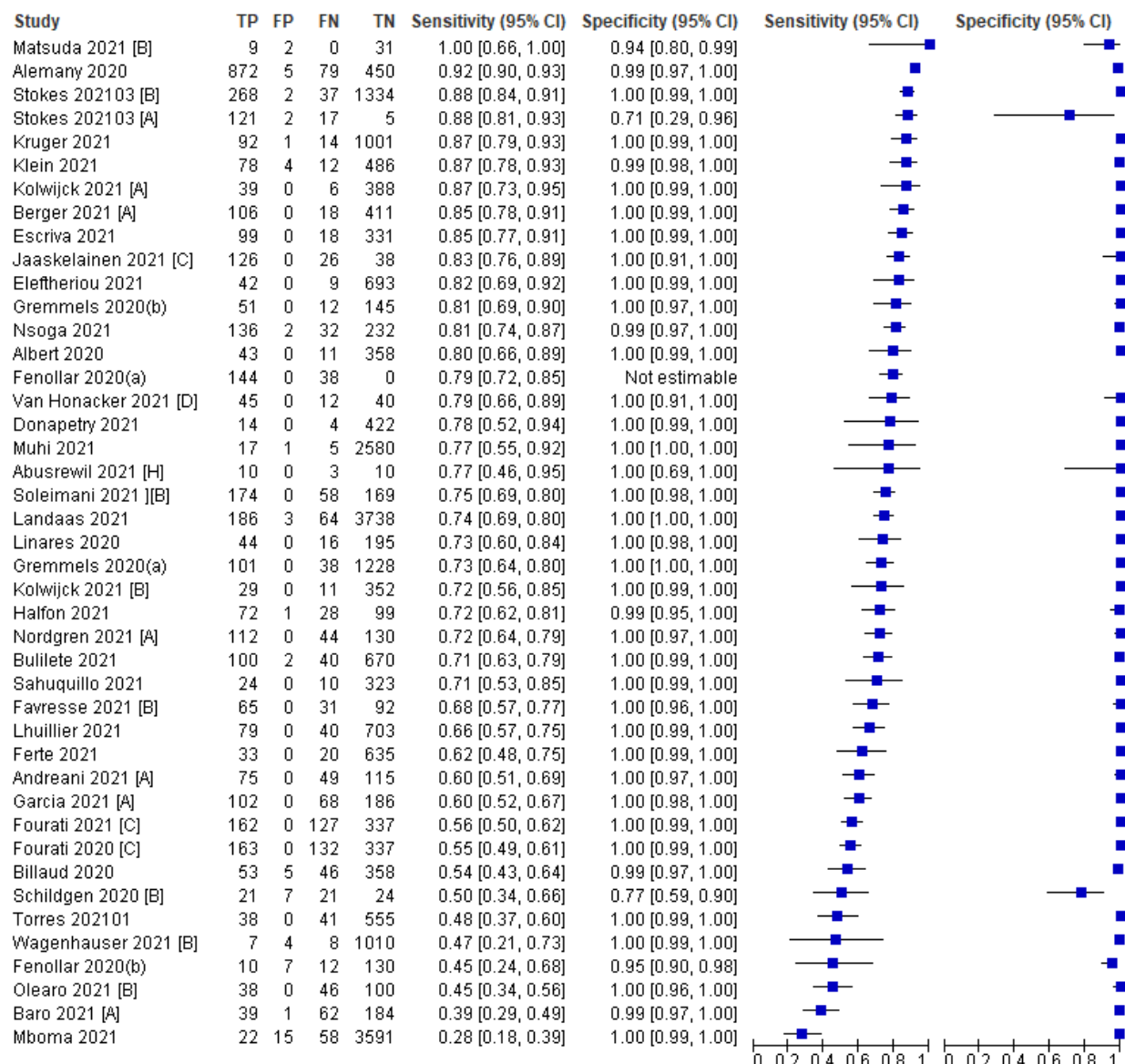


Figure 10. Sensitivity and specificity of Panbio™ Ag-RDT (Abbott)



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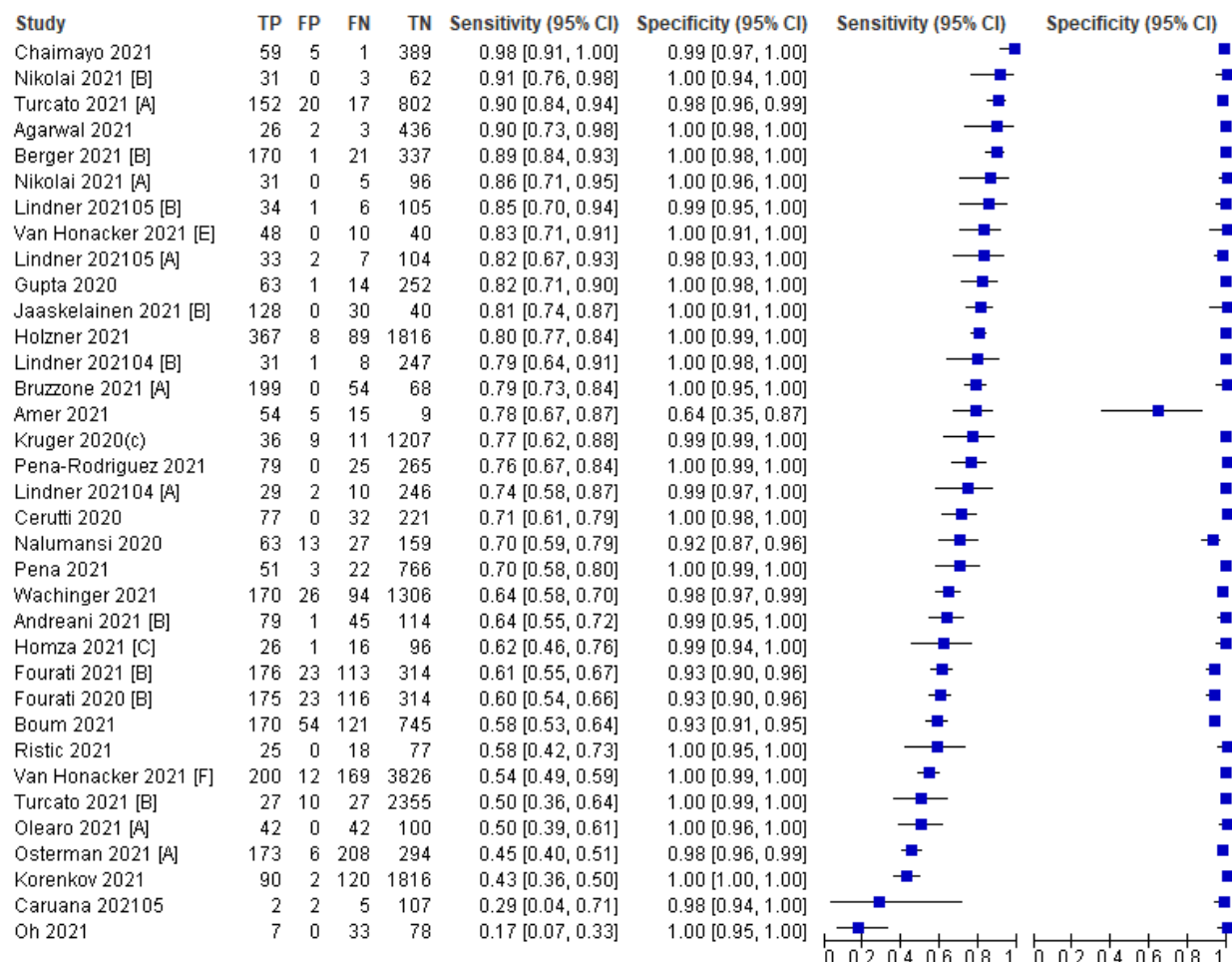


Figure 11. Sensitivity and specificity of Standard Q COVID-19 Ag Test (SD Biosensor)



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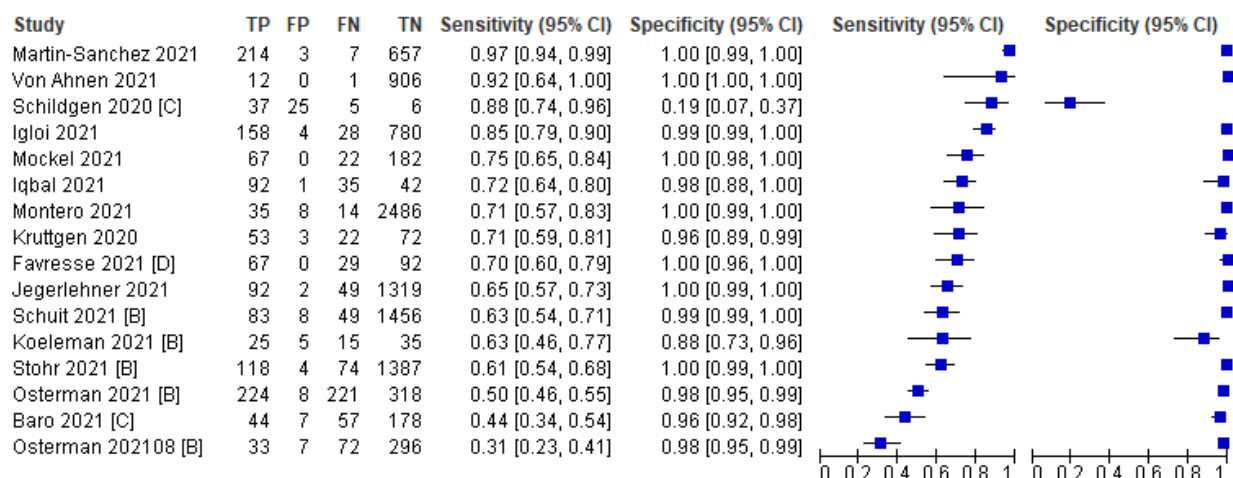


Figure 12. Sensitivity and specificity of Roche SARS-CoV-2 Rapid Antigen Test (Roche Diagnostics)

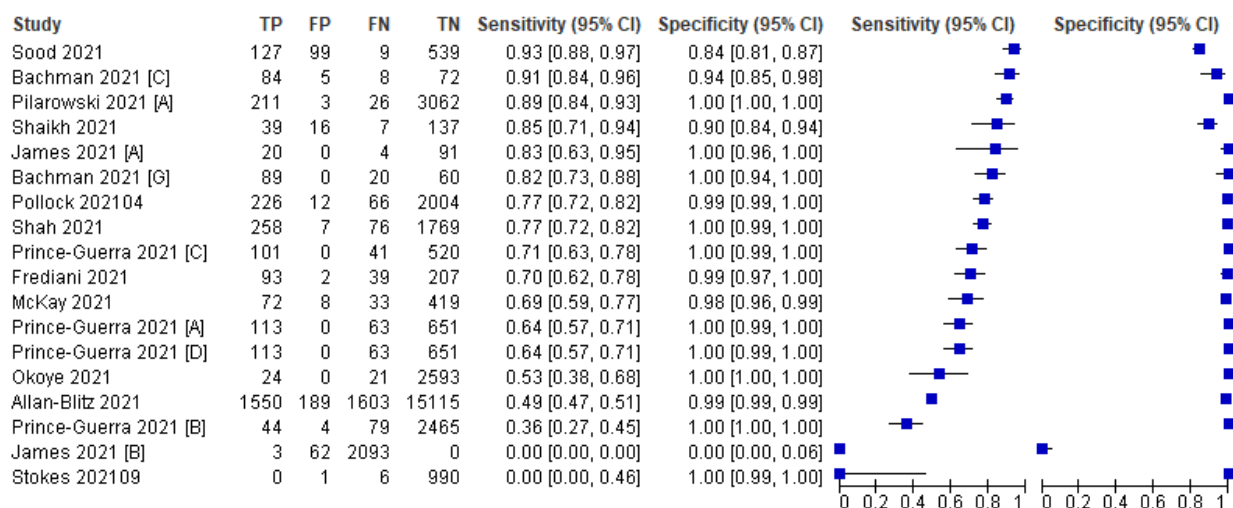


Figure 13. Sensitivity and specificity of Binax NOW SARS-CoV-2 (Abbott)

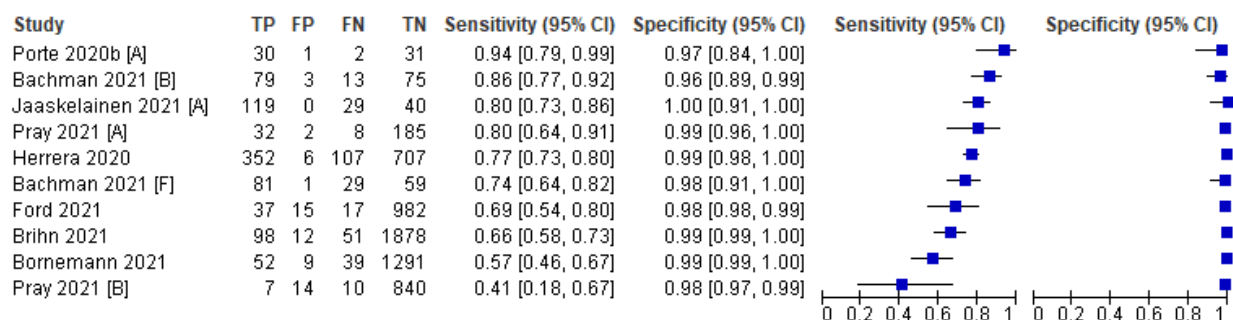


Figure 14. Sensitivity and specificity of Sofia SARS Antigen Fluorescent Immunoassay (Quidel)



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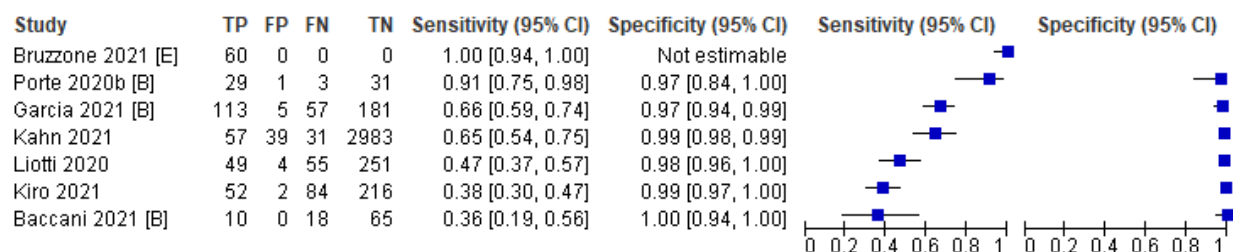


Figure 15. Sensitivity and specificity of Standard F Covid19 Ag FIA (SD Biosensor)

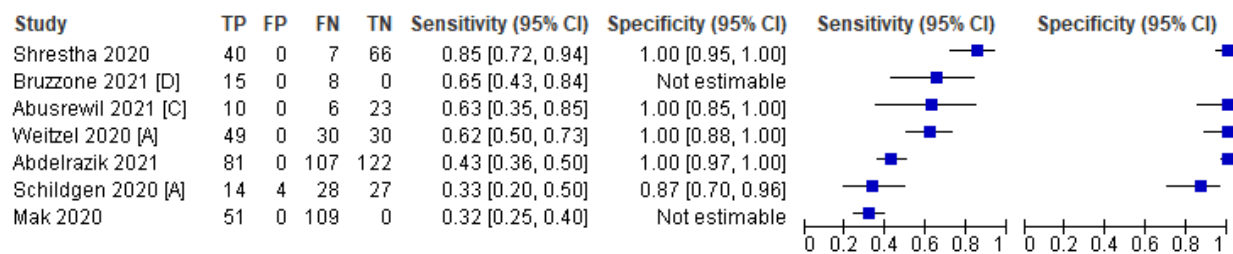


Figure 16. Sensitivity and specificity of Biocredit COVID-19 Ag test (RapiGEN)

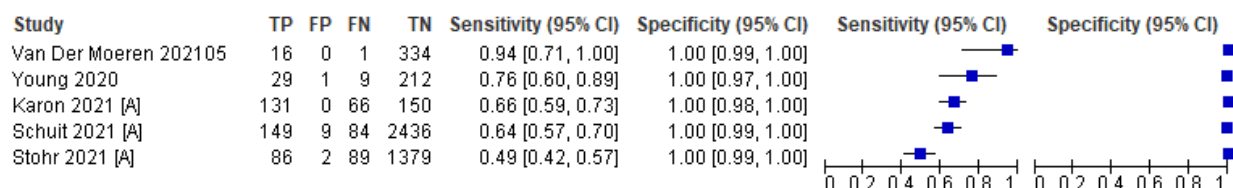


Figure 17. Sensitivity and specificity of COVID-19 Rapid Antigen Test (BD Veritor)

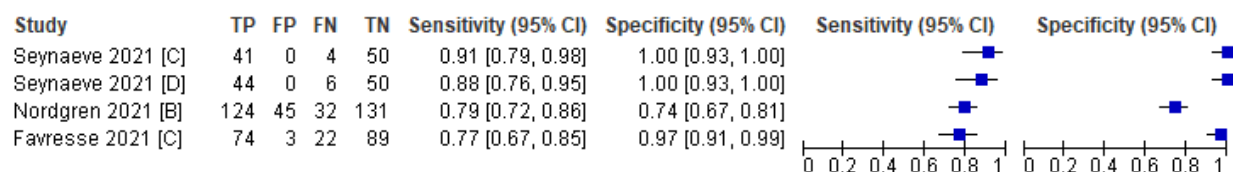


Figure 18. Sensitivity and specificity of Coronavirus Ag Rapid Test Cassette (Healgen Scientific)

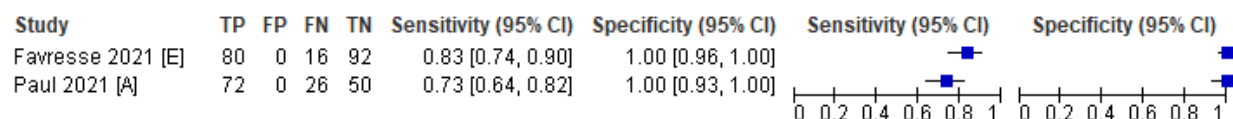


Figure 19. Sensitivity and specificity of VITROS Immunodiagnostic Products SARS-CoV-2 Antigen test (Ortho Clinical Diagnostics)

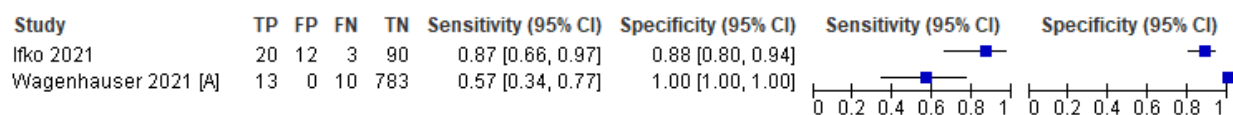


Figure 20. Sensitivity and specificity of NADAL COVID-19 Rapid Antigen Test (Nal Von Minden)



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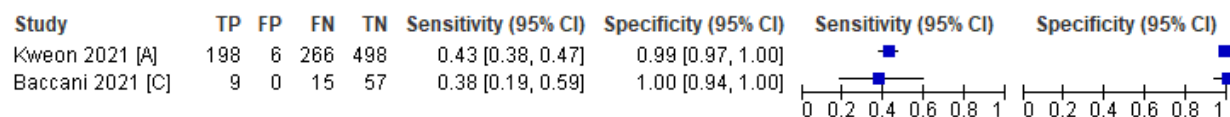


Figure 21. Sensitivity and specificity of AFIAS COVID-19 Ag (AFC) (Menarini)

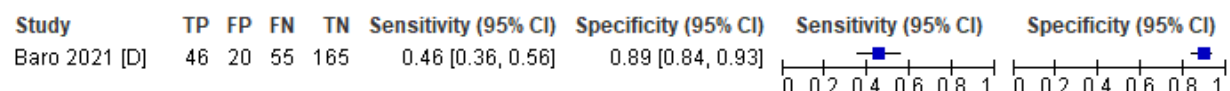


Figure 22. Sensitivity and specificity of SARS-CoV-2 Antigen Rapid Test Kit (Lepu Medical)

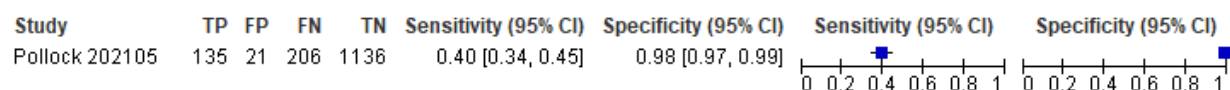


Figure 23. Sensitivity and specificity of CareStart COVID-19 Antigen Test (Access Bio)

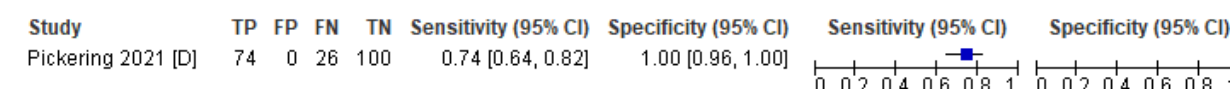


Figure 24. Sensitivity and specificity of Encode SARS-CoV-2 Antigen Rapid Test Device (Zhuhai Encode Medical Engineering)

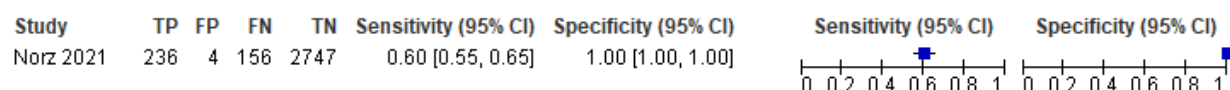


Figure 25. Sensitivity and specificity of Elecsys SARS-CoV-2 Antigen assay (Roche)



Figure 26. Sensitivity and specificity of SIENNA COVID-19 Antigen Rapid Test Cassette

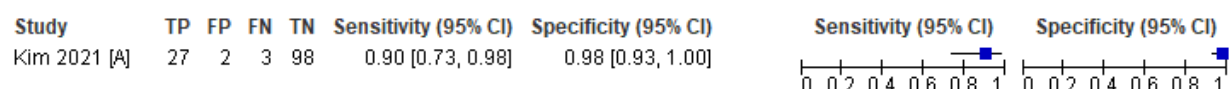


Figure 27. Sensitivity and specificity of GenBody COVAG025 Rapid Antigen Test

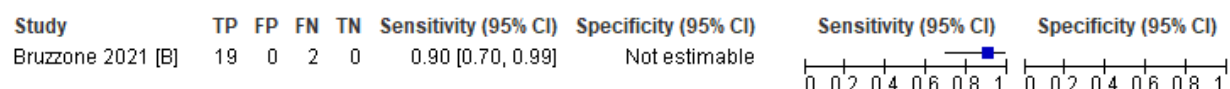


Figure 28. Sensitivity and specificity of Humasis COVID-Ag Test COVID-19 Antigen (Humasis)



Figure 29. Sensitivity and specificity of NowCheck COVID-19 Ag Test (Bionote)

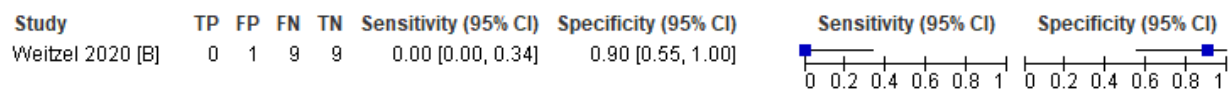


Figure 30. Sensitivity and specificity of StrongStep Rapid Antigen Test (Liming)



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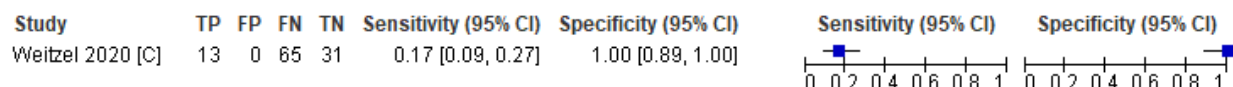


Figure 31. Sensitivity and specificity of Huaketai SARS-CoV-2 Rapid Antigen Test (Savant)



Figure 32. Sensitivity and specificity of SARS-CoV-2 Antigen Rapid Test Kit (JOYSBIO)

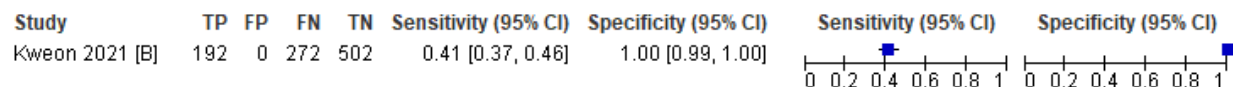


Figure 33. Sensitivity and specificity of ichroma™ COVID-19 Ag (Boditech)



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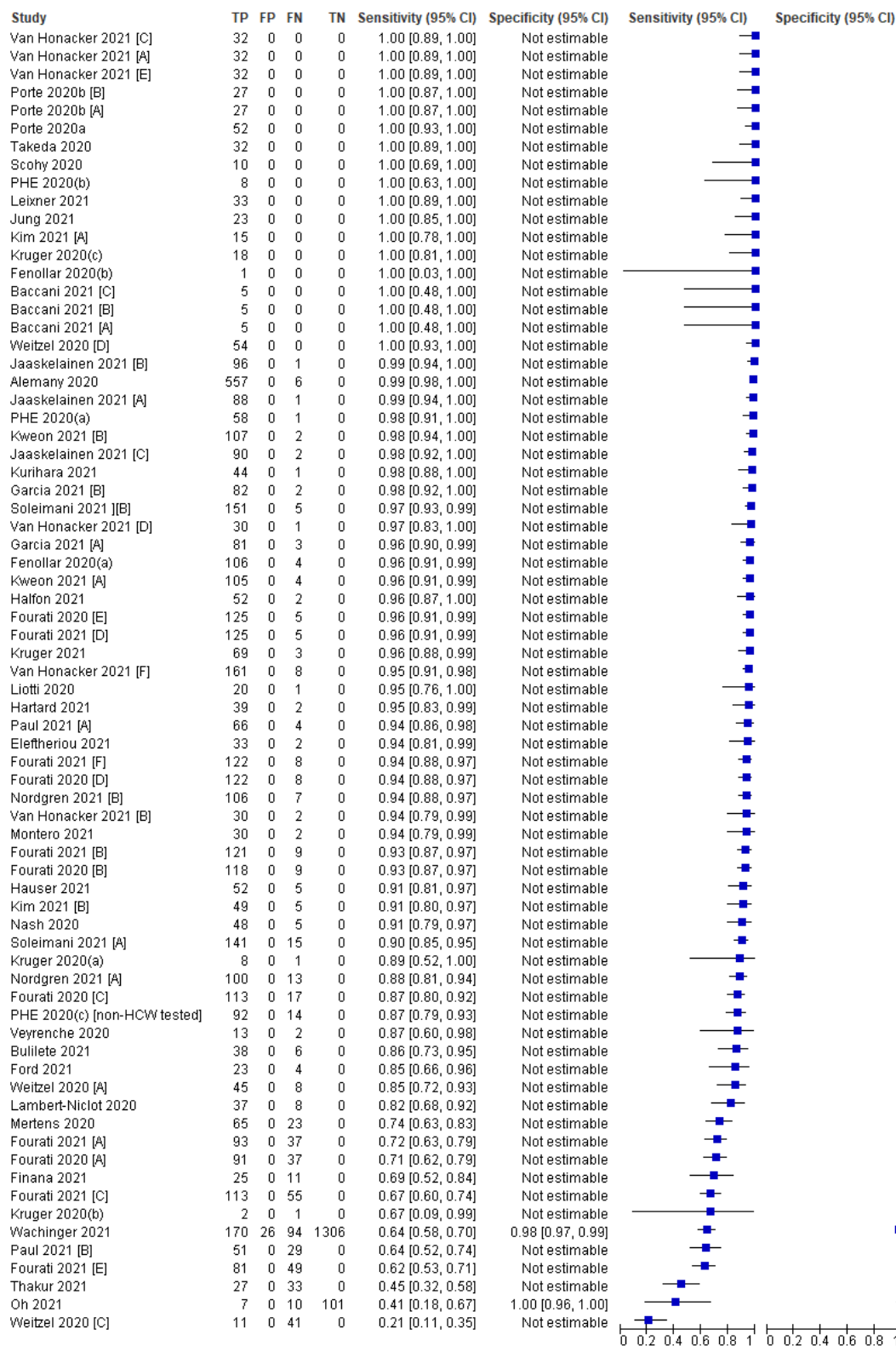


Figure 34. Sensitivity and specificity of RAgt if Ct value < 25



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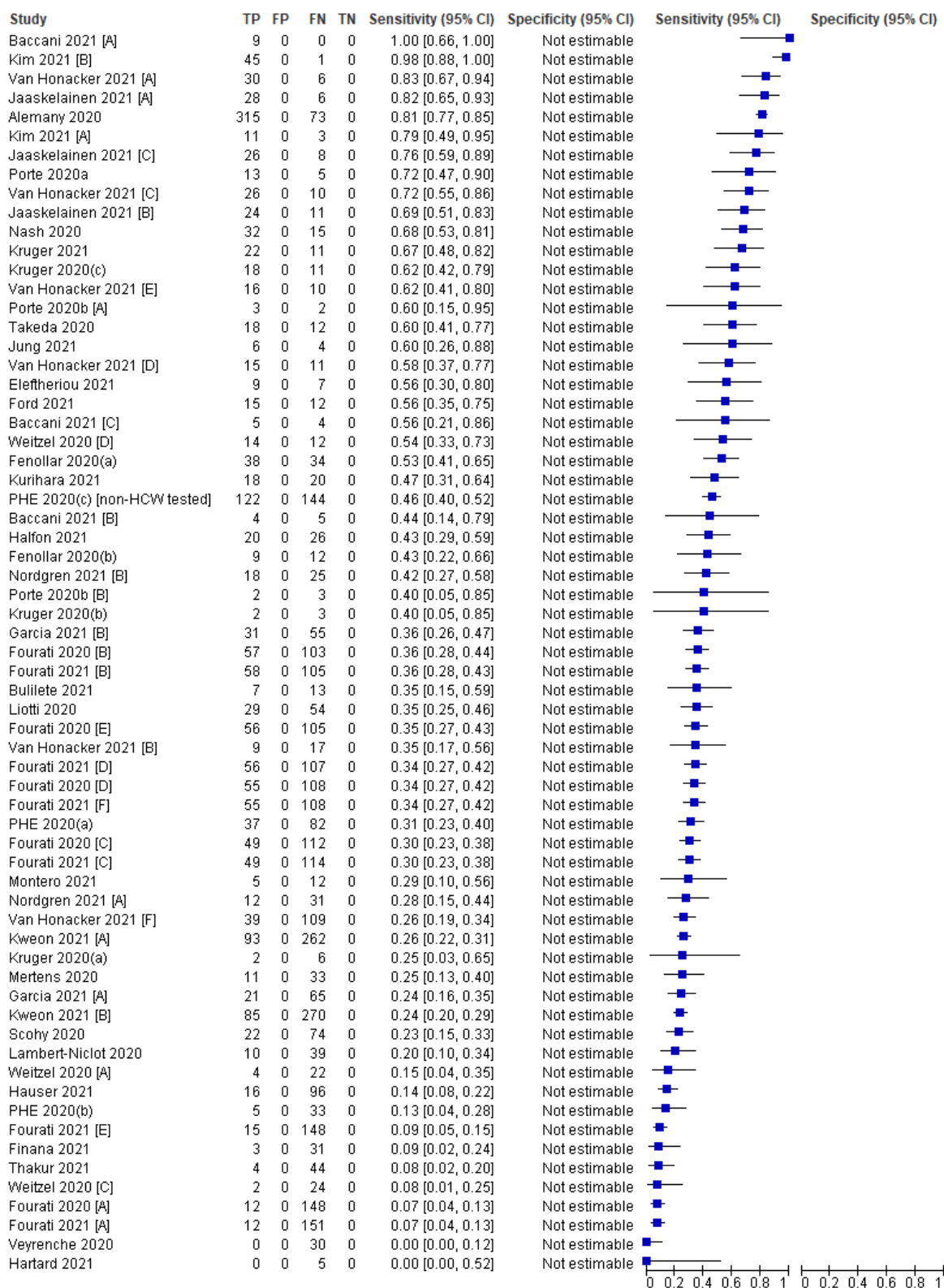


Figure 35. Sensitivity and specificity of RAgt if Ct value > 25



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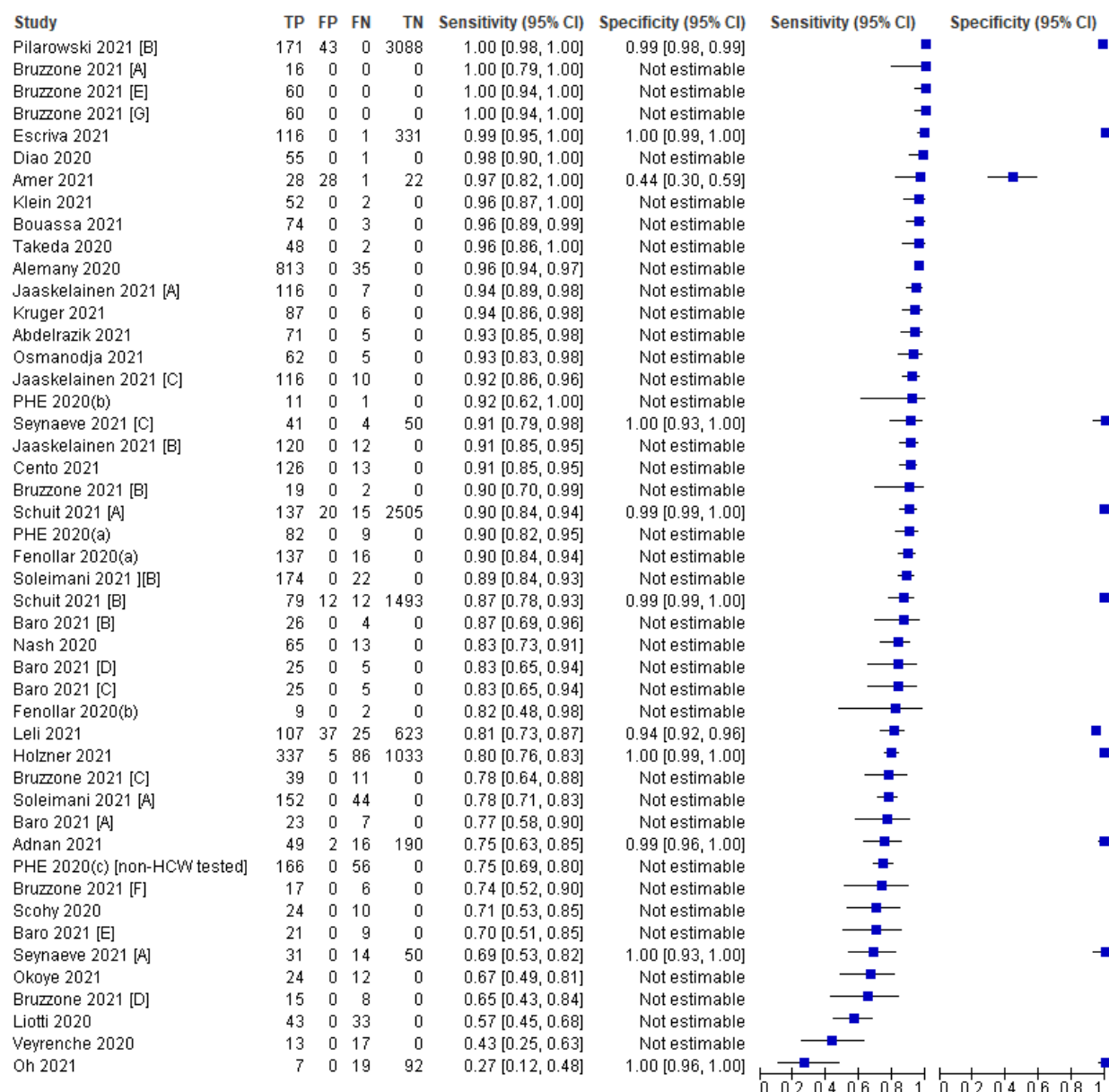


Figure 36. Sensitivity and specificity of RAgt if other Ct thresholds for 'higher' viral load



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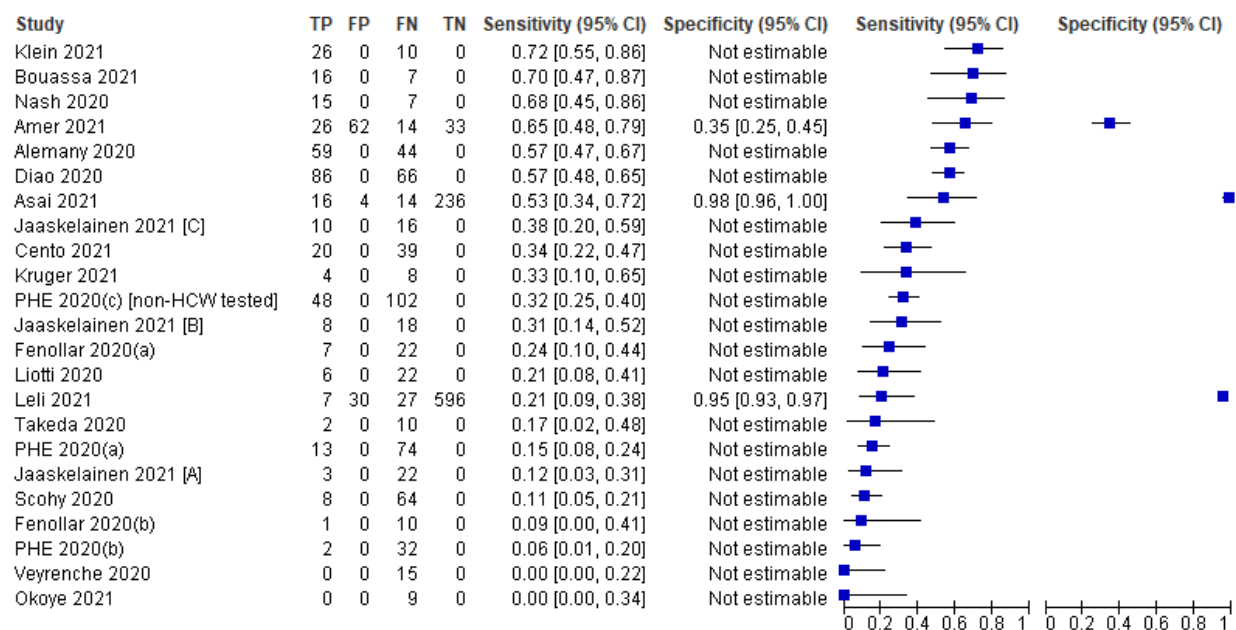


Figure 37. Sensitivity and specificity of RAgt if other Ct thresholds for 'lower' viral load



Philippine COVID-19 Living Clinical Practice Guidelines

Saliva

| Study | TP | FP | FN | TN | Sensitivity (95% CI) | Specificity (95% CI) |
|--------------------|-----|----|----|------|----------------------|----------------------|
| Ishii 2021 [D] | 8 | 1 | 0 | 123 | 1.00 [0.63, 1.00] | 0.99 [0.96, 1.00] |
| Kobayashi 2021 [B] | 111 | 3 | 3 | 5272 | 0.97 [0.93, 0.99] | 1.00 [1.00, 1.00] |
| Asai 2021 | 49 | 4 | 14 | 238 | 0.78 [0.66, 0.87] | 0.98 [0.96, 1.00] |
| Seitz 2021 | 8 | 0 | 10 | 22 | 0.44 [0.22, 0.69] | 1.00 [0.85, 1.00] |
| Mak 2020 | 18 | 0 | 27 | 0 | 0.40 [0.26, 0.56] | Not estimable |
| Ishii 2021 [B] | 3 | 0 | 6 | 84 | 0.33 [0.07, 0.70] | 1.00 [0.96, 1.00] |
| Nagura-Ikeda 2020 | 12 | 0 | 91 | 0 | 0.12 [0.06, 0.19] | Not estimable |
| Uwamino 2021 [B] | 4 | 0 | 37 | 0 | 0.10 [0.03, 0.23] | Not estimable |

Sputum

| Study | TP | FP | FN | TN | Sensitivity (95% CI) | Specificity (95% CI) |
|----------|----|----|----|----|----------------------|----------------------|
| Mak 2020 | 5 | 0 | 40 | 0 | 0.11 [0.04, 0.24] | Not estimable |

NP + saliva

| Study | TP | FP | FN | TN | Sensitivity (95% CI) | Specificity (95% CI) |
|-------------|----|----|----|-----|----------------------|----------------------|
| Yokota 2021 | 31 | 2 | 3 | 307 | 0.91 [0.76, 0.98] | 0.99 [0.98, 1.00] |

Exhaled breath

| Study | TP | FP | FN | TN | Sensitivity (95% CI) | Specificity (95% CI) |
|-----------------|----|----|----|----|----------------------|----------------------|
| Maniscalco 2021 | 12 | 1 | 1 | 91 | 0.92 [0.64, 1.00] | 0.99 [0.94, 1.00] |

OP

| Study | TP | FP | FN | TN | Sensitivity (95% CI) | Specificity (95% CI) |
|----------------------|-----|----|----|------|----------------------|----------------------|
| Nsoga 2021 | 136 | 2 | 32 | 232 | 0.81 [0.74, 0.87] | 0.99 [0.97, 1.00] |
| Kahn 2021 | 57 | 39 | 31 | 2983 | 0.65 [0.54, 0.75] | 0.99 [0.98, 0.99] |
| Wagenhauser 2021 [A] | 13 | 0 | 10 | 783 | 0.57 [0.34, 0.77] | 1.00 [1.00, 1.00] |
| Wagenhauser 2021 [B] | 7 | 4 | 8 | 1010 | 0.47 [0.21, 0.73] | 1.00 [0.99, 1.00] |
| Wagenhauser 2021 [C] | 23 | 12 | 40 | 3146 | 0.37 [0.25, 0.50] | 1.00 [0.99, 1.00] |

Nasal + OP

| Study | TP | FP | FN | TN | Sensitivity (95% CI) | Specificity (95% CI) |
|------------------|------|-----|------|-------|----------------------|----------------------|
| Allan-Blitz 2021 | 1550 | 189 | 1603 | 15115 | 0.49 [0.47, 0.51] | 0.99 [0.99, 0.99] |

Nasal (anterior nares + mid turbinate)

| Study | TP | FP | FN | TN | Sensitivity (95% CI) | Specificity (95% CI) |
|------------------------|-----|----|-----|-------|----------------------|----------------------|
| Denina 2021 | 16 | 14 | 1 | 160 | 0.94 [0.71, 1.00] | 0.92 [0.87, 0.96] |
| Sood 2021 | 127 | 99 | 9 | 539 | 0.93 [0.88, 0.97] | 0.84 [0.81, 0.87] |
| Bachman 2021 [C] | 84 | 5 | 8 | 72 | 0.91 [0.84, 0.96] | 0.94 [0.85, 0.98] |
| Bachman 2021 [D] | 84 | 4 | 8 | 78 | 0.91 [0.84, 0.96] | 0.95 [0.88, 0.99] |
| Nikolai 2021 [B] | 31 | 0 | 3 | 62 | 0.91 [0.76, 0.98] | 1.00 [0.94, 1.00] |
| Bianco 2021 | 269 | 48 | 29 | 561 | 0.90 [0.86, 0.93] | 0.92 [0.90, 0.94] |
| Pilarowski 2021 [A] | 211 | 3 | 26 | 3062 | 0.89 [0.84, 0.93] | 1.00 [1.00, 1.00] |
| Osmanodja 2021 | 62 | 1 | 8 | 308 | 0.89 [0.79, 0.95] | 1.00 [0.98, 1.00] |
| Nikolai 2021 [A] | 31 | 0 | 5 | 96 | 0.86 [0.71, 0.95] | 1.00 [0.96, 1.00] |
| Bachman 2021 [B] | 79 | 3 | 13 | 75 | 0.86 [0.77, 0.92] | 0.96 [0.89, 0.99] |
| Chiu 2021 [A] | 64 | 14 | 11 | 260 | 0.85 [0.75, 0.92] | 0.95 [0.92, 0.97] |
| Shaikh 2021 | 39 | 16 | 7 | 137 | 0.85 [0.71, 0.94] | 0.90 [0.84, 0.94] |
| Klein 2021 | 38 | 2 | 7 | 243 | 0.84 [0.71, 0.94] | 0.99 [0.97, 1.00] |
| Chiu 2021 [C] | 32 | 18 | 6 | 22938 | 0.84 [0.69, 0.94] | 1.00 [1.00, 1.00] |
| Bachman 2021 [A] | 75 | 2 | 15 | 77 | 0.83 [0.74, 0.90] | 0.97 [0.91, 1.00] |
| Chiu 2021 [B] | 62 | 10 | 13 | 264 | 0.83 [0.72, 0.90] | 0.96 [0.93, 0.98] |
| Lindner 202105 [A] | 33 | 2 | 7 | 104 | 0.82 [0.67, 0.93] | 0.98 [0.93, 1.00] |
| Kruger 202108 | 120 | 4 | 26 | 611 | 0.82 [0.75, 0.88] | 0.99 [0.98, 1.00] |
| Bachman 2021 [G] | 89 | 0 | 20 | 60 | 0.82 [0.73, 0.88] | 1.00 [0.94, 1.00] |
| Pray 2021 [A] | 32 | 2 | 8 | 185 | 0.80 [0.64, 0.91] | 0.99 [0.96, 1.00] |
| Bachman 2021 [H] | 87 | 1 | 23 | 59 | 0.79 [0.70, 0.86] | 0.98 [0.91, 1.00] |
| Pollock 202104 | 226 | 12 | 66 | 2004 | 0.77 [0.72, 0.82] | 0.99 [0.99, 1.00] |
| Muhi 2021 | 17 | 1 | 5 | 2580 | 0.77 [0.55, 0.92] | 1.00 [1.00, 1.00] |
| Shah 2021 | 258 | 7 | 76 | 1769 | 0.77 [0.72, 0.82] | 1.00 [0.99, 1.00] |
| Lindner 202104 [A] | 29 | 2 | 10 | 246 | 0.74 [0.58, 0.87] | 0.99 [0.97, 1.00] |
| Bachman 2021 [F] | 81 | 1 | 29 | 59 | 0.74 [0.64, 0.82] | 0.98 [0.91, 1.00] |
| Prince-Guerra 2021 [C] | 101 | 0 | 41 | 520 | 0.71 [0.63, 0.78] | 1.00 [0.99, 1.00] |
| Frediani 2021 | 93 | 2 | 39 | 207 | 0.70 [0.62, 0.78] | 0.99 [0.97, 1.00] |
| Bachman 2021 [E] | 75 | 2 | 33 | 57 | 0.69 [0.60, 0.78] | 0.97 [0.88, 1.00] |
| Leli 2021 | 114 | 30 | 52 | 596 | 0.69 [0.61, 0.76] | 0.95 [0.93, 0.97] |
| Ford 2021 | 37 | 15 | 17 | 982 | 0.69 [0.54, 0.80] | 0.98 [0.98, 0.99] |
| McKay 2021 | 2 | 0 | 1 | 134 | 0.67 [0.09, 0.99] | 1.00 [0.97, 1.00] |
| Brhnn 2021 | 98 | 12 | 51 | 1878 | 0.66 [0.58, 0.73] | 0.99 [0.99, 1.00] |
| Prince-Guerra 2021 [A] | 113 | 0 | 63 | 651 | 0.64 [0.57, 0.71] | 1.00 [0.99, 1.00] |
| Prince-Guerra 2021 [D] | 113 | 0 | 63 | 651 | 0.64 [0.57, 0.71] | 1.00 [0.99, 1.00] |
| Pray 2021 [B] | 7 | 14 | 10 | 840 | 0.41 [0.18, 0.67] | 0.98 [0.97, 0.99] |
| Pollock 202105 | 135 | 21 | 206 | 1136 | 0.40 [0.34, 0.45] | 0.98 [0.97, 0.99] |
| Prince-Guerra 2021 [B] | 44 | 4 | 79 | 2465 | 0.36 [0.27, 0.45] | 1.00 [1.00, 1.00] |

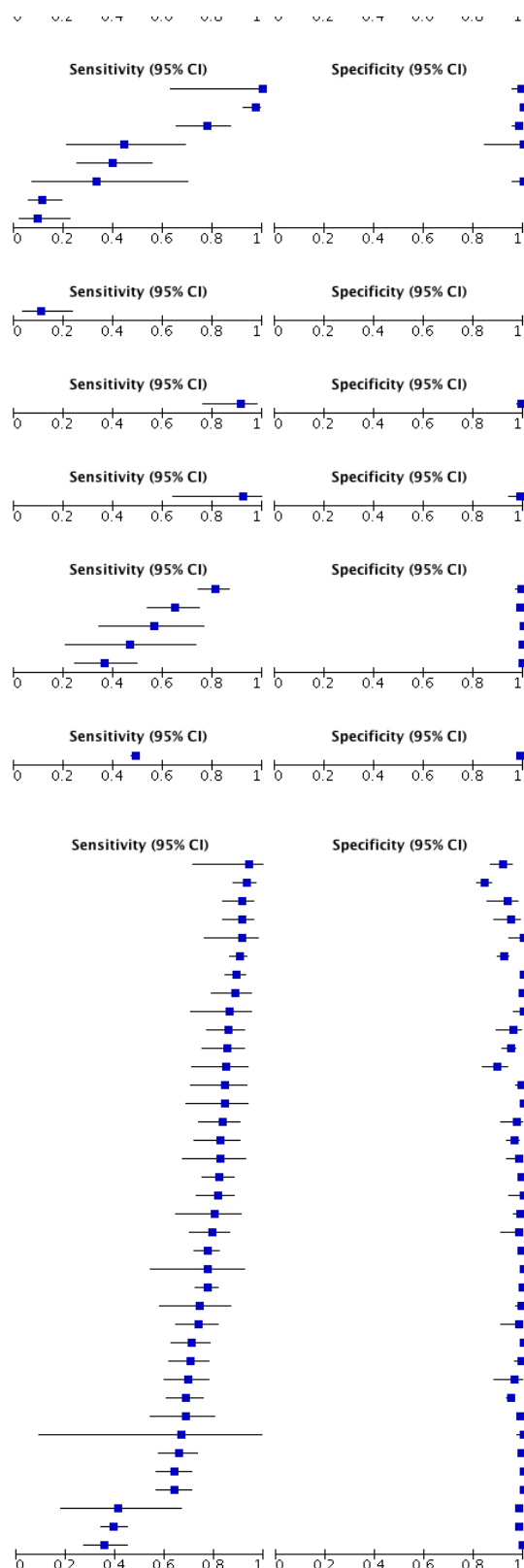


Figure 38. Sensitivity and specificity of RAgT per specimen type